## UNIVERSITY OF NATAL

# A CONTRIBUTION TO KNOWLEDGE OF THE GENUS HYPOXIS L. (HYPOXIDACEAE) IN NATAL, SOUTH AFRICA. 

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## FRONTISPIECE



Hypoxis lat1folia Hook.
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## INTRODUCTION


#### Abstract

The genus Hypoxis was established by Linnaeus in 1759. It was only a brief description, "Corolla 6 parts, stamens short, ovary Inferior." From this unimposing beginning Hypoxis has become known to be pantropical in distribution, and a well known geophyte from the Cape Peninsula to Ethiopia, but especially in South Africa.

Baker (1878 \& 1896) and Ne1 (I914) are the only authors to have revised the genus in southern Africa and indeed to have made any substantial contributions to knowledge of the species. However, both authors, but particularily Baker, were limited by the number of specimens avallable for study and the amount of field work that could be undertaken, if any at ald. Thus concepts of the genus were not complete and no effective key to identification was produced. Nel extended Baker's work but did not attempt to improve upon this author's species descriptions. This, in itself, is a short-coming in Nel's revision because, although he cited more specimens than did Baker, he did not revise the diagnostic characters by which species were recognized. Not surprisingly, the worker attempting to use Nel's monograph for identification of species, often becomes confused and hesitant to be definite about the application of names.

Nel covered all African species. Within this continental framework he recognized eleven groups, six of which were almost exclusively tropical except that in some instances, one southern African species was included.


Because of its greater conciseness and more direct relevance to the flora, it i.s Baker's work (1896), and especially his key, which

1s the more widely used by South African botanists. However, as Nel suggested, this work is somewhat unreliable because of the variable nature of the characters on which diagnoses are based.

This thesis has attempted to clarify knowledge of the genus in Natal, and to construct a more effective key to species identification. However as plants of Hypoxis are not restricted to political limits, the boundaries of Natal were not considered in studying species represented within this province: Choice of Natal species as a group for detailed consideration was for two reasons; firstly field work was practical in this area, and secondly, as the genus is not easily subdivided into groups of similar species, some criterion had to be used to decrease the number of species involved.

Thus twenty-one species of Hypoxis were worked on, following their distribution throughout South Africa. Obviously when revisions of the Cape and Transvaal species are undertaken amendments may have to be made to the Natal section. It is hoped that similar revisions for other areas will follow, and that in the foreseeable future there will be an amalgamated revision of the genus in southern Africa, which will include South West Africa, Swaziland, Lesotho and South Africa.

## CHAPTER <br> I

THE GENUS

### 1.1 History

The genus Hypoxis was established by Linnaeus in 1759. Of the three specimens cited by this author only two now belong in this genus, and they are the American species H. erecta L. and H. decumbens $L$. The third is a member of the genus Colchicum L. (C. montanum L.), of the family Liliaceae.

Since its establishment, Hypoxis has been placed in the families Amaryllidaceae (incIuding subfamily Hypoxidoideae and tribes Hypoxideae and Haemodoreae) and Hypoxidaceae by different authors. Geerinck (1969) in the introduction to his work on the genera of Haemodoraceae and Hypoxidaceae, summarized these placements without discussing them. He maintained Hypoxis in the family Hypoxidaceae, implying that this was the most satisfactory position for the genus. This relationship is accepted in this study of the Natal species.

Descriptions of the genus were compiled by Roemer $\&$ Schultes (1830), Herbert (1837), Endlicher (1840), and Salisbury (1866). The last named author grouped the genera Spiloxene and Ianthe with Hypoxis, placing them under Section 2 of the order Hypoxideae. There was no key to the genera, but he listed their main characters (see Table l). Salisbury did not include species in his work.

Baker in 1878 was the first author to prepare a synopsis of the genus Hypoxis and to construct a key to its world taxa. In his introduction he stated:


Table I. Sumary of characters distinguishing Spiloxene and Ianthe from Hypoxis (Salisbury, 1866)
"Another group of petaloid monocotyledons of which a synopsis is greatly needed for daily use is Hypoxidaceae. The latest that is practically usable is that of Roemer and Schultes, published in the second part of the 7th volume of their "Systema Vegetabilium" in 1830. This, for the date, is a full sumary of what was known and had been written about these plants; but of a large proportion of the species the authors had no opportunity of examining specimens, and could therefore only cite the descriptions of other writers, without being able to reduce them to one common formula. An author writing under such circumstances is both sure to admit too many species and to fail to appreciate their true relationship."

Baker recognized 4 genera within Hypoxidaceae: Hypoxis, Molineria, Curculigo and Pauridia, and between 60-70 species. He distinguished the genera thus (translation from the Latin):

Perianth tube above ovary lacking or very short. Stamens epigynous.

1. Hypoxis. Fruit capsulate, circumscissile, operculate. Leaves sessile not plicate. C.B. Spei, Afric. trop., Asia australis, America.
2. Molineria. Fruits berry-1ike. Leaves petiolate, plicate. Asia trop., Australia bor., Ins. Seychell?

Perianth tube extended above ovary. Stamens perigynous.
3. Curculigo. Tube elongated, filiform. Stamens 6. Reg. trop. "utriusque orbis", C.B. Spei.
4. Pauridia. Tube short, funnel-shaped. Stamens 3.

In his revision Baker (1878) subdivided the genus Hypoxis into two subgenera, Ianthe (Ianthe et Spiloxene, Salisb.) and Euhypoxis, (Rypoxis, Salisb.).

Ianthe: Plants totally glabrous. (Corm always small, annual. Anthers linear, basifixed. Stigma $\pm$ discrete).
Euhypoxis: Plants $\pm$ hairy. (Ovary nearly always clothed with setose, erect outspreading dense hairs. Leaves nearly always pilose. Tuber small or large. Anthers nearly always slightly versatile, base sagittate. Stigma nearly always concrete).

In 1896 in Flora Capensis, Baker revised the South African species of Hypoxidaceae. He formulated a more comprehensive, but nevertheless not easily usable, key to the species employing such characters as corm size (an unreliable parameter in perennial plants); flower size; form, nature and hairiness of the leaves; and inflorescence type. Nel commented later: "Such a key is virtually not usable."

Baker's use of "perianth tube" in the generic descriptions was regarded by Nel (1914) as "false terminology" and this was emended by both Bentham \& Hooker (1883) and Nel to "rostrum" (beak). It is the extended sterile apex of the inferior ovary.

The most recent monograph of the African genera of Hypoxidaceae is that of Nel (1914). In this he recognized six genera: Hypoxis (Baker's subgenus Euhypoxis), Ianthe (Baker's subgenus Ianthe), Curculigo, Forbesia, Molineria and Rhodohypoxis (Nel, new genus).

Nel was of the opinion that previous authors had attached too great an importance to the nature of the fruit in classification of the genera, and thus species which had nothing in comon except the same fruit, were found together in one genus. Forbesia and Curculigo could be separated from Hypoxis and Molineria by the presence of a beak on the fruit, but within the genus Rhodohypoxis several species are beaked (for example R. rubella (Baker) Nel) and others (for example R. baurii (Baker) Nel) are not. This is confirmed by Hilliard \& Burtt (1973) who stated: "Thus we find an ovary beak occurs in several groups of monocotyledons, while at the same time it is not always constant in a genus, even in such a small and distinctive one as Rhodohypoxis."

Ne1 referred to characters used to differentiate genera by previous authors as "by no means constant," and "ambiguous." He distinguished his genera primarily on the form of attachment of the anthers to the filaments.

Molineria, Forbesia, Ianthe: anthers adnate, connate with the base of the filaments (translation from the Latin):

Rhodohypoxis, Curculigo, Hypoxis: anthers basifixed.
Curculigo was distinguished from Hypoxis by the presence of a thread-like beak and berry-like fruit, whereas the latter genus has perianth segnents free to the base while the fruit is a dehiscent capsule.

Rhodohypoxis is the closest genus to Hypoxis (Baker's species H. baurii and H. milloides (Baker) Hilliard \& Burtt, were reinstated under Rhodohypoxis). Rhodehypoxis was separated from Hypoxis by Nel by virtue of its red or white flowers, the presence of a perianth tube
and Its eplpetalous, sessile anthers which are different in shape from those of Hypoxis. On these characters Nel felt justified in establishing Rhodohypoxis as a new genus. Another characteristic used to distinguish these two genera was commented on by Hilliard \& Burtt (1973):

> "Rhodophypoxis shows marked generic individuality on account of the form of the flower which Milne-Redhead (in Bot. Mag. t 9412 , 1935) has very aptly described as having a "blind look": a phrase that sums up the way in which the inner tepals are inflexed in their lower part so that the mouth of the perianth is almost entirely closed."

Nel studied all the African species of Hypoxis and classified them into what he called natural groups, six, of the total eleven, of which almost exclusively comprised tropical species. Diagnostic characters used in dividing these groups were the form of the anther apex, the relationship of stigra to style, inflorescence type and leaf venation.

The main criticism of Nel's work is that he did not redescribe any of the species which had already been established, but merely cited more specimens, and thus his concept of these species was not always clear. The characters he used in species differentlation are discussed in detall under "diagnostic characters."

A number of new species of Hypoxis have been described since Nel's work, but no other major revision of the genus has been undertaken. At the present time, within the family Hypoxidaceae, revisionary work is being done on Rhodohypoxis (Hilliard \& Burtt; University of Natal, Pietermaritzburg and Edinburgh University, Scotland respectively) and on Empodium and Spiloxene (Thompson, University of Stellenbosch, Cape).


Table 2 : A summary of the genera established under the family Hypoxidaceae from 1878-1914.


Fig. 1. General known world distribution of Hypoxis Linn. (excluding Ianthe Nel) (sources: Baker (1878), Engler (1964), Geerinck (196.9, 1971), Index Kewensis, (1896-1970).

### 1.2 Geographical Distribution

The genus Hypoxis occurs throughout most of the warm temperate and tropical zones of the world (Fig. 1). It is difficult to determine the precise distributional limits, but the genus is not known to be present in Europe, northern and central Asia, north Africa, Canada and extra-tropical South America.

The largest number of species (53) is found in South Africa, with the greatest concentration in the Cape Province. Figures extracted from Index Kewensis (1895-1970) show that within the rest of Africa there are 46 species in Tropical Africa (including 2aire, East Africa, Angola, Malawi and Rhodesia), 3 in Ethiopia, 1 in Guinea, and 5 in the Cameroons. Australasia has 5 species, Asia 2, and there are 16 in the Americas (including Mexico). According to Baker (1878), "No known species extends its range from one continent to another."

Distribution and Habitat in southern Africa

Hypoxis species are found throughout southern Africa, except in the arid karroid regions of southern South West Africa and the north western Cape. . The majority of species that occur in Swaziland, Lesotho, the Transvaal, the Orange Free State, Natal and the northern Cape are perennial, grassveld species and would be classed as typical geophytes following Raunkiaer's (1937) classification of plant life forms. The rootstock, which is responsible for the plant bcing able to withstand harsh winters, droughts and annual veld burning, is a mucilagenous and resiniferous corm capable of continued growth throughout the duration of life of the plant. One of the genera first to produce aerial parts in spring, Hypoxis plants flower and are pollinated before other constituents of the vegetation, particularly the
grasses, reach a competetive height.
Some taxa show little altitudinal preference and the high veld, the mountains and the coast may support the same species. But there are exceptions: H. zululandensis S.E. Wood is known only at the coast; H. interjecta Nel and HI . neliana Schinz are not represented at low altitudes.

H: membranacea Bak. and H. parvola Bak., which are among the more fragile, less robust species in the genus, although sometimes occurring in open veld areas, prefer moister, ravine habitats and forest margins. Their corms are very small, (approximately 1 cm in diameter) and the leaves are membranous.

The Hypoxis species found south of Kougaberge, that is in the southern Cape, are distinct from those that occur in the remainder of southern Africa. They tend, on the whole, to be smaller, and are obviously adapted to winter rainfall conditions. Ianthe, a glabrous genus of Hypoxidaceae (separated from Hypoxis by Nel, 1914) is confined to this area.

### 1.3 Cytology

Very little work has been done on the cytology of Hypoxis. Wilsenach, Warren \& Papenfus (1967) compiled chromosome numbers for seven named species and one that was not identified. They also listed chromosome numbers for $H$. rooperi $S$. Moore that had been obtained by a previous group of workers (Fernandes \& Neves, 1961). The results
are sumarized in Table 3 , which also includes a chromosone number for H. pusilla Hook. (Darlingtọn \& Wylie, 1961).

## Chromosome Number

Species
H. stellepilis Ker
H. zeyheri Bak.

32
H. multiceps Buching. 36
H. longifolia Bak.
H. rooperi S. Moore (76-)96(-114)
H. rooperi S. Moore (76-)96(-114)
H. acuminata Bak.

Diploid
Haploid

16
H. Longifolia Bak. 72
$(38-) \pm 43(-58)$
$18(-20)$
H. filiformis Bak.

7
H. sp.
H. pusilla Hook.

Table 3. Chromosome numbers compiled for Hypoxis species so far investigated.

A basic chromosome number for Hypoxis has not been determined. The suggestion has been made that it is likely to be below 9, as at least one species has 16 somatic chromosomes.

For some at least of the species in which plants are large, chromosome numbers are higher than those recorded for species in which plants are small (for example for $\underline{H}$. rooperi $n=38- \pm 43-58$, while for H, filiformis $\Omega=7$ ), but no clearly defined polyploid series is evident within the genus at the present stage of knowledge.

Apomixis has not been confirmed for any species so far studied, but Wilsenach et al. (1967), regarded this phenomenon as likely because
of the degree of polymorphism, the variation in chromosome numbers, the abnormalities in meiosis of pollen mother cells in the high chromosome number species, the variation in chromosome numbers of pollen grains of the same species, the degeneration of megaspores, and the common occurrence of twin embryo-sacs, that they encountered.

A great deal more research is needed in this field. Because of complexities indicated by the preliminary work already undertaken, this is likely to prove a project in itself. A cytogenetic study was not attempted as part of this thesis. Clarification of the genetic history of Hypoxis may enable better understanding of patterns of hybridization and of variation within the genus than is possible at present. In conjunction with comparable studies within related genera, Ianthe Salisb., Curculigo Gaertn., Forbesia Eckl., Molineria Colla, Rhodohypoxis Nel, Spiloxene Salisb. and Empodium Salisb.; it may also perhaps contribute to the understanding of generic origins within Hypoxidaceae.

### 2.1.1. Herbarium studies

The foundation of this thesis was laid by the examination of herbarium material in conjunction with reference to relevant literature. From study of dried specimens from a range of localities and habitats, entities were delimited and the gross range of their morphological variation, distribution and ecological preferences determined. Problems requiring further study either in the field or the laboratory were exposed.

Specimens from the following herbaria were made available for study: Albany Museum, Grahamstown; Bolus Herbarium, University of Cape Town; Botanical Research Institute, Pretoria and Stellenbosch; British Museum (Natural History), London; Compton Herbarium, ? Kirstenbosch; Natal Herbarium, Durban; Natal Parks Board, Cathedra1 Peak; Royal Botanic Gardens, Kew; and the University of the Witwatersrand. A11 specimens studied are cited (see Chap. 2.5 Enumeration of Species, for citations, and Appendix for an index to collectors, collectors' numbers and taxa to which the specimens belong).

The method of specimen citation used in this thesis is a modification of the Degree Reference System proposed by Edwards \& Jessop (1967), and modified by Edwards and Leistner (1971). The herbarium abbreviations used in the citations are those of Holmgren \& Keuken (1974), except for the Natal Parks Board, Cathedral Peak
'which is designated NPC. Where a collector's number is lacking, either the herbarium number, or the date of collection is quoted, for example (SAM 92654), or (SAM, Oct. 1964), respectively.

### 2.1.2. Field Studies

Pield work was confined to Natal. Population studies enabled more accurate observation of plant habitats, habit and duration of growth, range of morphological variation and duration of flowering. It also often resulted in the extension and refinement of locality records. Of particular value was the opportunity it provided to study hair types and densities, both within plants during seasonal growth, and within and among whole populations. Inflorescence form and the developmental stages of this; ovary form, fruit development and dehiscence and seed maturation were also able to be studied. Character variation with ageing, especially changes in vesture of leaves and inflorescences, could be followed effectively only in living material. Corm size in relation to overall plant size and the rature of the contractile roots were also best studied in the field.

Where possible at least one plant was transplanted and growi under cultivation. This provided a constant source of living material for comparison and for dissection. Specimens collected in the field, especially leaves and inflorescences, were also preserved for anatomical study. Additional herbarium material was prepared where necessary in order to supplement existing collections.

### 2.1.3. Laboratory Studies

Anatomical investigations of hair types and seed coat structure were made eithex from hand sections or by embedding in wax and micro-
toming. A detailed anatomical study, however, was not attempted as part of this thesis. Surface structures of the seed coat were examined on the Transmitting Electron Microscope and the Scanning Electron Microscope. In preparation for scanning, dry seeds from mature capsules were mounted on stubs with double sided adhesive tape, precoated with carbon, and then coated with 250 angstroms of gold palladium in an Hitachi HUS3B coating unit. They were examined in an Hitachi SSM-2 Scanning Electron Microscope and photographed by an Asahi Pentax camera.

NOTE: In all figures the scale-1ine represents 1 cm unless otherwise stated.

### 2.2 Diagnostic Characters

Classification within the genus Hypoxis depends main1y upon the vegetative characters because of the high level of uniformity in floral structure. The classificaLory value of any one of the characters discussed below may differ from species to species, thus identification depends not on single features but on correlations of characters.

In discussing characters used in species diagnosis, I have included comments on features employed by baker (1878 \& 1896) and Nel (1914) in their treatments of the genus.

### 2.2.1. Rootstock

Baker (1896) used corm size to differentiate $\underline{\text { K. filiformis and }}$ H. kraussiana but this is unsatisfactory, especially as overlap in size range occurs.

All taxa possess a fleshy mucilagenous corm that is bright yellow or white internally. Corm sizes range from $0,5-11,0$ cor in diameter and shapes vary from oblong to turbinate or globose.

Knowledge of the patterns of growth of corms is very Iimited. In this study seeds could not be germinated (there seems to be some dormancy system in operation), thus the study of seedlings and the development of young corms was not possible. Until experiment provides a method that will promote seed germination within a reasonable period, study of corm growth is not practical.

Preliminary observations show there to be at least three types of corm growth within the Natal species studied. The first is represented by the H. rooperi (Fig. 2a) in which a single corm persists throughout the life of the plant. Growth is from one apical meristem and usually only one aerial shoot per season is produced. The other types of corm development involve what is probably sympodial branching with the development of lateral buds. One exanple is H. acuminata where growth is initiated from one apical meristem (Fig. 2b). The corm either remains single or many lateral buds develop. These on growth result in the formation of a "plate" of rootstock material from which many aerial shoots arise (Fig. 3).
H. angustifolia var. buchananif exhibits the growth form in which lateral buds develop into slendcr rhizome-Iike portions that later thicken with the deposition of food reserves. The parent corm thus appears to become twisted or lmotted (fig. 4).

H. angustifolia: plant showing twisted single corm.


Fig. 4. H. angustifolia: twisted and knotted corm that has probably arisen as a result of sympodial branching.

Note: arrow indicates rhizome-like portion which probably later thickens with deposition of food reserves.

In addition to the above two species, corms of H . multiceps and $H$. costata were also found to exhibit sympodial growth.

Corm shape and the presence; or not, of sympodial branching often add confirmatory value to an identification.

### 2.2.2 Leaf

Plants of Hypoxis are acaulescent with the leaves arising directly from the corm, often in three ranks. There is a considerable range in the erectness and rigidity of the leaf, but this organ does provide features of value in diagnosis, especially vesture, venation and shape, in this order of importance.
shape: Leaves may be oblong, lanceolate, linear, acuminate, sickleshaped or a combination of these forms. The degree of variation in leaf shape differs from species to species. K. filiformis has linear, filiform, subterete leaves which exhibit little variation, while , leaves of $H$ : costata vary from oblang to lanceolate and exhibit the most extensive range of variation of any Natal species. Use of leaf shape in identification has its limitations as it is often difficult for the worker unfamiliar with the genus to distinguish between, for example, linear-lanceolate and linear-acuminate leaves. It is, however, a useful criterion in distinguishing extremes. Leaf shape was used by both Baker (1878, 1896) and Nel (1914) in their keys.

Leaf length cannot be used in classification because it is the most variable parameter in the genus. In one year's season length may vary from 10 cm to 100 cm (Figs. 5 \& 6), thus making it impossible to give reliably, an average leaf length for any taxon. Nel used this character in separating species within the groups, but it is not satisfactory.


Fig. 5. H. rigidula: plant a - present year's leaves, b - last years old leaves still attached


Fig. 6. H. rigidula: two plants showing extremes in leaf length; left: plant with spring growth, leaves short; right: plant with summer growth, leaves much elongated.

Leaf width also tends to exhibit a range of variation within a species, but it again is useful to differentiate extremes, for example the leaf of H . filiformis is not wider than 3,0 m wide while that of H. galpinii is $1,5-4,5 \mathrm{~cm}$ wide. Both Baker and Nel used this character but it is apparent that they often had available only one, or at the most a few, specimens of the taxon for study, and thus could not record accurately the range of variation. This led to difficulties in using their keys.
venation: Nel used leaf venation as one of his major diagnostic
characters for differentiating his groups within the genus. A
translation from his German text reads as follows:

The leaf laminae are throughout strongly ribbed, a characteristic as I will say at a later stage, which lends itself well to a systematic classification of the genus. It is established that the number of nerves in the different species is variable. The number of nerves provides not only a good characteristic for distinguishing the different species but it is also very valuable in building groups. Not only does the number of nerves vary but one finds that different nerves themselves are unevenly thickened. One observes that there are numerous species in which on every leaf half one, and more seldom, two ribs are more pronounced in comparison to the others; while by the way of contrast in other species the nerves remain equally thickened.

Diagnostically venation is a good character. However Nel
tended to be too adamant as to the number of nerves per leaf per species, and did not take into account range in number according to range in leaf width. The presence of prominent nerves, and the regularity of the nerves are undoubtedly good characters for identification of species in this genus.
vesture: This is probably the most important character in diagnosing taxa within the genus.


Fig. 7. Hair types represented in the Natal species of Hypoxis - a-c, stellate; d - bifurcate; esimple.

Plants possess simple, bifurcate or stellate hairs (Fig. 7) that clothe the leaves. Hair type is constant for a species, but often more than one type is represented within a taxon (either simple and bifurcate, or bifurcate and stellate, intermingled). A number of species exhibit similar hair types and thus identification is not possible on type alone. However in conjunction with hair positioning, angle, and colour, good criteria are established for identifying the Natal taxa. Baker used degree of hair covering as a criterion in his key, while Nel used this character as well as hair angle.

Hairs may clothe the entire leaf surface, they may be scattered over the leaf surface without a concentration on margins and keel, or the abaxial surface may be more densely clothed than the adaxial. Hairs may also occur only on margins and keel with the laminal surfaces glabrous. Hair positioning is constant within a species but density may vary. An extreme example of this is H. costata where, although hairs are always found on the keel and margins; the lamina may range from subglabrous to densely hairy (possibly a result of environment). Plants of $H$. oblonga were also noted to differ in hair density according to aspect. In many Natal species old leaves tended to have fewer hairs in comparison with the younger leaves.

Hair angle is from patent to appressed and is constant within a taxon. Hairs on the leaves of plants of H. argentea var. argentea, a Cape variety, produce a sericeous effect, not present in any Natal species.

Hair colours present within the genus are white, yellow, golden and light and red-brown. This feature is useful in conjunction with other aspects of vesture.

## 2.2 .3 <br> Peduncle

All taxa have one or more axillary peduncles which axe usually stellately hairy and terminate in an inflorescence. They are either ancipitous or terete, and in several species they become grooved and subglabrous towards the base. The peduncle is of little importance in classification.

### 2.2.4 Pedicel

Pedicels are also usually stellately hairy and range in length from 0,1 to $6,5 \mathrm{~cm}$. A range in length is always represented within a taxon. Pedicel length is useful in distinguishing species with subsessile flowers from those with long-pedicelled flowers. It is important to measure the pedicel from the bract and not to include the continuation of the main axis. Baker did not use characters of the peduncle nor of the pedicel in his keys, but Nel used pedicel length in a number of instances.

### 2.2.5 Inf1orescence

Baker (1896) recognized two main forms within Hypoxis, corymbose and racemose. Ne1 (1914) also recognized these types but substituted as an almost equivalent term, umbels for corymbs, and included spikes. However both authors experienced difficulty in grouping the inflorescences of all the species under these types and introduced the suffix "sub-" as an ald. But with such intermediate possibilities, it became confusing to attempt to recognize and delimit "sub-umbellate", "almost corymbose", "sub-racemose" and "spicate".

Lawrence (1969), based on the earlier work of Rickett (1944), put forward a schematic diagram suggesting the hypothetical evolution


Fig. 9. The range of inflorescence types within the Natal species of Hypoxis. a - raceme, for example H. latifolia; e - corymb, for example H. gerrardii; b, c, d - reduced types not readily placed as racemose or corymbose, for example H. costata, H. parvula and H. filiformis respectively.
of inflorescence types. The following series of diagrams, from this scheme, encompasses the range represented within Hypoxis (Fig. 8).


Fig. 8. Inflorescence types (Lawrence, 1969). 1 - compound corymb (not present in Hypoxis); 2 - simple corymb; 3 - raceme; 4 - spike.

However as can be seen from Fig. 9, the inflorescence of Hypoxis do not always fit conveniently into these types. The two extremes are well defined, corymbose (for example H. gerxardii, Fig. Pe) and racemose (for example H. latifolia, Fig. Pa). These extremes together with a reduced inflorescence (H. parvula which is always only 1-flowered, Fig. Pc), may be used in species diagnosis. However,
many examples of the Natal species fall between the extremes of corymb and raceme. In such intermediates the inflorescence type cannot be decided from consideration of the mature flowering head alone. Even If developmental stages are studied it is not always possible to differentiate corymbose from racemose examples. Thus inflorescence form is far from reliable in diagnosis, except in special cases.

### 2.2.6 Perianth segments

The six, free perianth segments of all taxa are oblong to oblong-1anceolate. A considerable range in size is represented within a taxon, and overlap among different species occurs. This character has not been used in the key or in the descriptions.

Baker placed considerable emphasis on perianth length in his key (1896). Nel also used this parameter occasionally but did not place emphasis on it, nor did he use shape. A translation from his text in German reads as follows:

The form of the perianth segments offers no characteristics for classification because it changes very frequently and is not even always constant in the same plant.

The segments are either yellow or white. Only H. parvula (which also has plants with yellow segments), and H. membranacea have white perianth segments.

### 2.2.7 Stamens

There are always six free stamens connate to the base of the perianth segments. The filaments are filiform or subulate, equal or unequal (usually in two groups of three with the longer filaments


Fig. 10. Stigmatic types represented within Hypoxis:
a - H. membranacea and H. parvola,
b - all other taxa
attached to the outer segments). Anthers are lanceolate, sagittate and versatile with their apex entire or bifurcate.

The most important features are the lengths of the filaments and the state of the anther apex (i.e. bifurcate or entire). These are constant within a taxon but I have not used them in the key firstly because I have concentrated more on vegetative features, and secondly because similar species often have the same type of stamen.

Ne1 attaches great importance to the anther apex, and in fact separates two groups on this feature alone:

At the point the anthers are either clefted or unclefted. A characteristic which I have used ror the classification of the species.
(translation from the German text)
Baker (1896) made no reference to this character.

### 2.2.8 Stigma and Style

There are two distinct types of stfgma and style within the Natal species:
a) that of H. membranacea and H. parvala: the style is long and filiform with a minute stigma that appears globose. This stigma is most probably a very reduced form of that described in b) (Fig. 10a);
b) that of all other taxa: the style is cylindrical, subulate, or the stigna is subsessile. The stigma is a concrete structure consisting of three fused lobes. It is pyramidal in shape and has three concave faces. (Fig. 10b).

Variation in stigma size between the taxa is negligible. In some specimens the stigma lobes separate, but this is not a constant phenomenon and is most probably attributable to age.


Fig. 11. Dehiscence mechanisms of capsules; a\&b-H. latifolia (capsule opens by circumscissile dehiscence, with no lateral splitting of valves); $c, d \& e-H$. angustifolia (circumscissile dehiscence followed by the splitting of three longitudinal valves).

I have only used these characters to separate out H. membranacea and H. parvula. Nel however attached great importance to the relationship of stigma to style and separated some of his groups on this character:

The groups Angustifoliae, Orbiculatae, Infaustae, Recurvatae and Subspicatae are characterized by a style which is either longer than the stigma or approximately the same length. A sessile stigma or a stigma which is much longer than the style is characteristic of the groups Argenteae, Nyassicae, Oligotrichae, Rigidulae and Obtusae.

(translation from the German text)
Ne1 also distinguished H. membranacea on this character, but Baker did not use it in his key.

### 2.2.9 Fruit

The fruit of Hypoxis has, from the earliest descriptions (Baker, 1878) been the chief character in distinguishing the genus within the family Hypoxidaceae. The fruit is a capsule which opens by circumscissile dehiscence below the apex (Fig. 11). In two species, H. angustifolia and $\underline{H}$. gerrardii, the dehiscence proceeds further This involves splitting of the three valves longitudinally. The three portions of the capsule bend backwards and expose the seeds and placenta. (Fig. 11). Capsules on plants of H. latifolia have also shown this feature but only after the seeds were shed; and the inflorescence had become dry and shrivelled. It therefore appears probable that all species have the potential for these valves to split, but that it is not often realized.

### 2.2.10 Seed

Mature seeds are black with a similar obconic shape and pronounced projecting funicle and microple that lie in close proximity to one another. There are two groups, smooth-coated and papillate


Fig. 12. H. costata: a - seed (plate width $=2,3 \mathrm{~mm}$ ); b -. external features of seed coat (plate width $=0,58 \mathrm{~mm}$ ).

a.

b
Fig. 13. H, latifolia: a - seed (plate width $=5,7 \mathrm{~nm}$ ); b - external features of seed coat (plate width $=0,57 \mathrm{~mm}$ ).

$\mathbf{a}$

b

Fig. 14. H. obtusa: $a$ - seed (plate width $=5,7$ mom); $\bar{b}$ - external features of seed coat (plate width $=0,57$ man).

a

b
Fig. 15. H. angustifolia var. buchananii: a - seed (plate width $=2,3 \mathrm{~mm}) ; \quad b-$ external features of seed coat (plate width $=0,57 \mathrm{~mm})$.
(Figs, 12-15 show the range that occurs within the Natal species). The seed coat was hard and impermeable (even when cracked, seeds would not germinate). This led to difficulty in cutting sections which were needed in order to determine whether the papillae developed with maturation of the seed coat, or whether they were present at an early stage of development.

Results showed there to be no clear-cut difference in seed surface structures between the species. Thus this feature is not satisfactory for diagnostic distinction between the two groups, nor in the identification of individual taxa.

### 2.3 Description of the Genus

This description is relevant to the Natal species only. With a full revision of the genus in southern Africa, amendments may be necessary.

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Hypoxis L. Syst. Nat. 10;7 (1759); Gen. P1. no. 417 (ex parte)
    (1764); Roem. et Schultes, Syst. Veg. 7, 46:759 (1830);
    Herbert, Amaryll. 65 (1837); Endl. Gen. no. 1264 (1840);
    Salisb. Gen. PI. Fragm. 44 (1866); Baker in J. Linn. Soc.
    17:98 (1878) (excluding subgenus Ianthe); Bentham &
    Hooker, Gen. Pl. 3, 2:712 (1883); Pax in Naturl. Pflanzen
    2, 5:121 (1887); Baker in F1. Cap. 6:178 (1896) (excluding
    subgenus Ianthe, H. milloides and H. baurii); Nel in Engl.
    Jahrb. 51:239.(in clavi) (1914); Phil1. Gen.: t. 1230
    (1951).
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Herbs perennial with a fleshy; mucilaginous corm, acaulescent.
Leaves 2-many, filiform, lanceolate, linear or oblong, hairy or glabrous, often conspicuously nerved, the first leaf is a membranous sheath surrounding a neck formed from the bases of the inner leaves, and usually situated below ground level; remnants of old leaves form bristles crowning the corm. Peduncles ancipitous or terete, hairy, especially immediately below the flowering portion. Inflorescence scapose, axillary, l-many flowered. Flowers pedicelled or subsessile, bisexual, regular, epigynous. Bracts subtending pedicels, rarely absent, linear or linear-subulate, hairy along keel. Ovary inferior, clavate-oblong, hairy, trilocular. Perianth segments six, free, ovate-oblong, in two ranks, the outer yellow or white and glabrous adaxially, green or green and yellow and hairy all over abaxially; the inner as above except that there is only a central vertical line of hairs abaxially. Stamens six; filaments attached to the base of the perianth segments, subulate or filiform, equal or unequal, if unequal usually the outer longer than the inner; anthers 2-celled,
versatile, lanceolate, sagittate, apex entire or bifurcate. Style cylindrical, subulate, filiform or stigma subsessile. Stigmas three, free or concrete with three concave faces; pyramidal and equal to or longer than the style, or spherical and minute. Fruit a capsule opening by circumscissile dehiscence below the apex, sometimes accompan1ed by the splitting of three valves of the capsule. Seeds many, black, round to elliptic-oblong; with funicle and cone-like micropyle forming two protuberances on the same side of the seed, outer coat smooth or papillate.

1. Leaves glabrous ..... 2
Leaves hairy ..... 3
2. Leaves $1,0-1,5 \mathrm{~cm}$ broad; inflorescence 2-f1owered3. H. Interjecta
Leaves 2,5-11,0 cm broad, often over1apping, at least basally,to form a funnel; inflorescence 6-21 flowered. 1. H. latifolia
3. Stigma spherical, minute, $\pm 5,5$ mim long, much shorter than thefiliform style that is usually $\pm 3,0 \mathrm{~mm}$ long4
Stigma concrete, pyramidal with 3 concave faces, equal to orlonger than the cylindrical or subulate style5
4. Inflorescence 1-flowered, perianth segments yellow or white adaxially, bracts absent or only one that is $1,0-4,0$ man long
5. H. parvulaInflorescence l-3 flowered (on a plant there will always be onepeduncle that is more than 1-flowered), perianth segments whiteadaxially, bracts 2, 0,3-0,6 cm long ..... 20. H. membranacea
6. Leaf hairs brown ..... 6
Leaf hairs yellow, golden or white ..... 7
7. Hairs light brown, scattered over both surfaces of the leaf
19a. H. sobolifera var. sobolifera
Hairs red-brown, densely covering both surfaces of the leaf
19b. H. sobolifera var. pannosa
8. Leaf hairs golden giving a sericeous effect to the leaf ..... 8
Leaf hairs white or yellow ..... 9
9. Hairs appressed, densely covering abaxial surface of the leaf 16a. H. argentea var. argentea Hairs appressed, situated predominantly on margins and keel, scattered over the lamina of the leaf
16b. H. argentea var. ..... sericea
10. Leaves membranous ..... 10
Leaves not membranous ..... 12
11. Leaves sickle-shaped, hairs stellate or bifurcate or both typesintermingled, patent, situated predominantly on margins and
keel 18. H. lata
Leaves lanceolate, linear-acuminate or oblong-lanceolate, hairssimple and bifurcate intermingled, not patent, scattered overthe leaf surface11
12. Leaves lanceolate or 1inear-acuminate, $0,2-0,9 \mathrm{~cm}$ at widestpoint, semi-erect .. 17a. F. angustifolia var. angustifolia
Leaves lanceolate to oblong-acuminate, $0,6-1,7 \mathrm{~cm}$ at widest
point, flaceid 17b. H. angustifolia var. buchananii
13. Leaves sickle-shaped ..... 13
Leaves not sickle-shaped ..... 15
14. Leaf hairs stellate and bifurcate, dense on margins and kee1,usually appressed, lamina glabrous to pilose14
Leaf hairs stellate and bifurcate, often U-shaped, mainly onkeel and margins, but not dense, patent, lamina glabrous topilose18. H. lata
15. Laminal hairs lying in channe1s between the ribs

Lamina glabrous or occasionally with scattered hairs at the base
9. H. obtusa
15. Leaves rigid, erect, linear, linear-lanceolate, linear-acuminate, (rarely oblong-lanceolate), filiform or subterete ........... 16 Leaves not rigid, or, if rigid, oblong to oblong-lanceolate or sickle-shaped 19
16. Leaves linear, $0,1 \sim 0,3 \mathrm{~cm}$ at widest point, hairs yellowish, predominantly bifurcate, appearing U-shaped, situated mostly along margins and keel ...................................... 13. H. neliana Leaves not as above, hairs white ................................... 17
17. Leaves with a prominent vein on either side of the keel, hairs, if present, not appressed ..................................................... 18

Leaves not as above ....................................................... 22
18. Leaves $0,2-1,5 \mathrm{~cm}$ at widest point, laminal hairs bifurcate, scattered to dense on both surfaces (abaxial surface usually more densely covered than adaxial surface) ...... 8. H. oblonga Leaves $1,5-4,5 \mathrm{~cm}$ at widest point, laminal hairs lacking, if present stellate, scattered on prominent ribs .. 2. H. galpinii
19. Leaves $0,1-0,3 \mathrm{~cm}$ at widest point, ribs $6-12$ regular, hairs predominantly bifurcate, scattered over the whole leaf surface not concentrated on margins and keel; inflorescence 1-4 flowered, first formed flower with longest pedicel, youngest flower usually subsessile 20

Leaves $0,2-1,3 \mathrm{~cm}$ at widest point, ribs 6-20, irregular, hairs stellate (predominant) and bifurcate intermingled, dense on margins and keel, and lying in channels between the ribs, or both surfaces of the lamina woolly or white appressed hairy;
20. Leaves lightly pilose all over, hairs fine, white, simple and bifurcate intermingled, not patent; peduncles silky villous below inflorescence becoming subglabrous towards the base
12. H. filiformis

Leaves sparsely to densely covered with white, stellate and bifurcate (predominant) hairs intermingled, usually patent; peduncles hairy all over, hairs patent .... 14. H. kraussiana
21. Laminal hairs appressed, lying in the channels between the ribs 6a. H. rigidula var. rigidula Lamina woolly or densely white appressed hairy on both surfaces, hairs not confined to channels between the ribs 6b. H. rigidula var. pilosissima
22. Leaves rigid or semi-rigid or markedly sickle-shaped ........ 23

Leaves not rigid, not sickle-shaped ............................. 26
23. Qeayes lanceolate to oblong-lanceolate, never twisted with age, hairs yellowish (especially on inflorescence); inflorescence 1-5 flowered 24

Leaves linear-lanceolate or lanceolate, occasionally long and twisted with age, hairs white or opaque, inflorescence 2-15 Elowered 25
24. Leaves with thickened submarginal vein, lamina glabrous to densely hairy, hairs if present, predominantly bifurcate 5. H. costata

Leaves without thickened submarginal vein, lamina densely hairy, hairs short, stellate, rarely with bifurcate intermingled
4. H. multiceps
25. Leaves sometimes in 3-ranks, linear-lanceolate, slightly sickle-shaped, becoming twisted and curving backwards with age, lamina glabrous or occasionally with scattered hairs at the base, keel and margins covered with white, opaque, appressed hairs ........................................................ 9. H. obtusa

Leaves clearly 3-ranked, lanceolate, sickle-shaped, not becoming twisted with age, hairy all over (abaxial surface more densely covered than adaxial) ..................... 10. H. rooperi
26. Leaves $0,1-0,4 \mathrm{~cm}$ at widest point, hairs dense on keel and margins, lamina glabrous or with hairs mainly at the base between the ribs; or leaf 1 ightly pilose all over, hairs not U-shaped

Leaves $0,4-1,2 \mathrm{~cm}$ at widest point, hairs woolly villous all over; or sparse on adaxial surface, dense on abaxial surface, hairs to keel and margins often U-shaped ................... 28
27. Leaves linear, $0,3-0,4 \mathrm{~cm}$ at widest point, with 10-20 uniform ribs, hairs stellate, dense on keel and margins, lamina glabrous or hairs mainly at the base between the ribs
11. H. zululandensis

Leaves linear $0,1-0,3 \mathrm{~cm}$ at widest point, with $6-8$ prominent ribs, hairs simple and bifurcate, lightly pilose over 1eaf surface
12. H. filiformis

Leaves linear-1anceolate, with usually 2 (seldom 4) prominent ribs, hairs fine, whitish-golden, woolly villous all over; hairs on margins and kee1 not U-shaped ...... 15. H. gerrardii Leaves linear-lanceolate or linear-acuminate, ribs 6-12, hairs patent, stellate or bifurcate, sparse on adaxial surface, dense on abaxial surface, hairs to keel and margins often U-shaped

1. Hypoxis latifolia Hook. in Bot. Mag. t. 4817 (1873); Baker in J. Linn. Soc. 17:115 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:232 (1893), et in Fl. Cap. 6:185 (1896); Medley Wood, F1. Natal:132 (1907); Bews, Fl. Natal \& Zululand: 65 (1921). Type: South Africa, Natal, Adlam s.n. (K, ho1o.!
H. colchicifolia Baker in J. Bot. (Lond.):3 (1889), in Dur. \& Schinz, Consp. P1. Afr. 5:231 (1893), et in F1. Cap. 6:186 (1896). Type: South Africa, s.n. Bull (K, holo.)
H. oligotricha Baker in J. Bot. (Lond.):3 (1889), in Dur. \& Schinz, Consp. FI. Afr. 5:232 (1893), et in Fl. Cap. 6:187 (1896); Medley Wood, Fl. Natal:132 (1907); Nel in Engl. Jahrb. 51:321 (1914); Bews, F1. Natal \& Zululand:65 (1921); Ross, FI. Natal:132 (1972). Type: South Africa, Natal, Clairmont, Medley Wood 1170 (NH, holo.!)

Corm 4, 0-7,0 cm in diam., globose-turbinate. Leaves 5-12, $11,0-60,0 \mathrm{~cm}$ long, $2,5-11,0 \mathrm{~cm}$ at the widest point, 3 -ranked, often overlapping, at least basally, to form a funnel, erect, coriaceous, oblong-lanccolate, strongly ribbed with $30-90$ veins, glabrous occasionally with sparse stellate hairs on the margins, (except BurttDavy 13457 which has stellate hairs all over), leaves deciduous. Peduncles ancipitous, with few, fine, white stellate hairs below raceme. Inflorescence up to $14,0 \mathrm{~cm}$ long, excluding pedunc1e, 6-21 flowered. Pedicels of lower open flowers $1,0-2,0 \mathrm{~cm}$ long, of upper $\pm 0,1-1,0 \mathrm{~cm}$ long. Bracts subulate, $1,0-2,5(-3,1) \mathrm{cm}$ long, glabrous to lightly setose. Ovary globose-turbinate, subglabrous with few, stellate hairs. Perianth segments yellow adaxially, in open flowers up to $2,0 \mathrm{~cm}$ long, $0,4-0,6 \mathrm{~cm}$ wide at the broadest point. Filaments subulate, equal. Anthers 4,0-7,0 mm long, lanceolate, sagittate, apex entire or slightly bifurcate. Style subulate, approximately 1,0 mm long. Stigma concrete, (of 3 loosely fused sections that


Fig. 16. Hypoxis latifolia Hook. - known distribution in Southern Africa up to 1975.
tend to separate with age), 3,0-5,0 mm long, $1,0-1,5 \mathrm{~mm}$ at widest point, pyramidal with 3 concave faces. Fruits a capsule, opening by circumscissile dehiscence below the apex.

Flowering Period: August-February.

Hooker in 1873 described Hypoxis latifolia. Baker in 1889 established Hypoxis colchicifolia and Hypoxis oligotricha. H. colchicifolia was described as having shorter leaves, fewer flowers, longer pedicels and a more hairy inflorescence than H. oligotricha. In his key in Flora Capensis (1896), Baker separated them on inflorescence type, that of $H$. colchicifolia being corymbose and that of H. oligotricha racemose. However, it has since been determined from the type that $\underline{H .}$ colchicifolia has a racemose inflorescence. Examination of the types of these two species has shown them to be conspecific with the type of $H$. latifolia, thus they must both become junior synonyms.

Plants of H. latifolia are concentrated in Natal south of the Tugela River, from the Estcourt district to the Mtamvuna River. It is not yet known if this river is the southernmost limit of this species or whether the apparent absence of specimens further south is due to inadequate collecting.

Only two specimens, Moss 13679 (Johannesburg) and Burtt-Davy
13457 (Vryheid) have been found outside southern Natal. Moss 13679 is glabrous and thus conforms with the species. Burtt-Davy 13457, possibly represents a variant since its leaves are covered with stellate hairs. Further collecting in northern Natal and the Transval is needed to determine if there is a range from the glabrous to


Fig. 17. H. latifolia: a - centre leaf, b - outer Ieaf.
the hairy forn, or whether the latter should be regarded as a distinct varlety. The discontinuities in the known distribution (Fig. 16) cannot, at present, be adequately explained.

Hypoxis latifolia is a very robust species usually found growing in poor and sandy soils in undisturbed or disturbed grassveld. The corm is large in the mature plant, turbinate and bright yellow internally when freshly cut. Adventitious, contractile roots, which arise from the top of the corm, are comparatively thick (4-6 mm diam.).

Towards the middle of the flowering season the leaves developed are characteristically large and glabrous (Fig. 17a), with a tendency for the margins to become wavy with age. The plant can reach up to 60 cm in height and although the leaves appear bunched or whorled on an herbarium sheet, they are in fact 3-ranked. The outermost leaves are smaller than those in the centre (Fig. 17b). The inflorescence is racemose with never less than 6 flowers per raceme. It is sparsely covered with hairs (that is, over peduncle, ovary and perianth segments).

This species is easily identified in the field and is without any close ally in the genus. Morphologically H. galpinii is closest to H. latifolia but the plants have narrower leaves with hairy margins, and hairier inflorescences.

Citations:

| TRANSVAAL | - | 2628 | $\begin{aligned} & \text { (Johannesburg): Johannesburg (-AA), Moss } \\ & 13679 \text { (J). } \end{aligned}$ |
| :---: | :---: | :---: | :---: |
|  |  | 2730 | (Vryheid): Vryheid (-DD) surtt-Davy 13457 (PRE). |
| NATAL | - | 2929 | (Underberg): Tabamhlope (nAB), Miller 150, |
|  |  |  | 202 ( $\mathrm{NH}, 3$ sheets); Estcourt ( $-\overline{\mathrm{BB} \text { ), Barker }}$ 21449 (NBG). |
|  |  | 2930 | (Pietermaritzburg): Near Howick (-AC), |
|  |  |  | Medley Wood 5193 (NH); Greytown (-BA), |
|  |  |  | Wylie s.n. (PRE, NH 27973); Gramond (-BC), |
|  |  |  | Staples s.n. (NH 18533); Hawthorns Hill |
|  |  |  | (-GB), A11 sopp 520 (NU), 909 (NH, NU); |



Fig. 18. Hypoxis galpinii Bak. - known distribution in Southern Africa up to 1975.

Hilton (-CB), Trace 59 (NU); Pietermaritzburg (-GB), Herbst s.n. (NBG 78, 441), Stirton s.n. ( $\mathrm{NU}, 21.10 .75$ ), S.E. Wood 73, 85 (NU); 11 km. to Mid-illovo from turn ofe from Eston Road (-CD), S.E. Wood 111 (NU).
2931 (Stanger): Clairmont, near Durban (-CC), Medley Wood 1170 (NH, SAM).
3030 (Port Shepstone): Station Dumisa (-AD), Rudatis 1791 (PRE); St. Michaels-on-Sea (-CB), Nicholson 797 (PRE); Marina Beach (-CB), Strey 5949 (NH); on road to P. Edward from Izingolweni (-CC), S.E. Wood 161 (NU); Beach Terminus (-CD), Thode s.n. (STE 2542, 4662); 7 miles north of P. Edward (-CD), Strey 4521 (NH).
3130 (Port Edward): Port Edward (-AA), Ward 6654 (PRE).
2. Hypoxis galpinii Baker in F1. Cap. 6:188 (1896); Nel in Eng1. Jahrb. 5l:320 (1914); Verdoorn in Wild Flowers of the Transvaal:69 (1962). Type: South Africa, Transvaal, Saddleback Range, near Barberton, Galpin 1098 (GRA, iso.!)

Corm 2,1-4,5 cm in diam., turbinate-globose, crowned with bristles. Leaves $4-8$, up to $50,0 \mathrm{~cm}$ long, $1,5-4,5 \mathrm{~cm}$ at widest point, not noticeably 3-ranked, erect, linear-acuminate to oblonglanceolate, ribs 20-50, not compact, 2 more prominent than the rest, hairs stellate, white, scattered on keel and margins, rarely on the prominent nerves, the abaxial surfaces of the flrst membranous leaves also covered with stellate (occasionally bifurcate) hairs. Peduncles ancipitous, hairs stellate, usually patent, most abundant below inflorescence grading to few at the base. Inflorescence 4-1l-flowered. Pedicels of lower open flowers $0,1-1,5 \mathrm{~cm}$ long (flowers often appearing subsessile), pedicels of upper open flowers $\pm 0,1-0,7 \mathrm{~cm}$ long, densely villous. Bracts subulate, $1,0-3,5 \mathrm{~cm}$ long, up to $0,3 \mathrm{~cm}$ wide, setose.


Fig. 19. H. galpinii: variation in leaf shape (from apex to half way down lanina)

Perianth segments yellow adaxially, in open flower $0,9-1,8 \mathrm{~cm}$ long. Filaments subulate, equals 2,0-3,0 mm long. Anthers lanceolate, sagittate, 5,5-7,0 mm longs apex entire or slightly bifurcate. Style subulate-cylindrica1 $1,0-2,0 \mathrm{~mm}$ long. Stigma concrete, pyramidal with 3 concave faces, 2,0-4,0 min long. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: September-February.
H. galpinii has not previously been recorded for Natal. The distribution as at present known is from Haenertsburg (N. Transvaal), through Swaziland, to Mount Currie (Cape Province) (Fig. 18). Populations grow in grassveld and disturbed grassveld, commonly at high altitudes, but plants in more southern latitudes occur near the coast.

Plants are characterized by obtuse-apiced, linear leaves (Fig. 19), which turn a dark red-brown when dry. The leaves are neither conduplicate nor arranged in 3 ranks, in which features they are unlike many of the Natal species, for example $\underline{H}^{\text {. rooperi, } H . ~ o b t u s a . ~}$ The lower pedicels of the inflorescence are unusually short ( 0,1 $1,5 \mathrm{~cm}$ long). H. latifolla is the closest morphological ally of H. galpinii (for details see under the former species).

Medley Wood 373 (NH) in my opinion is H. galpinif, but Baker (F1. Cap., 1896) cited it as H. rooperi S. Moore. It does not have the sickle-shaped, conduplicate leaves covered with stellate hairs, that are characteristic of H . rooperi.

## Citations:




Fig. 20. H. interjecta: variation in leaf shape
3. Hypoxis interjecta Ne1 in Engl. Jahrb. $51: 321$ (1914). Type: South Africa, Transvaal, Lydenburg, Wilms 1454 (BSB, holo.)

Corm 2,0-5,0 cm in diam., oblong-globose, crowned with bristles. Leaves $7-9,3,5-15,0 \mathrm{~cm}$ long, $1,0-1,5 \mathrm{~cm}$ broad at widest point, 3ranked, oblong-lanceolate, erect to semi-erect, with 20-40prominent ribs, glabrous. Peduncles ancipitous, densely hairy, hairs stellate with one arm longer than the remainder, usually yellow, opaque. Inflorescence 2-flowered. Pedicels $0,6-1,4 \mathrm{~cm}$ long at time of flowering, densely hairy. Bracts Iinear-subulate, setose, 0, 6-1,1 cm long. Perianth segments yellow adaxially, in open flowers l,0-1,5 cm long. Filaments subulate, equal, $\pm 3,0$ wan long. Anthers 4, 5-5, Onm long, lanceolate, sagittate, apex entire or very silghtly bifurcate. Style subulate or stigma subsessile. Stigma concrete, 1,5-2,0 man long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: September-December.

Hypoxis interjecta has small (usually less than 15 cm in length), glabrous leaves that vary in shape from oblong to lanceolate (Fig. 20). Peduncles are particularly hairy with stellate, spreading and usually yellow hairs. In all specimens seen the inflorescence was two-flowered. H. interjecta is a grassveld species, recorded from altitudes approximately between 1000-2000 metres.

The closest morphological ally of $H$. interjecta is H. multiceps. In this species the shape of the leaves is similar, but they are larger attaining approximately 20 cm in length and 5 cm in width. They are completely covered with ehort stellate hairs, in contrast to the


Fig. 2l. Hypoxis interjecta Nel - known distribution in Southera Africa up to 1975.
glabrous leaves of $H$. interjecta. The inflorescences of the species are notably alike, although H. multiceps may occasionally carry up to four flowers per inflorescence. H. latifolia is the only other Natal species with glabrous leaves, but otherwise these taxa are distinct.
H. interjecta has not previously been recorded for Natal.

Nel (1914) cited only the type specimen (Lydenburg, Transvaal). Other Transvaal specfmens have not been associated with H. interjecta, but were grouped together as an undescribed taxon (manuscript name $\underline{H}$. pretoriensis). Two specimens only are known from Natal, both from the Estcourt district. The discontinuity in the distribution (Fig. 21) shows the general need for collecting, especially in northern Natal and the northern Orange Free State.

| TRANSVAAL | - | 2528 | (Pretoria): Muckleneuk (-CA), Goossens 91 |
| :---: | :---: | :---: | :---: |
|  |  |  | (PRE); Pretoria (-CA), Leendertz 11970 |
|  |  |  | (PRE). |
|  |  | 2529 | (Wi.tbank): Staffberg (-BD), Young Al25 (RRE). |
|  |  | 2627 | (Potchefstroom): Witwatersrand: Frankenwald (-BB), van Rensburg 25556 (J). |
|  |  | 2628 | (Johannesburg): Witwatersrand: Bryanston (-AA), Gilliland 26127 (J). |
| NATAL | - | 2929 | (Underberg): Yorkshire Wolds (-BB), Thode |
|  |  |  | 2538 (STE); Kamberg Mountain (-BD), Wright |
|  |  |  | 1549 (NU). |



Fig. 22. H. multiceps: variation in leaf shape
4. Hypoxis multiceps Buchinger ex Krauss in Flora 28:311 (1845), et in Krauss Breitrage:163 (1846); Baker in J. Linn. Soc.: 117 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:233 (1893), et in Fl. Cap. 6:187 (1896); Medley Wood, Fl. Natal:132 (1907); Nel in Engl. Jahrb. 51:319 (1914); Bews, Fl. Natal \& Zululand:65 (1921); Guillarmod, F1. Lesotho:149 (1971); Ross, F1. Natal:132 (1972). Type: South Africa, Natal, Krauss 248 (BM, holo.!)

Corm l,5-4,1 cm in diam., globose-oblong, crowned with bristles. Leaves 4-8, oblong-1anceolate, $3,0-22,0 \mathrm{~cm}$ long, up to $5,5 \mathrm{~cm}$ broad at the widest point, 3 -ranked, rigid, with $30-40$ ribs, hairs on both surfaces of the lamina stellate (all arms $\pm$ equal in length, $0,4-2,0$ min long) or stellate and bifurcate intermingled (stellate always more numerous than bifurcate) keel and margins with stellate hairs only. Peduncles 1-3(-4), ancipitous, villous, hairs stellate (all arms $\pm$ equal in length). Inflorescence 1-4-flowered. Pedicels 1,5-2,6 cm Iong at time of flower opening, villous. Bracts linear-subulate, setaceous, $0,6-1,9 \mathrm{~cm}$ long. Perianth segments in open flowers $1,0-$ $2,0 \mathrm{~cm}$ long, yellow adaxially, villous abaxially, hairs atellate (one $\operatorname{arm} 1,2-2,0 \mathrm{~mm}$ long, the remainder $\pm 0,4 \mathrm{~mm}$ ). Filaments $\pm$ equal, subulate. Anthers 4,5-6,0 man long, linear-lanceolate, sagittate, apex entire. Style $\pm 1,0 \mathrm{~mm}$ long or stigma subsessile. Stigma concrete, 2,0-3,0 um long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: July-November.
H. costata is the only ally of H. multiceps. No other
species in Natal is likely to be confused with either taxon. Like H. costata, H. multiceps has short, oblong-lanceolate to long, narrow lanceolate leaves (Fig. 22) with the laminae varying from glabrous to


Fig. 23. H. multiceps and H. costata: hair types $\left.\begin{array}{l}\text { A, B, C }- \text { H. multiceps } \\ D, E, F-H \text { H. costata }\end{array}\right]$
densely hairy. The inflorescence is two or three flowered, supported by a particularly thick, ancipitous peduncle. There are, however, several features (see Table 3) that consistently differentiate these closely allied species.

|  | H. costata | H. multiceps |
| :---: | :---: | :---: |
| lamina | Submarginal veins thickened. | Submarginal veins not thickened. |
|  | Hairs predominantly bifurcate (arms up to 6,0 am | Hairs predominantly <br> stellate (arms of equal |
|  | long, Fig. 23C), inter- | length, not exceeding |
|  | mingled with stellate hairs | 2,0 um, Fig. 23D, E), |
|  | (one or two of the arms | occasionally inter- |
|  | long, up to $6,0 \mathrm{~mm}$, | mingled with bifurcate |
|  | Fig. 23A, B) . | hairs (arms up to 2,0 |
|  |  | mm long) |
| keel \&margins | Hairs predominantly | Hairs stellate only <br> (type as above) |
|  | stellate occasionally |  |
|  | intermingled with |  |
|  | bifurcate ones (types |  |
|  | as above) |  |
| inflorescence | Hairs stellate (one arm | Hairs stellate (one arm slightly longer, up to 2,0 min, Fig. 23F). |
|  | only longer, up to 6,0 |  |
|  | num) |  |

Table 3: Sumary of morphological features by which H, costata and H. multiceps may be distinguished.


Fig. 24. Hypoxis multiceps Buch. - known distribution in Southern Africa up to 1975.

Plants of H. multiceps occur in the Eastern Cape, Natal, the Transvaal, Swaziland and Lesotho (Fig. 24), up to altitudes of 2000 m .

$32^{2}$

Fig. 25. Hypoxis costata Bak. - known distribution in Southern Africa up to 1975.
5. Hypoxis costata Baker in J. Linn. Soc.:119 (1878), in Dur. \& Schinz, Consp. F1. Afr. 5:231 (1893), et in F1. Cap. 6: 188 (1896); Nel in Engl. Jahrb. 51:322 (1914); Bews, Fl. Natal \& Zululand:65 (I921); Guillarmod, F1. Lesotho: 149 (1971). Type: South Africa, Orange Free State, Nelson's Kop, Cooper 879 (K, holo.)

Corm l,0-3,5 cm in diam., globose-oblong, crowned with bristles. Leaves 4-10, oblong-lanceolate, $5,0-18,0 \mathrm{~cm}$ long, $0,8-4,2 \mathrm{~cm}$ wide at the broadest point, 3-ranked, usually rigid with $20-70$ ribs and the submarginal veins strongly thickened, hairs on margins and keel stellate with one arm longer than the remainder (up to 6,0 um long), both sides of the lamina glabrous to densely hairy, hairs predominantly bifurcate with some stellate intermingled. Peduncles ancipitous, villous, hairs patent, stellate with one arm longer than the remainder. Inflorescence 1-5 flowered. Pedicels $0,3-1,0(-1,6) \mathrm{cm}$ long at time of flowering, villous. Bracts subulate, setaceous $0,7-1,6 \mathrm{~cm}$ long. Perianth segments yellow adaxially, in open flowers $1,1-1,7 \mathrm{~cm}$ long. Filaments $\pm$ equal, subulate. Anthers (3,0-)4,5-6,0 mm long, sagittate, lanceolate, apex entixe. Stima concrete, 2,0-4,0 mun long, pyramidal with 3 concave faces, usually subsessile. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: October-January.

Hypoxis costata has a distribution sympatric with that of its closest ally H. multiceps (Fig. 25).

Like H. multiceps, plants of H. costata have strongly ribbed, oblong-lanceolate leaves, but in the latter species a thickened submarginal vein is present (Fig. 26). H. costata and H. multiceps are also distinguished according to hair type (for detalls see under the


Fig. 26. H. costata: variation in leaf shape
latter species). Peduncles are ancipitous and the pedicels are generally shorter than those of H. multiceps.

Within the limits of H . costata plants are not entirely uniform: one entity possesses short, oblong leaves with few hairs on the lamina; the other has longer, lanceolate leaves with a densely hairy lamina. However, these are extremes and are linked by intermediates that cannot clearly be placed with either group. Further field work is required to study populations in the field so that the range of variation may be better under stood in relation to habitat and environmental conditions. At the present time the extremes will not be formally recognized.

Citations:

| TRANSVAAL | - | 2430 | (Pilgrims Rest): Mount Anderson, Pilgrims Rest (-DD), Smuts \& Gillet 2465 (PRE). |
| :---: | :---: | :---: | :---: |
| O.F.S. | - | 2828 | (Bethlehem): Golden Gate National Park (-CC), Liebenberg 6956 (PRE). |
|  |  | 2829 | (Harrismith): Rensburgskop, Swinburne (-AC), Jacobsz 328 (PRE). |
| SWAZILAND | - | 2631 | (Mbabane): Ukutula, Mbabane (-AC), Compton 24490, 24578, 25210 (NBG). |
| NATAL | $\sim$ | 2730 | (Vryheid): Hlobane, Vryheid (-DB), Johnstone 528 (NU). |
|  |  | 2829 | (Harrismith): Catchment 9, Cathedral Peak (-CC), S.E. Wood 155, 158 (NU), Killick 1573 (NPC). |
|  |  | 2929 | (Underberg): Impendh1e (-DB), Levett 101 ( NH ) . |
| LESOTHO | - | 2828 | (Bethlehem): Leribe (-CC), Dieterlen 229b ( $\mathrm{NH}, \mathrm{PRE}, \mathrm{SAM}$ ). |
| CAPE | - | 3226 | (Fort Beaufort): Katberg (-DA), Galpin 8427 (PRE). |
|  |  | 3326 | (Grahamstown): Grahaustown (-BC), MacOwan 104 (SAM). |
| without precise locality: |  |  |  |
|  |  |  | Compton 21.311 (NBG), Basutoland; Thode 2543 (STE), Altemooi, Natal Province. |

6. Hypoxis rigidula Baker in J. Linn. Soc. 17:116 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:233 (1893), et in Fl. Cap. 6:186 (1896); Medley Wood, F1. Natal:132 (1907); Ne1 in Eng1. Jahrb. 51:331 (1914); Bews, F1. Natal \& Zululand: 65 (1921); S'blch in Merxm. F.S.W.A. Fam. 151:2 (1969); Guillarmod, F1. Lesotho:149 (1971); Ross, Fl. Natal:132 (1972). Type: South Africa, Cape, British Kaffrarig, Cooper 3239 (K, syn.!)

Corm 2,0-5,0 cm in diam., oblong, crowned with bristles.
Leaves $4-6(-9), 9,0->100,0 \mathrm{~cm}$ long, $0,3-1,3 \mathrm{~cm}$ at widest point, 1 inearlanceolate, erect, rigid, not 3-ranked, ribs 6-20(-32), close together, irregular: both surfaces of the lamina woolly villous, or white appressed hairy (in the latter case the adaxial surface with stellate, usually the more numerous, and bifurcate hairs intermingled in the channels between the ribs); abaxial surface more densely hairy (keel and margins with stellate hairs only). Peduncles 1-5, ancipitous, densely hairy below inflorescence grading to sparsely pilose towards the base. Inflorescence (excluding peduncle) up to $13,5 \mathrm{~cm}$ long, (2-)3-11 flowered (the first produced in each season tending to be limited to 2 flowers only). Pedicels up to $1,0 \mathrm{~cm}$ long at time of flowering or flowers subsessile, densely hairy. Bracts linearsubulate, setaceous, $0,7-2,7$ cm long. Perianth segments yellow adaxially, in open flowers ( $0,7-$ ) 1, $0-2,1 \mathrm{~cm}$ long. Filaments equal, subulate. Anthers 4,5-7,5 mm long, lanceolate, deeply sagittate, apex entire. Style cylindrical, 1,0-1,5 mm long. Stigma concrete, $2,5-3,5 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehtscence below the apex.

Flowering Period: September-March.

Within the limits of H. rigidula Baker recognized two entities, typical H. rigidula and II. rigidula var. pilosissima. These two taxa


Fig, 27. Hypoxis rigidula Bak. var. rigidula - known distribution in Southern Africa up to 1975, excluding South West Africa from which one gathering, Dinter 5601, has been recorded
are retained for they differ according to the hair covering of the leaf. The leaves of var. rigidula are covered with stellate and bifurcate hairs which lie in channels between the ribs; leaves of var. pilosissima have a much denser covering of stellate hairs only, which are patent, not appressed, and thus produce a woolly effect.

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Hypoxis rigidula var. rigidula
    Baker in J. Linn. Soc. 17:116 (1878), in Dur. & Schinz,
    Consp. F1. Afr. 5:233 (1893), et in F1. Cap. 6:186 (1896);
    Medley Nood, Fl. Natal:132 (1907); Nel in Engl. Jahrb.
    51:331 (1914); Bews, F1. Natal & Zululand:65 (1921);
    Sb1ch in Mermx. F.S.W.A. Fam. 151:2 (1969); Guillarmod,
    F1. Lesotho:149 (1971); Ross, F1. Natal:132 (1972).
    Type: South Africa, Cape, British Kaffraria, Cooper 3239
    (K, syn.!)
    H. elliptica Nel in Engl. Jahrb. 51:332 (1914); Bews, F1.
    Natal & Zululand:65 (1921); Ross, Fl. Natal:132 (1972).
    Type:' South Africa, Natal, betwecn Pietermaritzburg and
    Greytowa; Wilms 2317 (K, syn.:)
```

Leaves both surfaces of the lamina white, appressed hairy; adaxial surface with stellate (usually the more numerous) and bifurcate hairs intermingled in the channels between the ribs; abaxial surface more densely hairy (keel and margins with stellate hairs only).

Plants of var. rigidula are widely distributed in all four provinces of South Africa, Lesotho, Swaziland and South West Africa. The southern-most record at the present time is from East London (Fig. 27).

Examination of the syntype of H. elliptica Nel, showed it to to be conspecific with $H$. rigidula var. rigidula, and therefore it must now become a junior synonym.


Acocks 10833 (NH).
2830 (Dundee): top of Mpati M.t. (-AA), Shirley 76 (NU); Culvers, Weenen (-CC), Rogers 27786 (STE).
2831 (Nkandla): Eshowe (-DC), Lawn 1297, 1788 (NH).
2929 (Underberg): Giants Castle Game Reserve (-AD), Trauseld 661, 866, 867, (NU); Kamberg "Game Pass" (-AD), C.G. Gordon-Gray 117 (NU); near Gourton (-BB), Med1cy Wood 2627, 3456 (NH); hills above Mooi River (-BB), Medley Wood 3870 ( NH ) ; Mooi River Meteor Ridge ( -BB ), Mogg 3240 (PRE); nr. Willowford Stn., Estcourt (-BB), Acocks 10604 (NH); 85 km to Nottingham Rd. from Himeville ( -DA ), S.E. Wood 128 (NU); 75 km to Nottingham Rd. (-DA), S.E. Wood 128A (NU); 33 km to Underberg.from Pietermaritzburg ( -DC ), S.E. Wood 124 (NU).
2930 (Pietermaritzburg): Howick (-AC), Moll 1255 (NH), 1040 (NU); 8 m . from Greytown ( -BA ), Wylie s.n. (NH 28016); Greytown (-BA), Wylie s.n. (NH 21681); 1 km to New Hanover Station from Pietermaritzburg (-BA), S.E. Wood 92 (NU); 18 km to Greytown, nr. Seven Oaks (-BA), S.E. Wood 94 (NU); 20 km to Kranskop (-BA), S.E.Wood 95 (NU); 2 km from Estcourt (-BB), Stirton 1045 (NU); Pietermaritzburg (-CB), Allsopp 920 ( $\mathrm{NH}, \mathrm{NU}$ ) ; Chase Valley, Pietermaritzburg (-CB), S.E. Wood 140 (NU); Howick old main road, Pietermaritzburg (-CB), S.E. Wood 143 (NU); Arnold's Hill, Richmond (-CD), Wylie s.n. (NH 23357); 3 kn to Thornville on road to Richmond (-CD), Stirton 1219 (NU); 11 km to Mid-Illovo (-CD), S.E.Weod 106, 107 (NU); 8 ka to Mid-Illovo along railway line (-CD), S.E. Wood 114 (NU); Fields Hill near Pinetown (-DD), Medley Wood 754 (SAM).
3030 (Port Shepstone): 8 kin from Highflats towards Umzinto (-AB), S.E. Wood 175 (NU); Umgaye Flat (Alexandra County) (-BA), Rudatis 472 (STE); on road to P. Edward from Izingolveni (-CC), S.E. Wood 164A (NU).
LESOTHO - 2828 (Bethlehem): Leribe (-CC), Dieterlen 223,

CAPE - 3028 (Matatiele): Mt. Fletcher (ـCD), Weidermann \& Oberdieck 1147 (PRE).
3029 (Kokstad): Kokstad (-CB), Mi1dred 2 (NBG), Tyson 1211 (SAM), Piek 89 (NH).
3126 (Queenstown): Hangklip Mt., Queenstown (-DD), Galpin 1604 (GRA, PRE).
3128 (Umtata): nr. Ugie (-AA), Flanagan 2842 (SAM).
3226 (Fort Beaufort): Hogsback, Amatola Mts. (-DB), Peacock s.n. (SAM 65759).
3227 (Stutterheim): Mount Coke (-CD), Sim 1125
(NU); Stutterheim (-DA), Theron 2115 (NH);
Komgha (-DB), Flanagan 1171, (SAM).


Fig. 28. Hypoxis rigidula Bak. var. pilosissima Bak. known distribution in Southern Africa up to 1975.

3327 (Peddie): East London (-BB), Barker 7952 (NBG). without precise locality:

Gerstner s.n. (NH 28986), Zululand; MacOwan 1649 (GRA), Boschberg; Acocks 16275 (PRE), 4 m. s. of Groeneberg Pass, Adelaide; Lesotho: Dieterlen 1370 (NBG, PRE), near Joa's village, Likhoele, Mafeteng Distr.

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Hypoxis rigidula var. pilosissima Baker in J. Linn. Soc. 17:
    116 (1878), in Dur. & Schinz, Consp. Fl. Afr. 5:233 (1893),
    et in Fl. Cap. 6:186 (1896); Medley Wood, F1. Natal:132
    (1907); Nel in Engl. Jahrb. 51:331 (1914); Bews, Fl.
    Natal & Zululand:65 (1921); Ross, F1. Natal:132 (1972).
    Type: South Africa, Natal, Tugela, Gerrard 1826 (K, syn.!)
```

Leaves both surfaces of the lamina woolly villous, hairs not appressed.

The known distributional limits of var. pilosissima are similar to those of var. rigidula, but there are not as many collections and the southern-most limit, at present, is the Kei River Mouth (Fig. 28).

Of special interest is Moss 12080 (J), Houghton Rise, Johannesburg. The plant is particularly robust with leaves of $2,0 \mathrm{~cm}$ in width and a 13-flowered inflorescence. It would be interesting to determine whether polyploids are represented in the locality where this plant was collected.

Plants of both varieties of $H$. rigidula are tall, rigid and erect. The leaves are not 3-ranked and remain close together, which characters, later in the season when growth is complete, impart an impression of "whip-like" form. The first infloreseence is usually 2-flowered, a phenomenon which can cause confusion in distinguishing plants of this species from those of H. acuminata (for details see under this latter species). Subsequent inflorescences, however, are
generally more than 2-flowered. Plants occur mainly in grassland, but are occasionally recorded from damper areas such as vlef margins.



Fig. 29. Hypoxis acuminata Bak. - known distribution in Southern Africa up to 1975.
7. Hypoxis acuminata Baker in J. Bot. (Lond.):3 (1889), in Dur. \& Schinz, Consp. Fl. Afr. 5:230 (1893), et in Fl. Cap. 6: 186 (1896); Medley Wood, Fl. Natal:132 (1907); NeI in Eng1. Jahrb. 51:318 (1914); Guillarmod, Fl. Lesotho:148 (1971); Ross, F1. Natal:132 (1972). Type: South Africa, Natal, Inanda, Medley Wood 1347 ( NH , holo.!)

Corm 1,7-2,5 cm in diam., globose-oblong, crowned with bristles. Leaves (4-)5-9, up to $30,0 \mathrm{~cm}$ long, $0,3-1,2 \mathrm{~cm}$ at widest point, not noticeably 3-ranked, linear-lanceolate or linear-acuminate, semi-erect, V-shaped in cross section, ribs 6-12, hairs stellate and bifurcate intermingled, opaque, patent, sparse on adaxial surface, dense on abaxial with hairs to keel and margins often U-shaped. Peduncles ancipitous, slightly grooved, covered with patent hairs. Inflorescence $2-4(-6)$ flowered. Pedicels $0,2-1,4 \mathrm{~cm}$ long at time of flower opening, all approximately equal or that of first formed flower longer. Bracts. subulate, setaceous, $0,6-1,6 \mathrm{~cm}$ long. Perianth segments yellow adaxially, in open flowers $0,9-1,7 \mathrm{~cm}$ long. Filaments subulate. Anthers $4,0-6,0(-7,0) \mathrm{mm}$ long, those associated with outer perianth segments occasionally 0,5-1,0 mm longer than the inner ones, lanceolate, sagittate, apex entire or slightly bifurcate. Style 1,0-2,0 malong, or stigma subsessile. Stigma concrete, $2,0-3,5 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a capsule opening by circunscissile dehiscence below the apex.

Flowering Period: September-May.

Hypoxis acuminata is predominantly a grassveld species with a wide altitudinal range. Its distribution is sympatric with that of H. rigidula (its closest morphological ally), and the present southernmost limit is Weza, Natal (Fig. 29).

Both H. rigidula and H. acuminata have erect, narrow leaves but the latter species has a smaller, less rigid, fewer-veined leaf that is V-shaped in cross section (the lamina of H. rigidula is flat). Also leaves of H. rigidula remain "bunched" together while those of H. acuminata spread out.

The type and position of hairs on the leaf also differ in the
two taxa. H. acuminata has non-localised, predominantly bifurcate, patent hairs, whereas $\underline{H .}$ rigidula has appressed, predominantly stellate hairs which lie in channels between the ribs of the lamina.

Inflorescences differ only in flower number: H. acuminata usually has two, but may have up to four flowers, whereas inflorescences of H. rigidula are 3-11 flowered, except for the first which is usually 2 flowered.


8. Hypoxis oblonga Ne1 in Engl. Jahrb. 51:332 (1914); Bews, Fl. Natal \& Zululand:65 (1921); Ross, Fl. Natal:132 (1972). Type: South Africa, Natal, Weenen Distr., Medley Wood 4372 (NH, iso.!)

Corm 2,0-3,0 cm in diam., oblong, crowned with bristles.
Leaves $4-7,12,0-80,0 \mathrm{~cm}$ long, $0,2-1,5 \mathrm{~cm}$ at widest point, linear acuminate-lanceolate, erect, not $3-r a n k e d, 8-20$ irregular ribs with 2 more prominent, hairs on margins and keel stellate (predominant) and bifurcate intermingled, up to $8,0 \mathrm{~mm}$ long, whitish, dense; hairs on lamina bifurcale only, scattered to dense (abaxial surface denser


Fig. 30. Hypexis oblonga Nel - known distribution
in Southern Africa up to 1975.
than adaxial). Peduncles ancipitous, patently hairy, becoming subglabrous towards the base. Inflorescence 2-6 flowered. Pedicels $2,0-5,0 \mathrm{~mm}$ long or flowers subsessile. Bracts subulate, setose, 1,1-2,2 cm long. Perianth segments yellow adaxially, in open flowers $1,0-1,7 \mathrm{~cm}$ long. Filaments subulate, equal. Anthers 4,0-7,0 man long, lanceolate, sagittate, apex entire. Style cylindrical, 1,0-2,0 uma long. Stigma concrete, $2,0-3,0$ um long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: June-December.

Hypoxis oblonga is known only from Natal (Fig. 30). The erect, essentially narrow leaves are variable in width ( $0,2-1,5 \mathrm{~cm}$ ) and hairiness within the limits of a population. The leaf blades are provided with a prominent rib on either side of the keel. The leaves are not 3ranked. Plants growing at Inchanga exhibited aspect differences: those from the south facing slope had narrower, less densely hairy leaves; those from the north facing slope were wider, harder and more densely hairy.
H. oblonga has two morphological allies, H. galpinij and H. rigidula var. rigidula. Their differences are summarised in Table 4. In all other respects they are similar.

Citations:
NATAL - 2828 (Bethlehem): Mont aux Sources (-DD), Pellatt 2 (NU).
2929 (Underberg): "South Downa", Weenen Distr.
(Estcourt) (-BB), Medley Wood 4372 (NH);
 (STE).
2930 (Pietermaritzburg): Inchanga Cutting (-DA), K.D. Gordon-Gray 10011 (NU), S.E. Wood 210
(NU): Botha's Hill (-DC), Medley Wood 2435
(NH): Westville (-DD), Cairns s.n. (NU, 8.8.55).

| Leaf | H. oblonga | H. galpinii | H. rigidula |
| :---: | :---: | :---: | :---: |
| width | 0,2-1,5 cm | - $1,5-4,5 \mathrm{~cm}$ | 0,3-1,3 cm |
| ribs | prominent rib on either side of keel | prominent rib on either side of keel | ribs irregular |
| lamina | usually conduplicate | not condup1icate | not conduplicate |
| laminal hairs | bifurcate, | absent, ox | stellate |
|  | scattered to | rarely stellate | (predominent) |
|  | dense on both | hairs, scattered | and bifurcate |
|  | surfaces | on prominent | intermingled, |
|  |  | ribs | 1ying in |
|  |  |  | channels |
|  |  | - | between the |
|  |  |  | ribs |

Table 4. Compaxison of leaf characters of H. oblonga, H. galpini.i and H . rigidula var. rigidula.


Fig. 31. Hypoxis obtusa Burch. - known distribution in
Southern Africa up to 1975, excluding South West Africa and Botswana
9. Hypaxis obtusa Burchell in Bot. Reg.t. 159 (1816); Roem. et Schultes, Syst. Veg. 7:765 (1818); Baker in Trans. Linn. Soc. 29:156 (1875), in J. Linn. Soc.:114 (1878), in Dur. \& Schinz, Consp. F1, Afr. 5:233 (1893), et in F1. Cap. 6:154 (1896); Medley Wood, F1. Natal:132 (1907); Nel in Engl. Jahrb. 51:334 (1914); Bews, F1. Natal \& Zululand: 65 (1921); Söich in Merxm. F.S.W.A. Fam. 151:2 (1969); Guillarmod, F1. Lesotho:149 (1971); Ross, F1. Natal:132 (1972). Type: South Africa, Bechuanaland, Pellat Plains near Takun, Burche11 s.n. ( $K$, holo.!)

Hypoxis nitida Verdoorn in Flow. Pl. Afr. 27:Plate 1058; (1949); Guillarmod, F1. Lesotho:149 (1971); Ross, Fl. Natal:132 (1972). Type: South Africa, Transvaal, Pretoria, Robertson 2 (PRE, holo.!)

Corm 4,0-7,0 cmin diam., oblong-globose, crowned with a dense ring of bristles. Leaves $6-16,7,0-56,0 \mathrm{~cm}$ long, $0,3-2,0 \mathrm{~cm}$ at widest point, linear-lanceolate, erect but becoming twisted and curving backwards with age, ribs $20-80$, prominent, usually regular in size, hairs white, opaque, stellate, usually appressed on margins and keel, lamina usually glabrous, occasionally with stellate hairs at the base, with or without bifurcate hairs intermingled. Peduncles (1-)3-5, ancipitous, densely hairy below inflorescence grading to subglabrous or glabrous at the grooved base. Inflorescence 2-12 flowered. Pedicels lowest $2-4$ at time of flower opening, $0,5-1,7 \mathrm{~cm}$ long, densely hairy. Bracts subulate, setose, $0,8-3,5 \mathrm{~cm}$ long, the first formed flowers with the longest bracts. Perianth segments yellow adaxially, in open flowers $1,2-2,3 \mathrm{~cm}$ long. Filaments subulate, equal. Anthers 6,0-8,0 0 m long, lanceolate, deeply sagittate, apex entire. Style cylindrical $1,0-2,5 \mathrm{~mm}$ long. Stigma concrete, $2,0-4,0 \operatorname{man}$ long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.


Fig. 32. H. obtusa: a - young leaves, b - last season's leaves


Fig. 33. H. obtusa with old leaves only

Mypoxis obtusa is southern African in distribution (Fig. 31), being recorded from South West Africa, Swaziland, Lesotho and all four provinces of South Africa. Populations commonly occur at high altitudes of up to approximately 2000 m , but at more southern latitudes they are present at lower elevations often not far from the coast.

Leaves of this species are lanceolate and strongly ribbed, with the younger, central leaves characteristically narrower than the older, outer ones. The keel and margins are clothed with stellate hairs, hut the lamina is usually glabrous (occasionally stellate or bifurcate hairs, or both types intermingled, occur over the lamina for a distance of about $4,0 \mathrm{~cm}$ from the base). During the growing season the leaves elongate to up to 60 cm , become recurved and twisted and are not as markedly conduplicate as are the younger leaves. Fig. 32 shows the difference between young and old leaves, while Fig. 33 shows a plant with old leaves only.

Specimens v. Rensburici s.n. (J 25557) and Godfrey s.n. (PRE 53555)(Fig. 31-1 \& 3) are thought to be variants of H . obtusa because the leaves are covered throughout in stellate hairs. Further collectIng in the central area of the Transvaal, from where these plants were collected, is needed to determine if a range from the glabrous to the hairy form is present, or whether the hairy plants should be recognized as a distinct variety.

Verdoorn in 1949 established H. nitida (type: Robertson 2, Pretoria (PRE). In the type description this species was said to have "leaves 12 to 15 at time of flowering, shining, glabrous, or occasionally with a patch of appressed pubescence, the margins and midrib conspicuously fringed with whitish pubescence, sub-equally 30-75 nerved, at first erect then recurving, trifarious ... at the
time of flowering, 18 to 23 cm long and 1 to 2 cm broad, the outer shorter and broader and the innermost narrower". In the discussion comparison was made with H. obtusa. "In respect of the white fringes to the leaf it $\overline{\mathrm{H}}$. nitidā resembles $\underline{H .}$ obtusa, but in that species the leaves are much narrower, with fewer nerves, and they are not arranged trifariously".

According to the characters used by Verdoorn, I have not been able to distinguish $\underline{H}$. Jitida from $H$. obtusa. The critical characters do not hold, firstly because the leaves of both species are not always obviously trifarious, and secondly because leaf width varies between extremes of 0,3 and $2,0 \mathrm{~cm}$, with a wider leaf possessing more nerves than a narrower one.

Comparison of the types has shown these to be conspecific, and H. nitida must now become a junior synonym of H . obtusa.

The only close ally of H . obtusa is H . rooperi S . Moore, but in the latter species the leaves are more markedly 3-ranked, wider, hairy all over with the ribbing not as compact, and not becoming twisted in age.

Thinges 1904 (Fig. 31-2) falls into this complex but has hairy leaves like those of H . rooperi. A concentration of hairs on the keel and margins is, at the same time, suggestive of similar aggrégations of hairs on the leaves of H. obtusa. Thinges 1904 cannot be confused with either Godfrev s.n. (PRE 53555) or v. Rensburg s.n. (J 25557), as its leaf hairs are not appressed, and are stellate and bifurcste. Field study in the central Transvaal is necessary if the relationships among these variants are to be understood.

Citations:

| TRANSVAAL |  | 2428 | (Nylstroom): Flats near Warmbaths (-CC), Hofström \& Acocks 251 (PRE). |
| :---: | :---: | :---: | :---: |
|  |  | 2527 | (Rustenburg): Rustenburg (-AC), Leendertz 10203 (PRE), Moss 2655 (J). |
|  |  | 2528 | (Pretoria): Pretoria (-CA), Pegel s.n. (NH, 23.1.68, 4 sheets), Robertson 2 (PRE); |
|  |  |  | Muckleneuk, Pretoria (-CA), Bouwer 6055 (PRE); |
|  |  |  | Doornpoort, Pretoria (-CA), Mogg 272 (PRE); |
|  |  |  | Axalikloof, Pretoria (-CA), Mogg s.n. (PRE 9358); Doornkloof, Irene (-CC), Gillett s.n. (STE 30403). |
|  |  | 2529 | (Witbank): Buffelsvlei, Middleburg, Transvaal (-CB), Rudatis 39 (STE); Middleburg (-CB), |
|  |  |  | Jenkins 10202 (PRE). |
|  |  | 2530 | (Lydenburg): 18 mls from Nelspruit, Johannesburg road (-BC), Buitendag 619 (NBG). |
|  |  | 2531 | (Komatipoort): Barberton (-CC), Galpin 412 (GRA, PRE). |
|  |  | 2627 | (Potchefstroom): Witwatersrand, Frankenwald <br> (-BA), V. Rensburg s.n. ( $J$ 25556, 25557), |
|  |  |  | Maguire s.n. (J 25555); Parys (-CD), Godfrey s.n. (PRE 33555); Vereeniging (-DB), Gunn s.n. (PRE 33557). |
|  |  | 2628 | (Johannesburg): Johannesburg (-AA), Moss 2556 |
|  |  |  | (J), Pege1 s.n. ( NH 23.1.68); The Wilds, Johennesburg (-AA) Lambert s.n. (J 24471); |
|  |  |  | Melville Koppies (-AA), Macnae 1496 (J); Zoo |
|  |  |  | Koppies, Johannesburg (-AA), Moss 16184 (J); |
|  |  |  | Kempton Park (-AA), Prosser s.n. (J 29380); |
|  |  |  | 5 km past Balfour on road to Volksrust (-DA), Arnold 836 (NU). |
|  |  | 2630 | (Carolina): Piet Retief (-DD), Compton 22360 (NBG). |
| O.F.S. | - | 2626 | (Klerksdorp): Lichtenburg (-AA), Thinges 1904 (PRE). |
|  |  | 2828 | (Bethlehem): Beth1ehem ( -AD ), Potgieter 34 |
|  |  |  | (PRE); Golden Gate National Park (-DB), Liebenberg 7329 (PRE). |
|  |  | 2829 | (Harrismith): Harrismith (-AC), v.d. Zeyde |
|  |  |  | 8.n. (NBG, 4.2.70, 12.10.70). |
| SWAZILAND | - | 2631 | (Mbabane): Manzini (-AD), I'ons 63/19 (NBG). |
| NATAL | - | 2729 | (Volksrust): 20 km before Newcastle from |
|  |  |  | Volksrust (-DB), Arnold 842 (NU); Newcastle (-DD), Booysen 1707 (NGG). |
|  |  | 2730 | (Vryheid): Kafir Drift (-CB), Thode A249 |
|  |  |  | (NH) ; Vryheid (-DD), Strey 9101 (NH, PRE). |
|  |  | 2731 | (Louwsburg): Nongoma (-DC), Markotter s.n. (STE 8851). |
|  |  | 2829 | (Harrismith): Oliviershoek Pass, Bergville (-DA), Laughton s.n. (J 34644). |
|  |  | 2830 | (Dundee): 30 km from Dundee to Vryheid (-AB), |
|  |  |  | Stirton 1317 (NU); Weenen (-CC), Rogers 28311 (STE). |


| LESOTHO |  | 2929 | (Underberg): Giants Castle (-AD), Symons 72 (PRE), Trauscld 868 (NU); Mooi River, Meteor Ridge ( -BB ), Mogg 3281 (PRE, 2 sheets). |
| :---: | :---: | :---: | :---: |
|  | - | 2828 | (Bethlehem): Leribc (-CC), Dieterlen 223 ( NH ), |
|  |  |  | 230 (PRE, SAM), 310 (SAM). |
| CAPE | - | 2624 | (Vryburg): Vryburg (-DC), Sharpe s.n. (PRE 7234), no name (STE 12805). |
|  |  | 3026 | (Aliwal North): Aliwal North (-DA), Bolus 181 (PRE, STE). |
|  |  | 3029 | (Kokstad): Kokstad (-CB), Mildred I (NBG). |
|  |  | 3126 | (Queenstown): nr. Molteno (-AD), Flanagan 1642 (SAM); Hangk1ip Mtn., Queenstown (-DD), Galpin |
|  |  |  | 1605 (PRE); Queenstown (-DD), Baur 904 (SAM). |
|  |  | 3227 | (Stutterheim): Cathcart (-AC), Kemp s.n. (NBG 65826); Welcome Wood (King Williams Town) (-CD), Sim Il26 (NU). |
| without precise locality: |  |  |  |
|  |  |  | MacOwan 1649 (SAM), Boschberg, Somer |
|  |  |  | Moss s.n. (J, 22.10.2.7); Rennie 610 (NJ), |
|  |  |  | Polela distr., Glengariff; Weeks 25 (J) |
|  |  |  | Eckstein Park; Winter 617 (NBG). |

(Note: The South West African specimens have not been cited since it has not yet been possible to examine them.)
10. Hypoxis rooperi s. Moore in Gard. Comp. I:65 cum icone (1852), in J. Linn. Soc. 17:118 (1878), in Dur. \& Schinz, Consp. F1. Afr. 5:234 (1893), et in F1. Cap. 6:188 (1896); Medley Wood, Fl. Natal:132 (1907); Nel in Engl. Jahrb. 51:337 (1914); Bews, Fl. Natal \& Zululand:65 (1921); Guillarmod, Fl. Lesotho:149 (1971); Ross, Fl. Natal:132 (1972). Type: South Africa, without locality, Cooper 154 (BM, syn.!)

Corm 2,6-6,5 cm in diam., g1obose-oblong, crowned with bristles. Leaves many, 6,0-95,0 cm long, $1,0-5,2 \mathrm{~cm}$ at widest point, 3-ranked, lanceolate, -erect usually sickle-shaped, ribs 20-50, slight1y irregular in size, keel, margins and usually abaxial surface densely covered with stellate and bifurcate, white hairs, adaxial surface less densely covered with similar hairs. Pedunc1es ancipitous, densely hairy below inflorescence, grading to subglabrous at the grooved base.


Fig. 34. Hypoxis rooperi S. Moore - known distribution in Southern Africa up to 1975.

Inflorescence 2-15•flowered, up to $12,0 \mathrm{~cm}$ long (excluding peduncle). Lower pedicels ( $0,5-$ ) $1,0-3,0 \mathrm{~cm}$ long at time of flower opening. Bracts subulate, setose, $0,7-3,3 \mathrm{~cm}$ long. Perianth segments yellow adaxially, in open flowers ( $0,8-$ ) $1,0-2,3 \mathrm{~cm}$ long. Filaments subulate, equal. Anthers ( $5,0-$ ) $7,0-7,5(-8,0) \mathrm{mm}$ long, lanceolate, sagittate, apex usually entire, seldom slightly bifurcate. Style cylindrical, 1,0-3,0 mm long. Stigma concrete, 2,0-3,5 mm long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: (July-) August-January (-May).

Hypoxis rooperi is perhaps the best known Natal species. Plantis are characterized by the many, obviously 3-ranked, sickleshaped leaves that are densely hairy abaxially.

Plants occupy a wide range of habitats from sandy hills on the margins of dune forests to damp areas around streams: they are commonest, however, in grassveld, where, in disturbed areas, quite extensive almost pure stands often occur.
H. rooperi is eastern, southern African in distribution (Fig. 34), being recorded from all four provinces of South Africa', Lesotho and Swaziland, up to altitudes of approximately 1800 m. There is considerable variation in leaf size and hairiness over this range, but no distinct infra-specific entities are recognizable.

There is no difficulty in the identification of this species in the field. H. obtusa is its closest ally, but in this taxon the leaves are not as distinctly 3-ranked as in H. rooperi, while the narrower laminae are glabrous, except for opaque, stellate hairs
confined to the margins and keel.

Citations:

TRANSVAAL - 2431 | (Acornhoek): Kruger National Park (-AB), |
| :--- |
| Codd 5673 (PRE). |

2527 | (Rustenburg): Rustenburg (-AD), Hards 19 |
| :--- | (PRE).

2528 (Pretoria): Pretoria (-CA), Leeman s.r. (PRE 33554, Smith 1724 (PRE).
2529 (Witbank): Zondagsfontein, Witbank (-CC), Thode A2856 (NH).
2627 (Potchefstroom): Mooi River (-CA), Nelson 11768 (PRE); K1ipdrift (-CA), theron 1190 (NH).
2628 (Johannesburg): Modderfontein (-AA), Haagner s.n. (GRA, Oct. 1904); Hills south of Johannesburg (-AA), Arnold 834 (NU).
2630 (Carolina) Piet Retief (-DD), Pienaar 74 (NU).
O.F.S. - 2727 (Kroonstad): Kroonstad Convent (-CA), 6 (PKE).

2827 (Senekal): Senekal (-BD), Goossens 779 (PRE).
2828 (Bethlehem): Bethlehem (-AD), v. Ginkel 257 (PRE).
2926 (Bloemfontein): Bloemfontein (-AA), Bouwer 2229 (PRE).
SWAZILAND - 2631 (Mbabane): Timbutini River Bank (-AB), Compton 31790 (NBG); near Bremersdorp (-AB), Compton 28104 (NBG); Komati Bridge (-AC), Compton 28843 (NBG); Umtintegwa (-BD), Compton 27337 (NBG).
NATAL - 2632 (Bela Vista): Ndumu Game Reserve (-CC), Pooley 662 (NU); NPB camp, Kosi Bay (-DD), Moll \& Strey 3807 (NH).
2729 (Volksrust): 20 km before Newcastle from Volksrust (-DB), Arnold 840 (NU).
2731 (Louwsburg): Louwsburg (-CB), v. Rooyen 5 (NU).
2732 (Ubombo): Lake Sibaya (-BC), Hart 29 (NU);
Lala Nek (-CA), Stephen, v. Graan \& Schwabe 1151 (PRE); Sordwana Bay Park (-DA), S.E. hood 86 (NU).
2829 (Harrismith): 8 mls to Bergville on Rustenburg Rd. (-CA), Edwards 2407 (NU); Hart's Hill, Colenso (-DB), Strey 9962 (NH, NU); Ladysmith (-DB), Geekie 4l (NU); Colenso (-DD), Dixon 6 (NU).
2830 (Dundee): Dundee (-AA), Shirley 241 (NU); Krantzkloof (-DD), Haygarth 18 (STE), Rogers 24612 (STE).
2831 (Stanger): Hluhluwe Game Reserve ( -BB ), Ward 1586 (NAl, NU, PRE); Eshowe (-CD), Lawn 446 (NH), 1161 ( NH ), Gerstner 2590 (NH); Ngoye

2832 (Mtubatuba): Hluhluwe (-AA), We11s 2113 (INA); Hluhluwe Game Reserve (-AA), Bourquin 458 (NU); Richards Bay (-CC), Lawn 335 (NH).

2929 (Underberg): 8 km from Estcourt (-BB), S.E. Wood 145 (NU).
2930 (Pietermaritzburg): Albert Falls (-AD), Comins 304 (NU); $\pm 10 \mathrm{~km}$ from Wartberg (-AD), S.E. Wood 75 (NU); Greytown (-BA), Wylie s.n. (NH 22420); Kingscliff. (-BC), S.E. Wood 77B (NU); Impolweni (-BC), Rump s.n. (NH 20300); Kings Hill Railway siding (-BC), S.E. Wood 76 (NU); Pietermaritzburg (-CB), Lachman 7 (NU), Mauve 29 (NU), Fisher 81 (NU), Fairall 110 (SAM), Barker 4367 (NBG), Leisgang 46 (NU), Douwes Dekker 11 (NU), Pienaar 16 (NU), Stixton 105 (NU), S.E. Wood 72, 82 (NU); 11 km to MidIllovo from turn off on Eston Rd. (-CD), S.E. Wood 106 (NU); Nagel Dam (-DA), Todd 33 (NU); Cato Ridge (-DA), Ross 2157 (NH); Camperdown (-DA), Wells 1578 (NU); Inanda (-DB), Strey 5163 (NH), Medley Wood 47 (SAM); Bothas Hill (-DC), Hutchinson, Forbes \& McClean 31 ( NH ); 8 km to Kingburugh on Eston Rd. (-DC), S.E. Wood 118 (NU); 36 kra to Umbumbulu on Eston Rd. ( -DC ), S.E. Wood 100 (NU); 32 km to Umbumbulu ( -DC ), S.E. Wood 102 (NU); Isipingo North (-DD), Ward 398,459 (NU); Cowies Hill (-DD), Lawson 1220 (NH); Umlazi, Adams Mission station (-DD), Wilker 116 (NU).
2931 (Stanger): Umhlanga River (-CA), Ross \& Moll 2281 (NH, PRE); Tongaat (-CA), Farquaharson 3 (NU); 5 ml s west of Verulam ( -CA ), Moll 2068 (NU); Brighton Beach (-CC), Coleman 159 (NH); Greenwood Park (-CC), Wylie s.n. (NH 23107); Durban Flat (-CC), Medley Wood s.n. (NH 6922); Durban (-CC), Forbes s.n. (STE 12803).
3029 (Kokstad): Clydesdale (-BB), Tyson 2745 (PRE); Kokstad (-CB), Tyson 1125, 1210 (SAM), 1478 (GRA).
3030 (Port Shepstone): Hlutankungu (-AD), S.E. Wood 79 (NU); Umdoni Park (-BC), Guy \& Jarman 84 (NU); on rd. from Izingolweni to Port Edward (-CG), S.E. Wood 165B (NU); Beach Terminus (-CD), Thode s.m. (STE 2540); Ramsgate (-CD), S.E. Wood 174 (NU); Greenhart turn off from Port Edward to Port Shepstone (-CD), S.E. Wood 173 (NU); Marina Beach (-CD), Strey 5942 (NH, NU); St. Michaels-on-Sea (-CD), Nicholson 238 (NH); Shelley Beach (-CD), Strey 9256 (NH).
3130 (Port Edward): Entrance into Port Edward from Izingolweni ( -AA ), S.E. Wood 167, 168 (NU); outside Port Edward (-AA), S.E. Wood 170 (NU). Port Edward (-AA), S.E. Wood 171 (NU).

| LeSOTHO | - | 2828 | (Bethlehem): Leribe (-CC), Dieterlen 229A |
| :---: | :---: | :---: | :---: |
|  |  |  | (NH, PRE, SAM, 2 sheets), Dieterlen 229 (SAM). |
| CAPE | - | 3128 | (Umtata): Ugie (-AA), Britten 4653 (GRA); |
|  |  |  | Umtata waterfall (-DA), Schonland 3925 (GRA) |
|  |  |  | The Haven (-DC), J.L. Gordon-Gray 1438 (NU). |
|  |  | 3129 | (Port St. Johns): Port St. Johns (-DA), |
|  |  |  | Schonland 4179 (GRA). |


| 3227 | (Stutterheim): Mount Coke (-CD), L.E. Taylor 3535 (NBG), King Williams Town (-CD), Sim <br> 1129 (NU, SAM), 1855 (NU), Batten 6 (NBG, PRE); <br> Komgha ( -DB ), F1anagan 1810 (SAM). |
| :---: | :---: |
| 3228 | (Butterworth): near Kei Mouth (-CB), Flanagan 1810 (PRE). |
| 3325 | (Somerset East): Zuurberg (-AD), Paterson s.n. (GRA, Oct. 1911); Zuurberg Sanatorium, Uitenhage (-CD), Lang 177 (GRA). |
| 3326 | (Grahamstown): Grahamstown (-BC), Daly 78 <br> ( $\mathrm{PRE}, 2$ sheets); Bathurst (-DB), L. E. Tay 10 or 4966 (NBG). |
| 3327 | (Peddie): East London (-BB), L.E. Taylor 5584 (NBG), Courtenay-Latimer s.n. (NBG, Oct. 1945), Barker 3428 (NBG, 2 sheets). |
| y: |  |
| Compton 24706 (NBG), Ranches, Swaziland; |  |
| Rudatis 1736 (PRE), Campbellton, Natal; Gerstner |  |
| 3638 (NH), Zululand; Thode 2541 (STE), Smith's |  |
| Crossing, Natal; Kotze 507 (PRE), Umtlatuzi, |  |
|  | Zululand; Harrison 183 (NH), Palm Ridge Farm, |
| Zululand; Pegler 1143 (PRE), District Kentani; |  |
| Sim 1120, 1124,1129 (NU), Kabongaba, Transkei; |  |
| Scully 117 (SAM), Stockenstroom; Rogers 28367 |  |
| (STE), Manly Flats, Albany; Gerrard \& McKen1828 (NH); Tyson 2745 (STE). |  |
|  |  |

11. Hypoxis zululandensis S.E. Wood sp. nov.

Type: South Africa, Ubombo, Manzengwenya, Moll 4740 (SAKG, holo.!)

Cormus $\pm 4,0 \mathrm{~cm}$ diametro., sub-globosus, coronatus setis.
Folia $4-10,15,0-45,0 \mathrm{~cm}$ longitudine, $0,3-0,4 \mathrm{~cm}$ maxima latitudine, linearia, semi-execta, instructa $10-20$ costis, aequalibus magnitudine, approximatis, pili stellati, praediti solo brachio attingenti usque ad $2,0 \mathrm{~mm}$, residuum brachiarum $\pm 0,25 \mathrm{~mm}$ longitudine, pili opaci, densi marginibus carinaque, pili in lamina siti praecipue basi inter


Fig. 35. Hypoxis zululandensis S.E. Wood - known distribution in Southerm Africa up to 1975.
costas, hi pili instructi brachiis aequalibus longitudine $+0,25 \mathrm{~mm}$, praeditique interdum solo brachio longiore, similes pilis carina marginibusque, vel lamina glabra, (folia basalia infima subter vestita sunt pilis stellatis instructis brachiis aequalibus longitudine). Pedunculi ancipites, hirsuti. Inflorescentia praedita 2-5 floribus. Pedice11i. 0,5-1,5 cm longitudine tempore patendi florum, hirsuti. Bracteae lineares-subulatae, setosae, 0,5-1,2 cm longitudine. Perianthium seguenta lutea superficie, in floribus reclusis $1,0-1,5 \mathrm{~cm}$ longitudine. Filamenta subulata, inaequalia longitudine. Antherae 4,5-6,5 mm longitudine, lanceolatae, sagittatae, parum bifurcatae apice. Stylus cylindricus, $1,0-2,0 \mathrm{~mm}$ longitudine. Stigma concretum, $2,0-2,5 \mathrm{~mm}$ longitudine, pyramidale formatum tribus concavis faciebus. Fructus, capsula supra medium circumscissa, operculo deciduo.

Corm $\pm 4,0 \mathrm{~cm}$ in diam., sub-globose, crowned with bristles. Leaves $4-10,15,0-45,0 \mathrm{~cm}$ lang, $0,3-0,4 \mathrm{~cm}$ at widest point, 1 inear. semi-erect with 10-20 uniform ribs close together, hairs stellate, one arm up to $2,0 \mathrm{~mm}$, the remainder $\pm 0,25 \mathrm{~mm}$ long, opaque, dense on margins and keel, laminal hairs mainly at the base between ribs, these with equal arms $\pm 0,25 \mathrm{~mm}$ long, occasionally one arm longer as with hairs on keel and margins, or lamina glabrous, (the first basal leaves are abaxially covered with stellate hairs with arms of equal length). Peduncles ancipitous, hairy. Inflorescence 2-5 flowered. Pedicels $0,5-1,5 \mathrm{~cm}$ long at time of flower opening, hairy. Bracts 1inear-subulate, setose, $0,5-1,2 \mathrm{~cm}$ long. Perianth segments yellow adaxially, in open flowers $1,0-1,5 \mathrm{~cm}$ long. Filaments subulate, unequal. Anthers 4,5-6,5 mun long, lanceolate, sagittate, apex slightly bifurcate. Style cylindrical, 1,0-2,0 mm long. Stigna


Fig. 36. H. zululandensis: hair types
concrete, 2,0-2,5 mang, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: October-December.

This species is at present known only from the districts of Hlabisa and Ubombo in Zululand, Natal (Fig. 35), at altitudes of not more than 40 m . Plants appear to prefer grassland habitats but have been recorded near vleis. The specific name reflects the locale of the plants.

Hypoxis zululandensis has characteristically thin, linear leaves which are strongly nerved with often short, squat, stellate hairs (Fig. 36) on the keel and margins. The lamina is unusual in that there is a tendency for the hairs, which are situated mainly in the channels between the xibs, to be limited to the basal portion. H. rigidula is the only other Natal species in which the hairs 1ie within the channels, but this species is distinct because of its fewer, rigid leaves which are flat and not subterete in cross section as in H. zululandensis. The pedicels of H. zululandensis are also much longer than those of the subsessile flowers of H. rigidula.

Another close morphologically ally of H. zululandensis is H. obtusa, in which the dense covering of opaque hairs on the margins and keel, and the strongly nerved leaves axe similar. However, in this species the leaf lamina is wider, slightly sickle-shaped and glabrous.

The inflorescences of $\underline{H .}$ zululandensis and $\underline{H}$. filiformis are essentially similar except that first formed flowers are, respectively, either carried on pedicels $\pm 0,5 \mathrm{ca}$ long or are sub-sessile. The leaf
shape, ribbing and hair covering in H. filiformis are distinct from
 is unlikely.

## Citations:

NATAL - 2732 (Ubombo): Bazwana (-AB), Gerstner 3692 (NH);
just east of Vazi Swamp, Manzengwenya (-BA), Mol1 4740 (NPG); Redman's camp near False Bay (-CD), Gerstner 4936 (PRE).
2832 (Mtubatuba): Lake St. Lucia, eastern shores (-BA), Smook 636 (NU); R.H. Taylor 255 (NU).
12. Hypoxis filiformis Baker in J. Linn. Soc. 17:109 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:232 (1893), et in Fl. Cap. 6:180 (1896); Medley Wood, Fl. Natal:132 (1907); Nel in Engl. Jahrb. 51:305 (1914); Bews, Fl. Natal \& Zululand: 65 (1921); Guillarmod, Fl. Lesotho:149 (1971); Ross, Fl. Natal:132 (1972). Type: South Africa, Cape, Queenstown, Cooper 462 ( $K$, вуп.: )

Corm 0,6-2,0 cm in diam., globose, crowned with bristles.
Leaves $2-10,6,0-30,0 \mathrm{~cm}$ long, $0,1-0,3 \mathrm{~cm}$ at widest point, not 3 ranked, erect, rigid, filiform or linear with 6-8 prominent ribs usualIy close together; lightly pilose all over, hairs $\mathfrak{\text { Kine, }}$, white, simple and bifurcate. Peduncles $\pm$ terete, slender, grooved, silky villous below inflorescence, becoming glabrous to slightly pilose in the lower part. Inflorescence 1-3(.4) flowered. Pedicels 0,3~1,5 cm long at time of flower opening, the first-formed flower with the longest pedicel, densely villous. Bracts linearmsubulate, lightly setose


Fig. 37. Hypoxis filiformis Rak. - known distribution in Southern Africa up to 1975.

0,25-0,5 cm long. Ovary oblong, densely covered with silky, stellate hairs. Perianth segments yellow adaxially, in open flowers $0,4-1,0$ (-1,4) con long. Filaments unequal: 3 long, 3 short. Anthers 2,53,0 mm long, oblong-lanceolate, sagittate, apex bifid. Stigma subm sessile, concrete, approximately $2,0 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a capsule, opening by circumscissile dehiscence below the apex.

Flowering Period: July-February.

Hypoxis filiformis is a small species appearing "grass-like" amongst other vegetation. It is characterized by its small, globose corm; erect, subterete leaves; channelled peduncle and the arrangement of the flowers in the inflorescence (Fig. 9). Only within H. filiformis and H. angustifolia are aberrant plants known that produce flowers with 4 perianth segments instead of the usual 6 . These flowers have 4 anthers, a 2-loculed ovary and 2 connate bracts at the base of the pedicel, one of which is very much shorter than the other. Also, the perianth parts, ovary and pedicel are glabrous. Specimens of this form are recorded from the Underberg district, Natal (Kil.lick \& Vahrmeijer 4048); the Wakkerstroom district, Transvaal (Devenish 152la and Mauve 4504), and from Lesotho (Guillarmod , Getliffe \& Mzamane 274 (K)). They may be no more than occasional variants in normal populations, but this must be confirmed in the field.
H. filiformis occurs in all four provinces of the Republic (rig. 37). The species has a wide altitude range and a varied habitat tolerance (plants were found growing at sea level in Natal
and at 2500 m in Lesotho). Although there is a preference for marshy or damp areas, plants also lnhabit open grassland and sandy sofl near the coast. In the Port Shepstone area of Natal, H. filiformis tends to be more robust with the leaves up to 3.0 mm wide and not as subterete as usual. There are also up to six peduncles per plant, four flowers per peduncle and the bracts are 1 or 2 mm longer than average. H. filiformis is not readily confused with any other Natal species because of the nature of the leaves and forn of the inflorescence. Its closest ally morphologically is $H$, zululandensis but this 1ikeness is in the inflorescence rather than the leaves (see also H. zululandensis).

## Citations:




13. Hypoxis ne1iana Schinz in Vierteljahrsschr. Nat. Ges. Zurich
$71: 136(1926)$. Type: South Africa, Natal, mountains by
Estcourt, Schiechter 3348 (K, iso.!)

Corm approximately $1,0 \mathrm{~cm}$ in diameter, oblong, crowned with bristles. Leaves $4-7,9,0-12,0 \mathrm{~cm}$ long, $0,1-0,3 \mathrm{~cm}$ at widest point, erect, linear, hairs fine, yellowish, mainly 2 -armed appearing $U$ shaped, some stellate, occasionally on lamina, mostly along keel and margins, ribs 5-10, prominent, close together, the abaxial surfaces of the first one or two membranous leaves covered with hairs.

Peduncles l-4, with many stellate hairs at the apex grading to few or none at the channelled base. Inflorescence 2(-3) f1owered. Pedicels $0,6-1,5 \mathrm{~cm}$ long at time of flower opening, hairy. Bracts linear - slightly subulate, setose, $0,5-1,1 \mathrm{~cm}$ long. perianth segments yellow adaxially, $0,7-1,0 \mathrm{~cm}$ long in open flowers. Filaments subulate, equal. Anthers $2,0 \cdots 3,0 \mathrm{~mm}$ long, lanceolate, sagittate, apex entire. Style up to $1,0 \mathrm{~mm}$ long or stigma subsessile. Stigma concrete, $1,0-1 ., 5 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a
capsule opening by circumscissile dehiscence below the apex.

Flowering Period: September-December.

Baker in 1878 established H. sericea var. dregei (syntypes Drège 8525, Cape of Good Hope, and Cooper 1811, Kaffraria). The latter specimen is H. filiformis Bak. but before this was determined it was cited under H. argentea var. sericea (Bak., 1896) and under H. dregei (Bak.) Nel (Nel, 1914).

Baker himself sank H. sericea under H. argentea in 1896, as var. sericea and maintained the two original syntypes under this taxon.

Nel in 1914 upgraded to specific rank Baker's $\underbrace{H .}$ sericea var. dregei placing only Cooper 1811 under this taxon. Drege 8525 he referred to typical $H$. argentea, under which species he, unlike Baker, recognised no varieties. Under typical H. dregei Nel cited a number of other specimens. He also established H. dregei var. biflora (H. bi£lora de Wild. 1913: type Hock s.n. Belgian Congo, Katanga). Under this last variety Nel cited Schlechter 3348.

Schinz, in 1926, established H. neliana, a species he based upon Schlechter 3348, but without reference to either H. dregei or H. hiflora. An entity fitting the description of $\underline{H}$. neliana and agreeing with Schlechtex 3348 exists in Natal. I have not examined the t.ype of H . biflora and $s 0$ cannot comment upon the relationship of H. biflora and H. neliana. Until this comparison is made $I$ continue to use Schinz's name, H. neliana, for the Natal entity.

Plants of H. neliana are small and characterized by narrow, strongly ribbed leaves which are covered, especially on the keel and margins, with yellow U-shaped hairs. H. lata Nel has a similar


Fig. 38. Hypoxis neliana Schinz - known distribution in Southern Africa up to 1975.
vesture but the hairs are finer and whitish. The leaves of H . 1ata are broader, membranous and the ribs are not pronounced, Young plants of $H$. gerrardii Baker may resemble plants of $\underline{H}$. neliana, but the leaves of the former have a denser, woolly hair covering and there are only 2-4 prominent ribs on a wider lamina.

Only 5 specimens of $\underline{H}$, neliana have been recorded, all from
Natal (high areas of the Drakensberg, Estcourt and Impendhle - Fig.
38). Further collecting is likely to determine the distributional
limits, range of habitat and altitudinal preferences of this taxon.

## Citations:

NATAL - 2828 (Bethlehem): National Park (-DB), Hutchinson, Forbes \& Verdoorn 133 (NH).
2829 (Harrismith): Cathedral Peak (-CC), Ruch 1477 (PRE).
2929 (Underberg): Estcourt (-BB), Schlechter 3348 (GRA, NH, PRE); 28 km to Impendhle from Himeville (-DA), S.E. Wood 130 (NU); Impendhle (-DB), Levett 40 (NH).
14. Hypoxis kraussiana Buching. ex Krauss in Flora 28:311 (1845); Baker in J. Linn. Soc. 17:109 (1878), in Dur. \& Sckinz, Consp. Fl. Afr. 5:232 (1893), et in F1. Cap. 6:180 (1896); Medley Wood, Fl. Natal:132 (1907); Nel in Engl. Jahrb. 51:306 (1917); Bews, Fi. Natal \& Zaluland: 64 (1921); Ross, Fl . Natal:132 (1972). Type: South Africa, Natal, hills near Pietermaritzburg, Krauss 104 (B.M. holo.!)

Coxm l,0-2,0 cm in diam., globose-oblong, crowned with bristles. Leaves $4-7$, up to $28,0 \mathrm{~cm}$ long, $0,1-0,3 \mathrm{~cm}$ at widest point, lanceolale to linear-acuminate, erect, subterete, ribs 6-12, hairs stellate and


Fig. 39. Hypoxis kraussiana Buching. - known distribution in Southern Africa up to 1975.
bifurcate (more predominant) intermingled, sometimes up to 6,0 man long, white, usually patent, sparsely to densely scattered over surface. Peduncles densel.y hairy, hairs patent. Inflorescence 2-3(-4) flowered. Pedicels 0,5-1,7 cm long at time of flower opening, that of the first formed flower longest. Bracts linear-subulate, setaceous, 1,0-1,5 cm long. Perianth segments yellow adaxially, in open flowers $0,6 \mathrm{~m}, 4 \mathrm{~cm}$ long. Filaments unequal, subulate. Anthers, 3,5-5,0 mm long, lanceolate, sagittate, apex slightly or deeply bifurcate. Style cylindrica1, 0,5-1,0 mm long. Stigma concrete, 1,5-3,0 mom long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: July-November.

Hypoxis kraussiana is known from Natal and Swaziland (Fig. 39).
Its closest ally is H. neliana which occurs in the Drakensberg and south-western Natal (south of the Tugela River). Both species have narrow, erect leaves but those of H. neliana are more strongly ribbed with a predominance of yellow, bifurcate hairs on the keel and margins. The hairs are usually more numerous, shorter and more abundant than those of the leaves of $\underline{H .}$ kraussiana. The inflorescence of $\underline{H}$. kraussiana closely resembles those of H. neliana and H. filiformis, with the first formed flower having the longest pedicel.

## Citations:


maritzburg (-CB), Joliffe 52 (NU); Worlds View (-CB), Morris 366 (NU), Hodson 10 (NU); Swartkopskloof (-CB), C. Robertson 12 (NU).
15. Hypoxis gerrardii (=Gerrardi) Baker in J. Linn. Soc. 17:110 (1878), in tur. \& Schinz, Consp. Fl. Afr. 5:232 (1893), et in Fl. Cap. 6:181 (1896); Medley Wood, Fl. Natal:132 (1907); Nel in Engl. Jahrb. 51:306 (1914); Bews, F1. Natal \& Zululand: 65 (1921); Guillarmod, F1. Lesotho:149 (1971); Ross, Fl. Natal:132 (1972). Type: South Africa, without locality, Gerrard \& McKen 1827 (NH, holo.!)

Corm 1,0-2,5 cm in diam., obconic, crowned with fine bristles.
Leaves $5-19,10,0-50,0 \mathrm{~cm}$ long, up to $1,1 \mathrm{~cm}$ broad at widest point, 3-ranked, linear-lanceolate, semi-erect, with usually 2 , but occasionally 4, prominent ribs, woolly-villous all over, especially on margins and keel, hairs fine, whitish-golden, predominantly bifurcate, very occasionally stellate. Peduncles densely villous. Inflorescence 2-4 flowered. Pedicels $1,5-4,0 \mathrm{~cm}$ long at time of flowering, densely hairy. Bracts 1inear-subulate, setose, 0,5-1,2 cm long. Ovary oblongturbinate, densely hairy. Perlanth segments yellow adaxially, 0,6$1,3 \mathrm{~cm}$ long in open flowers. Filaments unequal. Anthers $3,0-4,0 \mathrm{~mm}$ long, lanceolate, deeply sagittate, apex bifurcate. Style subulate or stigma subsessile. Stigma concrete, up to $4,0 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex, followed by longitudinal splitting into 3 valves.

Flowering Period: All year.

Hypoxis gerrardii is a very common Natal species, occurring predominantly in grassland habitats where the soil is sandy and often rocky. Plants are also known from the Transvaal, N.E. Cape and Swaziland, where they occur over an extensive distributional range (Fig. 40).

The species is characterized by its narrow, linear leaves, which arc densely covercd with bifurcate hairs and thus appear woolly, and in which there is usually a single prominent rib (sometimes 2) on either side of the keel. The pedicels are comparatively long (1,5$4,0 \mathrm{~cm}$ ).
H. gerrardii is the only species known to flower all year under field conditions. During the months of May, June and July (1935) this species was in flower in the Highflats area. Most of the plants were sheltered by rocks and were growing on the north-eastern aspect of a slope. It is more likely that these plants were sheltered from fire and frost by the rocks and thus they realized their potential for continued growth throughout the year.
H. gerrardii is very closely allied to H. argentea (for details see under this species).

## Citations:

| TRANSVAAL | - | 2530 | (Lydenburg): Belfagt (-CC), Leendertz 7892 (PRE). |
| :---: | :---: | :---: | :---: |
| SWAZIIAND | - | 2631 | (Mbabane): Mbabane (-AC), Compton 27211 (NBG). |
| NATAL | - | 2732 | (Ubombo): Ubombo Hill (-CA), Ward 1655 (NH, NU). |
|  |  | 2829 | ```(Harrismith): Cathedral Peak (-CC), S.E. Hood 156, 149A (NU).``` |
|  |  | 2830 | (Dundee): Krantıkloof (-DD), Rogers 24583 |
|  |  |  | (STE); Haygarth 76 (STE); Kranskop (-DD) |

2831 (Nkandla): Spind Estates, Melmoth (-AD), S.E. Wood 90 (NU); Eshowe (-CD), Lawn 1019, 1019a, 1746 (NH); Umhlatuzi valley (-DA), Lawn 1684 (NH); Mtunzini (-DD), Lawn 783 (NH).
2832 (Mtubatuba): H1uhluwe Game Reserve (-AB), Fakude 15 (NH), Ward 1448 ( NH ).
2929 (Underberg): Cathkin Park (-AB), Howlett 9 (NH); Kamberg ( -AD ), Wright 24, 1552 (NU), C.G. Gordon-Gray 107 (NU); Giants Castle ( -AD ), Skead 203 (NU), Trauseld 869 (NU), BruynsHaylett 89 (NU); Yorkshire Wolds (-BB), Thode 2533 (STE); Lowlands (-BB), Schelpe 13 (NU); Balgowan (-BD), Mogg 3518 (PRE); Sani-Pass (-CC), Werdermann \& Oberdieck 1440 (PRE); Lot 7 Mawaqua, Polela (-DA), Rennie 91 (NU); Impendhle (-DB), Levett 25 (NH); near Bulwer ( -DD ), Landsdel1 5.n. (NH 34266).
2930 (Pietermaritzburg): Greytown (-BA), Wylie s.n (PRE, Nov. 1936; NH 28020); 10 km from Wartberg ( -BC ), S.E. Wood 75 (NU); Kingscliffe (-BC), S.E. Wood 78 (NU); between Glenside and Kingscliffe (-BD), S.E. Wood 77A (NU); Pietermaritzburg (-CB), Allsopp 728, 921 (NH, NU); Pennefather 28 (NU), Irwin 18 (NU), Fairall 57 (SAM), Barker 5162 (NBG), S.E. Wood 74, 83, 137, 142 (NU), Ram s.n. (NU, 2.3.74), Stirton s.n. (NU, 21.10.74); Elandskop (-CB), Webb 26 (NU); Table Mtn. (-CB), Killick 225 (NU); Thornville (-CD), Moll 3394 (NU); Cato Ridge (-DA), Ross 2159 (NH, PRE); 30 km to Umbumbulu on Eston Rd. (-DA), S.E. Wood 103A (NU); Inanda (-DB), Medley Wood 327 (NH), Strey 5170 (NH); Botha's Hil1 (-DC), Hutchinson, Forbes \& McClean 52 (NH); Drummond (-DC), Stirton 1091 (NU); 32 km to Umbumbulu ( -DC ), S.E. Wood 103 (NU); 8 km to Mid-Illovo (-DC), S.E. Wood 109, Il5 (NU); Hillcrest (-DD), Nieuwoudt 33 (NU); Umlaas River (-DD), Stirton 513 (NU); Key Ridge (-DD), S.E. Wood 120 (NU); Cowies Hill (-DD), Lawson 1219 (NH, PRE).
2931 (Stanger): Kearsney (-AB), Milner s.n. (NH 23260).

3029 (Kokstad): Harding (-DB), L.E. Taylor 5376 ( NBG ).
3030 (Port Shepstone): Highflats (-AA), S.E. Wood 81 (NU); 'Campbellton' ( -AB ), Rudatis 1381 (STE); Hlutankungu (-AB), S.E. Wood 80 (NU); on road to Port Edward from Izingolweni (-CC), S.E. Wood 164 (NU).

CAPE - 3029 (Kokstad): Cedarville (-AC), Baudert 24 (GRA); Kokstad (-AA) Bayliss 2547 (SAM).
3128 (Umtata): between Qumbu \& Shawbury (-BB), Schonland 4144 (GRA).
3326 (Grahamstown): Grahamstown (-BC), Rogers 27546 (STE).
without precise locality:
Gerrard \& McKen 1827 (NH); Markotter 8692 (STE), Nhlopenkulu Mission station, 2ululand.
16. Hypoxis argentea Harv. ex Baker in J. Linn. Soc. 17:110 (1878), in Dur. \&r Schinc, Consp. F1. Afr. 5:231 (1893), et in Fl. Cap. 6:181 (1896); Medley Wood, Fl. Natal:132 (1907); Ne1 in Eng1. Jahrb. 51:305 (1914); Bews, Fl. Natal \& Zululand:64 (1921); Guillarmod, Fl. Lesotho:149 (1971); Ross, Fl. Natal:132 (1972). Type: South Africa, Cape, Grahamstown, MacOwan 50 (GRA, SAM, iso-syn.!)

Corm 0,5-2,0 cm in diam., globose-oblong, crowned with few, fine bristles. Leaves $4-14,5,0-50,0 \mathrm{~cm}$ long, $0,1-0,7 \mathrm{~cm}$ at widest point, linear to linear-lanceolate, prominent ribs 2-8, covercd with long, fine, golden (occasionally whitish especially on older leaves), appressed, stellate or bifurcate hairs, or both types intermingled, which produce a sericcous effect abaxially and on the margins. Peduncles hairy. Inflorescence 1-4(-5) flowered. Pedicels 0,9$3, O(-3,9) \mathrm{cm}$ long at time of flower pening, hairy. Bracts linear, setose, $0,5-1,2 \mathrm{~cm}$ long. Perianth segments yellow adaxially, 0,5 $1,2 \mathrm{~cm}$ long, in open flowers. Filaments 3 long, 3 short. Anthers 1,5-3,5 mm long, lanceolate, deeply sagittate, apez bifurcate. Style $1,0 \mathrm{~mm}$ long or stigma subsessile. $\quad \underline{\text { Stigma concrete, } 1,0-2,5 \mathrm{~mm}}$ long, pyramidal with 3 concave faces. Fruit a capsule, opening by circumscissile dehiscence below the apex.



Fig. 40. Hypoxis argentea Rak. and Hypoxis gerrardii Bak. - knøwn distribution in Southern Africa up to 1975 .

At the time of publishing $H$. argentea, Baker, in the same journal, also established H. sericea (syntype: Zeyher 950, Uitenhage). This latter species was said to differ from H. argentea in possessing a greater number of larger and wider leaves in which the veins were "distinctly sculptured" (unlike $H$. argentea where the veins were said to be not prominent), and with the "underneath clothed with sericeous, appressed hairs," (not as in H. argentea where they were "finely silky all over"). Baker established two varieties under H. sericea: var. drege1 (syntype Drège 8525 from the Cape of Good Hope), and var. flaccida (syntypes Williamson s.n., Albany, and Burke s.n. "Seven Fountains"). In 1896 Baker himself sank H. sericea under H. argentea. H. sericea var. sericea and H. sericea var. dxegei he placed with H. argentea as var. sericea, but H. scricea var. flaccida he maintained as H. argentea var. flaccida. Examination of the specimen Burke s.n. (syntype of H. argentea var. flaccida) showed it to be synonymous with the cited material of $H$. argentea var. sericea. The specimen was collected in February, (thus at the end of its growing season) and has exceptionally long leaves. This range of leaf length was not taken into consideration when var. flaccida was established.

In the course of this study it has become apparent that two entities are evident within the limits of $H$. argentea. The distributions of these taxa are not discrete (Fig. 40), and they are only slight1y differentiated morphologically. Thus they are treated as varieties, namely $\underline{H}$. argentea $v a r$. argentea and $\underline{H}$. argentea var. sericea (Baker's variety in which is norv included his var. flaccida).

Vars. argentea and sericea differ by the hair covering on the leaves. Plants of var. argentea are densely clothed with stellate hairs on the keel, margins and abaxial lamina surface; the
leaves of var. sericea Baker are less densely hairy, the predominantly bifurcate hairs, that are occasionally intermingled with the stellate type, being confined mainly to the margins and keel.

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H. argentea var. argentea
    Baker in J. Linn. Soc. 17:110 (1878), in Dur. & Schinz,
    Consp. Fl. Afr. 5:231 (1893), et in Fl. Cap. 6:l81
    (1896); Nel in Engl. Jahxb. 51:305 (1914). Type:
    South Africa, Cape, Grahamstown, MacOwan 50 (GRA, SAM,
    iso-byn.!)
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Leaves 4-14, 5,0-30,0 cm long, densely covered with appressed, stellate hairs which produce a sericeous effect especially on the abaxial surface, prominent ribs 2-4, - equal. Inflorescence 1-4 flowered. Pedicels $0,9-3,0(-3,9) \mathrm{cm}$ long at time of Elower opening.

Flower Period: August-February.

Hypoxis argentea var. argentea is not known in Natal, but it occurs in the eastern Cape, the Orange Free State, the Transvaal and Lesotho (Fig. 40).

## Citations:



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CAPE - 3126 (Qucenstown): Jamestown (-BE), Barker 2124
    (NBG); Fincham's Nek (-DD), Galpin 2194 (GRA);
    Mtns. Queenstown (-DD), Galpin 1062 (K).
3224 (Graaff-Reinet): near Graaff-Reinet (-BA), Bolus
176 (K).
3226 (Fort Beaufort): Fairford (-BD), Cotterrell 63
    (GRA); Katberg (-DA), Sole 393 (GRA); nr.
    Seymour (-DB), Scully 107 (SAM).
```

3325 (Somerset East): Uitenhage (-CD), Thode A703
(NH), Zeyher 6 (STE), 950 (K, SAM, STE);
Uitenhage \& Drostdy Farm (-CD), Burchell 4469
(K).
3326 (Grahamstown): nr. Sidbury (-AC), Daly 766
(GRA); Albany (-AD), L.E. Tavlor 5946 (NDG);
Grahamstown (-BC), Glass 378 (SAM), L.E. Taylor
3634 (NBG), Rogers 27404, 28367 (STE), MacOwan
50 (GRA, K, SAM), Daly \& Cherry 977 (GRA);
Southwe 11 (-DB), Bayliss 3138 (NBG).
3424 (Humansdorp): between Jeffries Bay and Ferreisa-
town (-BB), Gillett 2316 (SIE); Humansdorp
(-BB), Thode Al049 (NH).

Hypoxis argentea var. sericea Baker in F1. Cap. 6:181 (1896);
Medley Wood, F1. Nalial:132 (1907); Bews, Fl. Natal \&
Zululand: 64 (1921); Guillarmod, Fl. Lesotho:149 (1971);
Ross, Fl. Natal:132 (1972). Type: South Africa, Cape
of Good Hope, Drege 8525 ( BM, syn.!)

Leaves $5-9,5,0-50,0 \mathrm{~cm}$ long, ribs $4-8$, of which 2 are more
prominent than the others, hairs long, fine, mainly bifurcate, occasionally with a few stellate intermingled, appressed, situated predominantly on margins, keel, and abaxial surface, but not as dense nor as markedly sericeous as in var. argentea. Inflorescence 2-4(-5). £lowered. Pedicels of open flowers $1,2-2,8 \mathrm{~cm}$ long.

Plowering Period: August-March.

Plants of var. sericea are present in all four provinces of
South Africa (the southern-most limit at the present time being Katberg, Cape Province), and Lesotho (Fig. 40, overlay).

The closest morphological ally of H . argentea is H . gerrardii
Bak. Leaves of the latter species are covered with bifurcate hairs which are not appressed, so the indumentum appears woolly: in all other characteristics it is similar to H , argentea, but it is more eastern in distribution (Fig. 40). Comparison of distributions shows that $\underset{H}{ }$ argentca var. argentea occurs inland, H. gerrardii is more coastal, while H. argentea var. sericea is sympatric over the distributional area of the two first-named taxa. It is tempting to postulate that H. argentea var. sericea had its origin in hybridization between the two adjacently distributed taxa $\underline{H \text {. argentea and }} \underline{H \text {. gerrardii, }}$ and that progeny of this crossing now constitutes an established entity showing some morphological similarity with each of the putative parents.

Proof of this is impossible without experimental crossings and more exhaustive field work. Until such time as further evidence supports this conjecture, I have maintained $\underline{H}$. gerrardii as distinct, since the species is already established and since plants are reasonably easily differentiated from $\underline{H \text {. argentea }}$ var. argentea, and $\underline{H \text {. argentea }}$ var. sericea.

Citations:

| transvail | - | 2529 | (Witbank): Middleburg (-CB), Rudatis 75 (STE). |
| :---: | :---: | :---: | :---: |
|  |  | 2531 | (Kømatipoort): Plaston (-AD), Halt 27 (NiI) |
|  |  | 2628 | (Johannesburg): Dunswart (-AB), Moss 13981 (J). |
| O.F.S. | - | 2827 | (Senekal): Gumtree (-CD), Ross 45 (NU); Ficksburg (-DD), Galpin 13840 (PRE). |
|  |  | 2829 | (Harrismith) : Swinburne (-AD), Jacobsz 141 (PRR). |
|  |  | 2926 | (B1oemfontein): Bloemfontein (-AA), Pretorius |
|  |  |  | s.n. (STE, Jan. 1936), Relmann 3761 (K); Thaba Nchu (-BB), Roberts 2388 (PRE); Dewetsdorp. (-DA), Steyn 933 (NBG). |
| NATAL | - | 2730 | (Vryheid): Utrecht (-CB), Thode Al296 (NH). |
|  |  | 2831 | (Nkandla) : 曹 m. s. of Nkwalini (-DA), Codd |
|  |  |  | 1849 (PRE); Nkwalini (-DA), Acocks 12960 (PRE). |
|  |  | 2929 | (Underberg): Estcourt (-BB), Mogg 3449 (PRE). |


17. Kypoxis angustifolia Lam. Encycl. méth. Bot. 3:182 (1789); Schult. Syst. Veg. 7:767 (1819); Fisch. et Mey. Animad. bot. in Ind. sem. hort. petrop. 10:49 (1845); Baker in F1. of Maur.:369 (1877), in J. I.inn. Soc. 17:111 (1878), in Dur. \& Schinz, Consp. F1. Afr. 5:231 (1893), et in F1. Cap. 6:180 (1896); Medley Wood, F1. Natal:132 (1907); Nel in Engl. Jahrb. 51:303 (1914); Bews, Fl. Natal \& Zululand:64 (1921); Perrier de la Bathie in F1. Madag. 41:10 (1950); Guillarmod, Fl. Lesotho:148 (1971); Ross,



Corm 0,6-2,5 cm in diam., oblong-globose, crowned with fine bristles. Leaves 5-9 (-15), 7,0-60,0 cm loag, $0,2-1,7 \mathrm{~cm}$ at widest point, 3-ranked, lanceolate, oblong-lanceolate or linear-acuminate, flaccid or seni-erect, membranous, usually with $2-4$ prominent ribs,
hairs white, fine, pilose, simple or simple and bifurcate intermingled, at $45^{\circ}$ angle, predominantly along margins and keel. Inflorescence 1-7 flowered. Pedicels $1,0-6,5 \mathrm{~cm}$ long at time of flower opening, pilose. Bracts linear-acuminate, with sparse keelar hairs, 0,6-1,2 cm long. Ovary oblong, in some cases up to $1,0 \mathrm{~cm}$ long. Perianth segments yellow adaxially, $0,6-1,2 \mathrm{~cm}$ long in open flowers. Anthers $2,0-4,0 \mathrm{~mm}$ long, lanceolate, sagittate, apex bifurcate. Filaments not equal, 3 long, 3 short. Style subulate, approximately $1,0 \mathrm{~mm}$ long. Stigma concrete, up to $3,0 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex, followed by the splitting of 3 longitudinal valves.

Flowering Period: July-May.

Within the limits of H . angustifolia, Baker recognized two entities, typical H. angustifolia and H . angustifolia var. buchananii. These two taxa are retained because the difference in their leaves renders them morphologically distinct. Var. angustifolia has narrow, 1anccolate or linear-acuminate, semi-erect leaves, while the leaves of var. buchananii are wider (up to $1,7 \mathrm{~cm}$ ), oblongwlanceolate, and flaccid. Var. buchananii also tends to have a larger corm, longer pedicels; and more flowers per inflorescence than var. angustifolia.

Hypoxis angustifolia var. angustifelia
Lam. Encycl. méth Bot. 3:182 (1789); Schult. Syst. Veg. 7:767 (1819); Fisch. et Mey, Animad bot. in Ind. sem. hort. petrop. 10:49 (1845); Baker in Fl. of Maur.:369 (1877), in J. Linn. Soc. 17:111 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:231 (1893), et in Fl. Cap. 6:180 (1896); Medley Wood, Fl. Natal:132 (1907); Ne1 in Engl. Jahrb. 51:303 (1914); Jews, Fl. Natal \& Zululand:64 (1921); Perrier de la Batra in Fl. Madag. 41:10 (1950); Guillarmod, Fl. Lesotho:148 ( 71); Ross, F1. Natal:132 (1972).


Fig. 41. Hypoxis angustifolia Lam. var. angustifolia known distribution in Soutnern Africa up to
1975.

Type: South Africa, Orange Free State, Cooper 1089 (? ho10.)

Corm 0,6-2,0 cm in diam., oblong-globose. Leaves 7,0-30,0 cm long, $0,2-0,9 \mathrm{~cm}$ at widest point, lanceolate or linear-acuminate, semierect. Inflorescence $1-4$ flowered. Pedicels $1,0-3,5 \mathrm{~cm}$ long.

Flowering Period: September-May.

Plants of var. angustifolia are known from all four provinces of South Africa and Lesotho (Fig, 41). Distribution tends to be concentrated in the eastern sector of the country, and records for the Transvaal are sparse. H. angustifolia extends to North Africa and is also the only species of Hypoxis to occur on the islands of Mauritius and the Seychelles.

Plants of this variety are generally small. The membranous leaves arc subglabrous carrying, as a rule predominantly or the margins and keel, only a few scattered, fine, usually white, hairs.

Citations:

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var. angustifolia
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TRANSVAAL - 2230 (Messina): Sibasa (-CD), Smuts \& Gillett 3165 (PRE, STE), Junod 25453 (PRE).
O.F.S. - 2926 (Bloemfontein): O.F.S. Botanic Garden, Bloemfontein (-AA), Mi1ler 333, 369 (NBG).
NAI'AL - 2732 (Ubombo): Sordwana Bay Park (-AD), S.E. Wood 86A (NU); Road to Maputa (-BA), Strey 4772 ( $\overline{\mathrm{NH}}$ ); False Bay Park ( -CD ), Ward 4131 (NH).
2829 (Harrismith): Van Reenen (-AD), Medley Wood 9649 (SAM), Haygarth s.n. (NH 9923); BrakFontein mr. Frere (-DD), Acocks 10832 (NH).
2830 (Dundee): Krantzkloof (-DD), Rogers 24665 (STE).
2831 (Nkand1a): MLunzini (-DD), Mogg 5880 (NH).
2832 (Mtubatuba): Hluhluwe Gane Reserve (-AA), Mncomezulu 7 (NH).



Fig. 42. Hypoxis angustifolia Lam. var. buchananii Bak. - known distribution in Southern Africa up to 1975.
H. angustifolia var. buchananii Baker in J. Linn. Soc. 17:111 (1878), in Dur. \& Schinz, Consp. F1. Afr. 5:231 (1893), et in F1. Cap. 6:180 (1896); Medley Wood, Fl. Natal:132 (1907); Bews, F1. Natal \& Zululand: 64 (1921). Type: South Africa, without locality, Buchaman s.n. (K, holo.!)
 Consp. Fl. Afr. 5:236 (1893), et in F1. Cap. 6:183 (:296); Medley Wood, Fl. Natal:132 (1907). Type: Sonth Africa, Natal., Inanda, Medley Wood 426a (K, isolo.!)
H. oti. $\frac{\text { Ear }}{}$ var. woodif Nel in Eng1. Jahrb. 51:309 (1914); Berss, F1. Natal \& Zululand:64 (1921); Ross, Fl. Natal:132 (1972). Type: South Africa, Natal, Inanda, Medley Wood 426a (K, syn.! )

Corm up to $2,5 \mathrm{~cm}$ in diam., eblong. Leaves up to 60 cm
long, $0,6-1,7 \mathrm{~cm}$ at widest point, oblong-1anceolate, flaccid, membranous.
Inflorescence (1-)2-7 flowered. Pedicels usually very long, up to 6,5 cm.

Flowering Period: July-December.

The distribution of var. buchananii is similar to that of var. angustifolia except that one gathering is known from Swaziland (Fig. 42).

Baker in 1889 established H. woodii (ty, : Medley Wood 426a). Seven years later he cited (Baker, 1896) Medlev ie 426 as H. angustifolia var. buchananii. Examination of both the a Medley Wood specimens (Fig. 43) showed them to be conspecific, thus $\mathrm{H}^{\text {. woodiz must become a }}$ junior synonym of H. angustifolia var. buchanarii.

Nel in 1914 placed Baker's H. wood. 1 in as variety of 11 . obligua, (type: Medley Wood 426a). This variety must also become a synonym of H. angustifolia var. buchananii. H. obligua is a distinct species 1 imited to the Cape and is not affected by this synonymy.
$\left.j^{1}, d\right\}$


a

b

c

Fig. 44. Herbarium specimens of putative parents and their hybrid

[^0]The closest morphological ally of $\underline{H}$. angustifolia is $\underline{H}$. lata. These taxa differ in that the leaves of the latter species are sickleshaped (not linear-lanceolate), with more frequent, less fine, patent hairs situated predominantly on the margins and keel. The perianth segments of H. lata also tend to be longer ( $1,2 \sim 1,7 \mathrm{~cm}$ ).

There is some evidence that H. angustifolia var. buchananii hybridizes in nature. Studies of natural populations of this species that grow sympatrically with populations of $H$. acuminata have revealed, among the putative parents, a range of intermediates that are probably hybrids (for further details see under Chapter 4 - Hybridization).

Another species with which var. buchananii seems to hybridize is H. membranacea, but intermediates in this case havc only been detected by study of herbarium specimens (Fig. 44).

In these two species there are differences in the styles and stimas, namely:
H. membranacea: stigma minute, ( ${ }^{+} 0,5$ man $)$ spherical; style filiform, $\pm 3,0 \mathrm{~mm}$ long (Fig. 45 c ).
H. angustifolia var. buchananii: stigma concrete, up to 3,0 mun long, pyramidal with 3 concave faces; style subulate, $\pm 1,0$ mm long (Fig. 45a).

Herbarium specimens were found which exhibited characters of both the above taxa. All specimens possessed flaccid, membranous, lanceolate leaves with bifurcatc hairs on the margins and keel (characteristic of $H$. angusti.folia var. buchenanii). Perianth segments were yellow or white adaxially (characteristic of $H$. membranacea where flower colour is white or yellow). One specimen of H . angustifolia var. buchanarii (Medley Wood 77l) was stated to have white flowers. This possibl.y

a
H. angustifolia var. buchananii

Umzinto:
Strey 5974
Weintroub 17124
Komgha:
Compton
17668
Stutterheim:
de Vries 85
King Williams Town: Dyer 298
putative var. buchananti $x$ membranacea

Port Shepstone:
Strey 5990, 9149, 6879
Elliotdale:
J.L. Gordon-Gray 949

## Kentani:

Compton 17716
Port St. Johns:
Strey 10241
c
H. membranacea

Transke i: Taylor 3710

Port Edward: Nicholson 713

Port Shepstone:
Klopper 5.n.
Glen: 274
East London:
Smith 3770
Galpin 5810

## Komgha:

Flanagan 1172
Kentani:
Pegler 109

Fig. 45. Stigmatic and haix types of:
a - H. angustifolia var. buchananii, b - putative hybrids, c - H. membranacea
indfcates a similar range in flower colour in this species. The stigmas and styles of the putative hybrids ranged between those of the putative parents (see Fig. 45b).

Both H. angustifolia var, buchananii and H. membranacea have a coastal distribution being sympatric over part of their range. H. membranacea occurs from East London to Barberton, but var. buchananii has wider limits extending into the N . Transvaal and Rhodesia. Both species prefer moist, shady habitats and it seems probable that some degree of hybridization occurs where the taxa overlap (a region from Umzinto to East London).

Population studies in the field, and intensive experimental work are necessary to confirm hybridization.

## Citations:

var. buchananii


|  |  | 3030 | (Port Shepstone): 'Ellesmere' (-AD), Rudatis 1150 (STE); Amanzimtoti (-BB), Franks s.n. (NH 14174); Park Rynie (-BC), Strey 5974 (NH, NU); Umdoni Park ( -BC ), Moss 17124 (J). |
| :---: | :---: | :---: | :---: |
| LeSotho | - | 2828 | (Bethlehem): Leribe (-CC), E. Phillips (SAM 6755, 2 sheets), Dieterlen 536 (SAM). |
| CAPE | - | 3226 | (Fort Beaufort): Hogsback, Amatola Mts. (-DB), Peacock s.n (SAM 65757): Hogsback Forest Reserve (-DB), Dah1strand 1793 (STE); Hogsback (-DB), Noel 1485 (GRA). |
|  |  | 3227 | (Stutterheim): Keiskama Hoek, King Williams Town (-CD), Dyer 298 (PRE); 3 ml . from Amabele (-DA), de Vries 85 (PRE); Komgha (-DB), Compton 17668 (NBG). |
| without precise locality: ${ }_{\text {Bell }}$ s.n. ( NH 14151). |  |  |  |
|  |  |  |  |

putative hybrids - H. angustifolia var. buchananii X H. membranacea

18. Hypoxis lata Nel in Engl. Jahrb. 51:324 (1914); Bews, F1. Natal \& Zululand:65 (1921); Ross, Fl. Natal:132 (1972). Type: South Africa, Natal, Van Reenen, Medley Wood 6254 (NM, syn.:)

Corm 0,5-1,2 cm in diam., globose-oblong, crowned with fine bristles. Leaves 5-9, 9,5-37,0 cm J.ong, $0,4-1,2 \mathrm{~cm}$ at widest point, lanceolate or linear-acuminate, sickle-shaped, usually membranous, with 2-8 prominent ribs, hairs stellate or bifurcate, or both types inter-


Fig. 46. Hypoxis lata Nel - known distribution in Southern Airica up to 1975.
mingled, patent, predominantly on keel and margins, lamina with few, scattered hairs. or glabrous. Peduncles ancipitous, hairs patent, whitish, dense below inflorescence grading to sparse at base. Inflorescence 2-4(-5) flowered. Pedicels $0,5-4,0 \mathrm{can}$ long at time of flower opening. Bracts linear-subulate, setaceous, $0,6-1,6 \mathrm{~cm}$ long. Perianth segments yellow adaxially, in open flower $1,2-1,7 \mathrm{~cm}$ long. Filaments unequal, subulate. Anthers 4,0-5,0 mon long, lanceolate, sagittate, apex bifurcate. Style $\pm 1,0$ mun long, or stigma subsessile. Stigma concrete, $2,0-3,5 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.

Flowering Period: September-December.

Hypoxis lata is known only from Natal and Lesotho (Fig. 46), at altitudes from 500 to 2000 m . Plants are 2-4(-5) flowered with the inflorescence and peduncle covered with white, patent hairs. Leaves are sickle-shaped, membranous, with a predominance of patent hairs on keel and margins. The lamina of the leaf is usually glabrous but occasionally scattered hairs are present. Plants are recorded from moist or dry grassland habitats.
H. lata is very similar morphologically to its closest ally H. acuminata which differs by possessing leaves that are not sickleshaped nor membranous: they are hairy all over with more prominent, more closely packed ribs than are the leaves of $\mathrm{H} .1 a t a$.

Inflorescences of H . acuminata are clothed with yellowish hairs (unlike the whitish hairs of H. lata) and the pedicels are shorter than those of H. lata. H. lata is also allied to H. angustifolia var. angustifolia (for details see under this species).

| NATAL | . | 2829 2929 2930 | (Harrismith): Van Reenen (-AD), Medley Wood 6254 (NH), Cathedral Peak; Catchment 1 (-CC), S.E. Wood 150 (NU). <br> (Underberg): 8 km from Estcourt (-BB), S.E. Wood 145A (NU); Pevensey turnmoff on Underberg Rd. ( $-D C$ ), S.E. Wood 125 (NU). <br> (Pietermaritzburg): Darg1e (-AC), Smook 61.4 (NU); Greytown (-BA), Wylie s.n. (PRE, Oct.Nov. 1931); 40 km to Bulwer (mCA), S.E. Wood 121A (NU); Chase Valley, Pietermaritzburg $(-C B), S . E$. Wood 139, 141 (NU); Howick old main road, Pietermaritzburg ( $-C B$ ), S.E. Wood 144 (NU); Ferncliff, Pietermarilizburg (-CB), S.E. Wood 138 (NU); 11 km from Mid-Illovo ( -CD ), S.E. Wood 110 ( NU ); 8 km Erom MidIllovo (-CD), S.E. Wood 113 (NU); Inchanga cutting (-DA), K.D. Gordon-Gray 10012 (NU); Botha's Hill (-DC), Stirton 1078 (NU). |
| :---: | :---: | :---: | :---: |
| LESOTHO | - | 2928 | (Marakabei): Mamalapi (-AC), Compton 21365 (NBG). |

## (Tharb.)

19. Hypoxis soboliferakJacq. Icon. pl. rar. II t. 372 (1786-93), in Coll. bot. (Suppl.):53 (1796) and in Bot. Mag. t. 711 (1804); Nel in Engl. Jahrb. 51:309 (1914); Bews,F1. Natal \& Zululand:64 (1921). Type: South Africa, Cape, Thunberg s.n. (स, holo.)

## UPG,

H. villosa var. sobolifera Baker in J. Linn. Soc. 17:114 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:236 (1893).
H. pannosa Baker in Gard. Chron.:134 (1874).
H. villosa var. pannosa Baker in J. Linn. Soc. 17:114 (1878), in Dur. \& Schinz, Consp. F1. Afr. 5:236 (1893), in Fl. Cap. 6:184 (1896); Ross, Fl. Natal:132 (1972).

Corm 1,3-.3,5 cm in diam., globose-oblong, crowned with bristles.
Leaves $5-14,11,0 \mathrm{~m} 41,0 \mathrm{~cm}$ long, $0,7-2,5 \mathrm{~cm}$ at widest point, abaxial
surface, keel and margins with a scattered to dense covering of light~


#### Abstract

brown to red-brown non-appressed hairs, up to 4,0 mong, mainly stellate, rarely bifurcate or both types intermingled, adaxial surface as above but with fewer hairs. Peduncles ancipitous, hairy all over or below the inflorescence grading to subglabrous at the base. Inflorescence 2-7 flowered. Pedicels hairy, $1,0-5,5 \mathrm{~cm}$ long at time of flower opening, the first formed flowers with the longest pedicels. Bracts linear-subulate, setose, 0,6-2,2 cm long. Perianth segments yellow adaxially, in open flowers $0,8-1,6 \mathrm{~cm}$ long. Filaments subulate, 3 long, 3 short. Anthers 2,5-5,0 mm long, lanceolate-oblong, sagittate, арек deeply bifurcate. Style cylindrical, $1,0-2,5$ mim long. $\underline{\text { Stigma }}$ concrete, $1,0-3,0 \mathrm{~mm}$ long, pyramidal with 3 concave faces. Fruit a capsule opening by circumscissile dehiscence below the apex.


Flowering Period: All year.
H. sobolifera was first established by Jacquin in 1796.

Baker (1878) sank it as a varicty of H. villosa L., but it was later reinstated as a species by Nel (1914).

Within H. sobolifera Nel recognized three infraspecific categories, namely var. sobolifera (Jacquin's species), var. pannosa Coriginally Baker's species $H$. pannosa, but later reduced by the same author to a variety within $H^{H .}$ villosa), and var. accedens (Nel's own taxon). Nel did not elaborate on the differences among these taxa except to state that the leaves of var. accedens were long and narrow. I have not been able to distinguish plants with such long, naryow leaves or to recognize a third entity within the species. I am thus maintaining only var. sobolifera and var. pannosa.


Fig. 47. H. sobolifera var. sobolifera: variation in leaf shape


These two taxa differ according to thc hairs on the leaves. Leaves of var. pannosa are denscly covered in red-brown hairs, whereas the leaf hairs of var. sobolifera are less dense and light brown. Within both vars. there is a similar range in leaf width, so this criterion cannot be used to distinguish them (Figs. 47 \& 48). The peduncles in the former variety are hairy all over, while in the latter they become subglabrous towards the base.

The closest ally of H. sobolifera is H. villosa Bak., a Cape species. Within H. villosa, light-brown haired plants have been placed under var. canescens (Fisch. \& Mey.) Baker. I consider H. sobolifera var. pannosa to be a junior synonym of H . villosa var. canescens (Fisch. \& Mey, ) Baker, but it is inadvisable to sink it formally until H. villosa is treated in detail and in conjunction with H. sobolifera.

Hypoxis sobolifera var. sobolifera
(Thunb.) Jacq. Icon. pl. rar. II t. 372 (1786-93), in Coll. bot. (Supp1.):53 (1796), in Bot. Mag. t. 711 (1804); Ne1 in Eng1. Jahrb. 51:309 (1914): Bews, Fl. Natal \& Zululand: 64 (1921). Type: South Africa, Cape, Thunberg: s.n. (\#, holo.) UPS,
H. villosa var, sobolifera Baker in J. Linn. Soc. 17:114 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:236 (1893).

Leaves 5-11, covered with light-brown hairs. Peduncles hairy below the inflorescence grading to subglabrous at the base.
H. sobolifera var. sobolifera is coastal in distribution except for four specimens which are recorded from the Natal Drakensberg. The range of distribution extends from Stellenbosch to Eshowe, Zululand (Fig. 49). This is the only Natal species which is recorded in the southern Cape.

Fig. 49. Hypoxis sobolifera Jacq. var. sobolif.era known distribution in Southern Africa up
to 1975.

Its closest morphological ally in Natal is H. angustifolia var. buchananii Bak. The hairs on the leaves of the last-named species are white and bifurcate only, whereas the hairs of H. sobolifera vax. sobolifera are brown and are both bifurcate and stellate. Pedicels of both species are exceptionally long; those of the two first formed
flowers of H. sobolifera var. sobolifera are often noticeably longer than the remainder, while this is not as pronounced in H. angustifolia var. buchananii where the pedicels are almost equal in length.

## Citations:

var. sobolifera
NATAL - 2828 (Bethlehem): Natal National Park (-DD) Martin 475 (NBG), Stanton 28 (NU); Mont aux Sources (-DD), Bayer \& McClean 196 (GRA; PRE).
2831 (Nkand1a): Eshowe (-CD), Lawn 1188 (NH);
'Hamewith' near Mtunzini (-DD), Mogs 4486 (PRE).
3029 (Kokstad): Rooi Vaal, Harding (-DB), L.E. Taylor 5362 (NBG).
CAPE - 3129 (Port St. Johns): Lusikisiki (-BC), Story 4221 (GRA, PRE).
3227 (Stutterheim): Komgha (-DB), Flanagan 813 (GRA, SAM), Compton 17622 (NBG).
3318 (Cape Town): Stellenbosch, University gardens (-DD), (STE 9010).
3322 (Oudtshoorn): Touws River, George (-CD), Martin 59 (NBG); Wilderness, George (-CD), Comyton 10705 (NBG); 3 ml . From George (-CD), Claasen 3 $\overline{(S T E)}$; Karbonatjieskraal (-CD), H.C. Tarlor 588 (NBG); Silver River, George Forest (-CD), Hall. 206 (NBG); Blanco, George (-CD), Meyer s.n. (STE 11292); George ( $-C D$ ), Paterson $1 \overline{218}$ (GRA).

3324 (Steytlerville): Uitkyk, Witte Els Bosch (-CB), Fourcade 4548 (STE).
3326 (Grahamstown): 10 ml . w. of Grahamstown (-AD), Compton 23389 (NBG); Governors Kop near Grahamstown (-AD), Barker 668 (NBG).
3327 (Peddie): Gonubie, East London ( -BB ), no col.I. (NBG 61923).
3420 (Bredasdorp): Swellendam Dist. ( $-A B$ ), Thode A2398 (NH) ; Maxloth Flower Reserve, Swellendam (-AB), Wurts 467 (NBG).
3423 (Knysna): Belvedere (-AA), Duthie 692 (J), s.n. (STE 15799); Concordia (-AA), Keet 608 (GRA); Barrington, Knysna (-AA), Steyn 75 I ( NB G ) ; The Crags, Knysna (-AA), Compton 23581 (NBG).
$22^{20}$

Fig. 50. Hypoxis sobolifera Jacq. var. pannosa (Baker) Nel - kncwn distribution in Southern Africa up to 1975.

Hypoxis sobolifera var. pannosa (Baker) Nel in Engl. Jahrb. 51: 309 (1914). Type: South Africa, without locality, grown in Kew Gardens (K, holo.!)
H. pannosa Baker in Gard. Chron.: 134 (1874).
H. villosa var. pannosa Baker in J. Linn. Soc. 17:114 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:236 (1893), in Fl. Cap. 6:184 (1896); Ross, Fl. Natal:132 (1972).
H. sobolifera var. accedens Ne1 in Engl. Jahrb. 51:310 (1914).

Leaves 5-14, densely covered with red-brown hairs. Peduncles hairy all over.

Flowering Period: October-April.
H. sobolifera var. pannosa is a predominantly Cape variety occurring in coastal areas in the southern latitudes, but in lower latitudes plants are found at higher altitudes (Fig. 50). Only one specimen is from Natal, namely, Medley Wood 3434, "slopes of Drakensberg".

This variety has no close ally in Natal. The dense covering of red-brown hairs is unique.

Gitations:
var. pannosa

(-AB), Rogers 27963 (STE).
3425 (Skoenmakerskop): Bethelsdorp (-DC), Paterson 370 (GRA).
20. Hypoxis membranacea Baker in J. Linn. Soc. 17:106 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:232 (1893), et in Fl. Cap. 6: 182 (1896); Medley Wood, Fl. Natal:132 (1907); Nel in Eng1. Jahrb. 51:308 (1914); Bews, Fu. Natal \& Zululand: 65 (1921); Ross, Fl. Natal:132 (1972). Type: South Africa, Natal, Tugela, Gerrard 1835 (K, holo.!)

Corm $0,4-0,5 \mathrm{~cm}$ in diam., $1,0-3,5 \mathrm{~cm}$ long, oblong, bristles few, fine. Leaves (4-)5-7(-9), 5,0-14,0 cm long, $0,8-2,2 \mathrm{~cm}$ at widest point, oblong-acuminate, membranous, veins not prominent, covered with scattered, white, stellate haixs, especially on the margins and keel. Peduncles 1-4, hairy. Inflorescence 1-3 flowered (at least one peduncle per plant is more than 1 -flowered). Pedicels $1,5-4,5 \mathrm{~cm}$ long at time of flowering, hairy. Bracts 2 per pedicel, 0,3-0,6 cm long, subulate, setose. Perianth scgments white adaxially, $0,5-0,8 \mathrm{~cm}$ long in open flowers. Filaments usually 3 long, 3 short, sometimes unequal. Anthers 1,5$2,0 \mathrm{~mm}$ long, lanceolate, sagittate, apex bifurcate. Style cylindrical 2,5-3,0 mrn long. Stigna spherical, $\pm 0,5 \mathrm{~mm}$ in diam. Fruit a capsule opening by circumscissile dehiscence below the apex.

## Flowering Period: October-January.

Hypoxis membranacea was first described by Baker in 1878.
In the same year and in the same journal he established another species,


Fig. 5l. Hypoxis membranacea Bak. and Hypoxis parvula Bak. - known distribution in Southern Africa up to 1975.
H. parvula. H. membranacea was said to be 2-flowered, with white perianth segments and two bracts subtending each flower, while $\underline{H}^{H}$ parvula was described as l-flowered, with yellow perianth segments and lacking bracts.

This author in 1896 sank H. parvula in H. membranacea. At this
 with yellow perianth segments and "leaves about 3, ... l-1立 ins. long." In 1914 Nel sank H. brevifolia Bak. in H. membranacea Bak.

In the present study of H. membranacea it became apparent that this species was not homogeneous. Two taxa were represented, one corresponding with Baker's H. membranacea of 1878 , the other with his H. parvula of the same date. H. brevifolia designated a synonym of H. membranacea by Ne1 (1914), falls under H. parvula.

The major differences between H. membranacea and H. parvula are that the latter always has only a 1-flowered inflorescence and usually no bracts subtending the pedicel (if present, there is only one which is short). The perianth segments may be white or yellow (further field study is necessary to determine whether other differences are correlated with these colour forms), while those of H. membranacea are always white. The leaves of H . parvula are smaller and usually less in number than those of H. membranacea, though the range for both species overlaps.

The geographical distribution of the two species is similar (Fig. 51). The range appears to be determined by the cool, moist habitat preference of the species. In the more temperate, southern latitudes, plants are found towards the coast, whereas in the lower Latitudes there is a tendency for thern to be limited to higher altitudes in order to achieve preferred habitat conditions. H. membranacea has a cendency, according to known rccords, to be more coastal and southern;
H. parvula is more limited to higher altitudes further inland.
H. membranacea and Il. parvula are distinct in the genus because of their characteristic stylar and stigmatic form. The style is filiform or cylindrical with a very small ( $\pm 0,5 \mathrm{~mm}$ ), spherical, stigma (Fig. 10) whereas other species in the genus have a pyramidal stigma, equal to or longer than, the subulate or cylindrical style. H. membranacea and H. parvila are also distinct in the form and texture of their leaves, which, as one would expect, are membranous with no prominent veins. $\underline{H}$. angustifolla is the only other species with a leaf of similar texture, but there the leaves are much longer and linear.

## Citations:


21. Hypoxis parvula Baker in J. Linn. Soc. 17:113 (1878), in Dur. \& Schinz, Consp. Fl. Afr. 5:233 (1893). Type: South Africa, Natal, Sanderson s.r., anno 1854 ( K, holo.:)
H. brevifolia Baker in Fl. Cap. 6:183 (1896); Medley Wood, Fl. Natal:132 (1907). Type: South Africa, Natal, Liddesdale, Medley Wood 3940 (NH, holo.!)

Corm 0,3-0,6 cm in diam., oblong, bristles few, fine. Leaves $2-5,2,5-7,0 \mathrm{~cm}$ long, $0,5-1,5 \mathrm{~cm}$ at the widest point, oblong-acuminate or oblong-1anceolate, membranous, covered with stellate hairs on the margins and keel, and simple or stellate hairs on the lamina. Peduncles 1-3, hairy. Inflorescence always l-flowered. Bracts usually absent, if present on $1 \mathrm{y} 1,1,0-4,0 \mathrm{~mm}$ long, membranous, subulate, slightly hairy. Perianth segments yellow or white adaxially, $0,5-1,0 \mathrm{~cm}$ long in open flowers. Filaments filiform, 3 long, 3 short. Anthers 1,0-2,0 man long, oblong-lanceolate, deeply sagittate, apex entire or slightly bifurcate. Style cylindrical or filiform, 3,0-3,5 man long. Stigma spherical, $\pm 0,5$ mm in diam. Fruit a capsule opening by circumscissile deliscence below the apex.

Flowering Period: Septcmber-March (-May).
H. parvula is closely allied to H. membranacea, under which species details of this relationship are given.

Hilliard and Burtt (pers. com.) are engaged upon a revision of the genus Rhodohypoxis Nel. They have established that intergeneric hybridization occurs between Hypoxis parvula and Rhodohypoxis milloides (Baker) Hilliard \& Burtt. Two hybrid specimens are cited here and further information will be available on publication of the above mentioned authors' work.

Of interest also are a number of specimens from the eastern Transvaal and Swaziland, which appears to constitute a distinct taxon probably within the limits of H. parvula, or closely related to this species. Plants are exceptionally small with the leaves not longer than $4,0 \mathrm{~cm}$, and $0,5-1,3 \mathrm{~cm}$ wide. They are nearly always 1 -flowered with the perianth segments yellow and only $2,0-3,0 \mathrm{~mm}$ long. There is usually one bract, $2,0-4,0 \mathrm{~mm}$ long, rarely two, or bracts may be entirely lacking. The anthers are only $\pm 0,5 \mathrm{~mm}$ long.

Their consistent "smallness" is the major reason for considering these specimens as constituting an infraspecific taxon within H . parvula, but the presence of one or two bracts subtending the flower is significan't, as is the observation that one plant possessed a 2-flowered

- inflorescence. No plants of this kind have been found in Natal, and In view of the limited knowledge of this entity, it is considered inadvisable to describe it formally before cited localities have been visited.

Citations:

| TRANSVAAL | - | 2430 | (Pilgrims Rest): Mariepskop (-DB), v.d. Schyff 4818 (PRE). |
| :---: | :---: | :---: | :---: |
| NATAL | - | 2829 | $\begin{aligned} & \text { (Harrismith): Cathedral Peak (-CC), Killick } \\ & 1539 \text { (NPC). } \end{aligned}$ |
|  |  | 2830 | (Dundee): Culvers, Weenen (-CC), Rogers 28312 (STE); Weenen County (-CC), Medley Wood s.n. (SAM 22265). |
|  |  | 2929 | (Underberg): on way to Grotto, Cathkin Park ( -AB ), Howlett 10 ( NH ); Tabamhlope ( -AB ), West 1404 (NI); top of Tabamhlope ( $-A B$ ), Evans |
|  |  |  | 438 (NH); Giants Castle Game Reserve (-AD), |
|  |  |  | Trauseld 683 (NU); Kamberg "Game Pass" (-DB), |
|  |  |  | C.G. Gordon-Gray 66 (NU); Bamboo Mt. Underberg |
|  |  |  | dist. (-CD), Grice s.n. (NU, 11.73). |
|  |  | 2930 | (Pietermaritzburg): Howick (-AC), Themas 17 |
|  |  |  | (SAM) ; N. part of Karkloof forests ( $-\Lambda D$ ), |
|  |  |  | Acocks 11833 (NH, PRE); Karkloof (-AD), Smook |
|  |  |  | 649 (NU); near York (-AD), Medley Wood 877 |
|  |  |  | (PRE); Umgeni Poort (-CA), Mo11 1362 (NU); |
|  |  |  | Pietermaritzburg (-CB), Fisher 724 ( $\mathrm{NH}, \mathrm{NU}$ ) ; |

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                                    Swartkop (-CB), Hillary 60 (NU); Liddesdale
                                    (-CB), Medley Wood 3940 (NH).
CAPE - 3029 (Kokstad): Ensekeni (Insikeni) (-BA), Medley
                            Wood 12077 (NH).
                            3228 (BuLterworth): Kei Mouth (-CB), Bokelmann 9
                            (NBG).
without precise locality:
                                    Thode 5293 (STE), Springfontein, Natal Province.
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taxon nov.
TRANSVAAL - 2330 (Tzaneen): Woodbush mountains (-CC), Moss
15429 (J).
2430 (Pilgrims Rest): near God's Window, Pilgrims
Rest area (-DD), Davidson 792 (J).
SWAZILAND - 2631 (Mbabane): Mbabane (-AC), Compton 27243, 29335
(NBG).
without precise locality:
Kerfoot 6459 (J), Belvedere, E. Tvl.
Putative hybrid: Hypoxis parvula $x$ Rhodohypoxis milloides
NATAL - 2929 (Underberg): Impendhle distr., farm "Ungeni
Vlei" (-CD), Wright 1600 (NU).
2930 (Pietermaritzburg): Fort Nottingham (-AA),
Wright 1567 (NU).

Indeterminate specimens

At the conclusion of this study, five herbarium sheets remained that could not be placed with certainty. It is believed that plants of Hypoxis can undergo hybridization especially within the genus. The "aberrant" specimens represented on these sheets may be the resuit of this phenomenon. The specimens are listed below together with an indication of the species they most closely resemble.

1. Coetzee 14 (NH), "Shawlands", Nottingham Road: - H. acuminata, but lamina glabrous.
2. Grice (NU, Dec. 1973), Bamboo Mt., Underberg: - H. oblonga, but 2 prominent nerves not apparent, hairs different, flowers pedicelled.
3. Trace 30 (NU), Wembly, Pietermaritzburg: - resembling H. rooperi - a putative hybrid of this species?
4. S.E. Wood 146 (NU), corner Estcourt highway and Winterton turn off: - H. kraussiana, but broader leaves, not subterete, nor as hairy, with different venation.
5. Thienel 12 (NU), Mariannhill: - ?

## CHAPTER 3

## DISCUSSION

The Natal species of Hypoxis are predominantly grassveld plants, but they have a wide habitat tolerance, for plants may occur on coastal sand dunes or on the fringes of marshy vleis.
H. membranacea, H. parvula and H. angustifolia var. buchananii do not exhibit this tolerance however, and grow only in damp areas. These are the only species with roembranous leaves. H. membranacea and H. parvala are also distinguished from other species in the genus because of their distinct stigmatic and stylar form (see Diagnostic Characters p. 23). The remainder of the Natal species are grassveld taxa. They are generally robust plants with hairy leaves. H. latifolia and H. interjecta, although falling within this group, have leaves that are usually glabrous. In no known plant of H. interjecta have the leaves been hairy, but in one plant of
N. Natal
H. latifolia (from Ehe-Tretrofeat) the leaves were covered in hairs. This probably indicates that all taxa have the genetic potential to develop hairy leaves. H. latifolia is distinct within the genus by virtue of its leaf size and form, having no close ally. Thus within the Natal species there are two small, isolated groups, still geophytic, but distinguished morphologically from the main section of grassveld forbs.

In South Africa many grasslands are subject to annual burning in winter. It is once this has occurred that plants of Hypoxis become obvious. No proof is available of the factors that stimulate the renewal of growth, but many hypothetical explanations, that are
supported by observation, have been suggested. This renewal of growth may depend on exposure of the soil to the sunlight (absorbtion of heat), or on the direct clearing of the old vegetation and with it unsatured hydrocarbons (ethylene) derived from it, that may possibly have acted as suppressors (alteration of the concentrations of these gases in the ambient and soil atmosphere may suppress then stimulate growth, but this needs experimental confirmation). Abeles, 1973, gives some evidence that ethylene may, in some cases suppress, and in other cases stimulate, the renewal of growth in underground organs (tubers and corms).

Bews (1925) in his early descriptive work on the vegetation of Natal, commented on growth renewal in grassland geophytes as follows:

> "In spring many grassveld plants commence growth even before the first rains, as soon as the temperature rises slightly, a fact which demonstrates that it is not in all cases the winter drought which causes the resting period in South African plants."

However, whether the veld is burnt, or the vegetation cleared by cutting, in autumn, winter, or early spring, the effect is the same. There is a need for physio-ecological studies in order to determine the factors that control the life-cycle of geophytes of the South African grassveld particularily the initiation of inflorescence development and the renewal of aerial growth.

Hypoxis plants rarely compete with other vegetation such as the grasses, despite growing with them. Flowering commences with the regeneration of aexial shoots, and often continues over a comparatively short period only so that on the same plant,flowers and dehisced capsules containing mature, black seeds may frequently be seen. Leaf growth continues throughout the growing season so that the new, short spring growth gives place in full sumer, in most cases at any rate,
to elongate leaves as tall as the associated grasses, (for example H. rigidula where leaves may range from 10 cm to 1 metre over the growing season).

Where plants occupied habitats where there was no competition, flowering took place all year. For example, H. gerrardii flowered all year on a north facing, rocky slope near Highflats. This strongly suggests that plants may have a potential to flower continuousiy, but that few individuals realize this potential.

As Hypoxis flowers are mostly bright yellow and clearly evident when the vegetation is sparse, collecting of herbarium material has been mainly during the short period of growth renewal.

Consequently this short period has often been taken to represent the range over the complete growing period. It is this that has led previous authors (who have relied predominantly on herbarium collections), to delimit leaf length in terms of restricted ranges. Thus when a specimen to be identified falls outside the range of possibilities covered by the key, confusion results.

There is evidence that plants of the genus undergo both interspecific and intergeneric hybridization on occasions. Detection of interspecific hybridization is difficult because many of the species are similar morphologically and diagnostic characters exhibit wide ranges of variation.

Two cases of putative interspecific hybridization were observed. One was detected from herbarium specimens, namely, $H$. angustifolia var. buchananii x H. membranacea (for details see page 89). The other case involved plants of $\underline{H}$. acuminata and $\underline{H}$. angustifolia var. buchananii, which were growing sympatrically. Specimens were found which exhibited


Fig. 52. H. acuminata, H, angustlfolia var. buchananii and putative hybrid:
a - H. acuminata, b - putative hybrid, c - H. angustifolia var. buchananii


Fig. 53. Enlargement of leaf and inflorescence of each plant shown in Fig. 52
a range of characters between those of the two putative parents. Leaves of these plants were similar to $H$. acuminata in shape and hairiness (Fig. 52), while inflorescences were intermediate (Fig. 53).

Intergeneric hybridization has been proved between Rhodohypoxis baurii and Hypoxis parvula (Hilliard pers. com.; see page 101). The occurrence of intergeneric hybridization strongly suggests that interspecific hybridization will also be possible, but this last cannot be proved without experimentation which needs to be undertaken.

This thesis has attempted to clarify the taxa within Hypoxis that exist naturally in Natal. Species have been redescribed in more detail, with careful checking for the presence of synonymous taxa. H. colchicifolia and H. oligotricha have become junfor synonyms of H. latifolia; H. elliptica of H. rigidula var. rigidula; H. nitida of H. obtusa; and H. woodii of H. angustifolia var. buchananii. Several taxa have not been recorded in Natal before, namely, H. galpinii, H. interjecta and H. costata. One new taxon has been established, H. zululandensis. Concluding this assessment a key to the species was constructed, using predominantly vegetative characters.

Further study (especially field work outside Natal) is required to clarify the relationships among some closely related taxa, for example H. gerrardii and $H$, argentea; or to elucidate aberrant specimens within individual species, for example H. 1atifolia and H. obtusa. Further field work outside Natal is also required to explore the extent of variation over their whole distributional range of widespread species such as H. costata and H. multiceps.

No attempt has yet been made to study the floral biology within this genus. The nature of the pollinating vectors is of particular interest. Outbreeding must occur since there is evidence of hybridization. Apomixis is also suspected to occur. Is this an aftermath of hybridization? Cytological studies are necessary to confirm the basic chromosome number for Hypoxis, and to throw more light on polyploidy within the genus.

Chemical and physio-ecological work on the corms, and detailed study of their growth patterns, including the functioning of the contractile roots, should prove interesting, as should studies on seed germination.

With collaboration in all fields of botanical study, a considerable contribution to the knowledge of the genus Hypoxis in southern Africa should be possible.

## SUMMARY

A revision of the Natal species of Hypoxis was undertaken. Some of the taxa represented in Natal extend beyond this province, and consequently work on these included the whole area of their distribution in Southern Africa.


#### Abstract

A history of the genus is given, together with details of world distribution. A review of cytological literature relating to the genus is also provided.


Study of specimens from a number of South African, and some European Herbaria, provided the founation of this revision. This was supplemented by both field work and laboratory investigations, Including the use of a Scanning Electron Microscope.

Diagnostic criteria used to delimit taxa are enumerated and critically considered. A key to the identification of species and infraspecific taxa, based predominantly on vegetative characters is provided. These characters are discussed in detail.

Detailed descriptions of the species are given, together with a range of flowering periods and brief habitat notes. Twenty one species are listed for Natal. During the course of the study, H. colchicifolia and H. oligotricha were found to be junior synonyms of H. latifolia; $H$, nitida of H . obtusa; and H . woodii and H. obligua var. woodii of H . angustifolia var. buchananii. H. parvula was reinstated as a species (previously regarded as synonymous with
H. membranacea), and one new species, H. zululandensis was established. Two taxa, not previously known in Natal, H. galpinii and H. interjecta were recorded for this province.

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## APPENDIX <br> INDEX OF NUMBERED COLLECTIONS

This index provides an alphabetical list of collectors, their collections, the herbaria in which the collections are housed and the names of the specimens. The collector's name is followed by the collector's numbers which are grouped in the sequence of the taxa as they are enumerated below. In this enumeration each taxon has a reference number.

Example: BEWS $650(N U=2)$ means the specimen was collected by BEWS, Universily number 650, and housed in the Natal Herbarium. It is Hypoxis galpinii Bak.

The herbarium abbreviations are those of Holmgren \& Keuken (1974), except for the Natal Parks Board, Cathedral Peak which is designated NPC. If no collectors number exists reference is then made either to the herbarium number or to the date of collection.

1. Hypoxis latifolia Hook.
2. Hypoxis galpinii Bak.
3. Hypoxis interjecta Nel
4. Hypoxis multiceps Buching.
5. Hypoxis costata Bak.
6. Hypoxis rigidula Bak. var. rigidula

6b. Hypoxis rigidula Bak. var. pilosissima Bak.
7. Hypoxis acuminata Bak.
8. Hypoxis oblonga Nel
9. Hypoxis obtusa Burch.
10. Hypoxis rooperi S. Moore
11. Hypoxis zululandensis S.E. Wood
12. Hypoxis filiformis Bak.
13. Hypoxis neliana Schinz
14. Hypoxis kraussiana Buching.
15. Hypoxis gerrardii Bak.

16a. Hypoxis argentea Bak. var. argentea
16b. Hypoxis argentea Bak. var. sericea Bak.
17a. Hypoxis argustifolia Lam. var. argustifolia
17b. Hypoxis argustifolia Lam. var. buchananii Bak.
18. Hypoxis lata Nel

19a. Hypoxis sobolifera Jacq. var. sobolifera
19b. Hypoxis sobolifera Jacq. var. pannosa (Baker) Nel
20. Hypoxis membranacea Bak.

21a. Hypoxis parvula Bak. var. parvula (will be established at time of publication of var. nov.)

21b. Hypoxis parvala Bak. taxon nov. (not established)

Putative hybrids:
H1. Hypoxis angustifolia Bak. var. buchananii Bak. x Hypoxis membranacea Bak.

H2. Hypoxis parvula Bak. var. parvala x Rhodohypoxis milloides (Baker) Hilliard \& Burtt
A. ACOCKS 10606 ( $\mathrm{NH}=4$ ), 10604, 10641, 10833 ( $\mathrm{NH}=6 \mathrm{a}$ ), 16275 ( $\mathrm{PRE}=6 a$ ) , $11771,10587(\mathrm{NH}=12)$, $9301(\mathrm{PRE}=12)$, 12960 ( $\mathrm{PRE}=$ 16b), 10832 ( $\mathrm{NH}=17 \mathrm{a}$ ) , 9331 ( $\mathrm{PRE}=17 \mathrm{a}$ ) , 11833 ( $\mathrm{NH}, \mathrm{PRE=21a);}$ ALLSOPP 909 ( $\mathrm{NH}, \mathrm{NU}=1$ ), 520 ( $\mathrm{NU}=1$ ), 920 ( $\mathrm{NH}, \mathrm{NU}=6 \mathrm{a}$ ), 682 $(\mathrm{NH}=10), 411,911(\mathrm{NH}=14), 728,921(\mathrm{NH}, \mathrm{NU}=15)$; ARNOLD $841(\mathrm{NU}=4), 835,837,838(\mathrm{NU}=6 \mathrm{a}), 839(\mathrm{NU}=7), 836,842$, $(N U=9), 834,840 \quad(N U=10)$.
B. BAIJNATH 187 ( $\mathrm{NU}=17 \mathrm{a}$ ); BAUDERT 24 ( $\mathrm{GRA}=15$ ); BARKER 2585 (NBG=2), 220, 7952 (NBG=6a), 3428-2 sheets, 4367 (NBG=10), 5162 (NBG=15), 2124 (NBG=16a), 7951 ( $\mathrm{NBG}=17 \mathrm{a}$ ), 688 ( $\mathrm{NBG}=19 \mathrm{a}$ ); BATTEN 6 (NBG, PRE=10); BAUR 904 (SAM=9); BAYER \& MCCLEAR 196 (GRA, PRE=19a); BAYLISS 2547 (SAM=15), 3138 (SAM=16a); BEETON 27 ( $\mathrm{NBG}=7$ ) ; BELL s.n. (NH 14151=17b) ; BEWS 650 (NU= $2), 193(N U=6 a), 8(N U=12) ; \quad B 0 K E L M A N N 1,3(N B G=17 a), 9$ (NBG $=21 \mathrm{a})$; BOLUS $65814(\mathrm{NBG}=6 \mathrm{a}), 181$ (PRE, STE=9); BOOYSEN 1707 ( $\mathrm{NBG}=9$ ) ; BOUROUIN 458 ( $\mathrm{NU}=10$ ); BOUWER 6055 (PRE=9), 2229 ( $\mathrm{PRE}=10$ ) ; BRITTEN 4653 (GRA=10); BRUYNS-HAYLETT 89 (NU=15); BUITENDAG 890 (NBG=6a), 158 ( $\mathrm{NBG}=6 \mathrm{~b}$ ) , 619 ( $\mathrm{NBG}=9$ ); BURTTDAVY 13457 ( $\mathrm{PRE}=1$ ) , 3823 ( $\mathrm{PRE}=6 \mathrm{a}$ ), 682 ( $\mathrm{NH}=6 \mathrm{~b}$ ).
C. CAIN s.n. (J 37931=2); CAIRNS s.r. (NU 8.8.55=8) ; CIAASEN 3 (STE=19a); CLIVER 403 ( $\mathrm{NH}=7$ ) ; CODD 9396 (PRE=2), 8049 ( $\mathrm{PRE}=7$ ) , 5673 ( $\mathrm{PRE}=10$ ) , 5750, $6659 ~(\mathrm{PRE}=12), 1849$ ( $\mathrm{PRE}=16 \mathrm{~b}$ ); CODD \& DE WINTER 3282 (PRE=2), 3361 (PRE=7); COHEN s.n. ( $J$ 18.9.1936=6a); COLEMAN 193 ( $\mathrm{NH}=6 \mathrm{~b}$ ), 159 ( $\mathrm{NH}=10$ ), 465 ( $\mathrm{NH}=16 \mathrm{~b}$ ); COMINS 304 ( $\mathrm{NJ}=10$ ); COMPTON 21449 ( $\mathrm{NBG}=1$ ), 22371, 26341, $27095(\mathrm{NBG}=2), 30881$ ( $\mathrm{NBG}, \mathrm{PRE}=2$ ), 27121 ( $\mathrm{NBG}=4$ ), 21311, 24490, 24578, 25210 ( $\mathrm{NBG}=5$ ), 32393 ( $\mathrm{NBG}=6 \mathrm{a}$ ), 24492, 26342, 26924, 26988, 28102 (NBG=6b), 24551, 25179, 26071, 27182, 28279, 28289 ( $\mathrm{NBG}=7$ ), 26128, 31728 (NBG, PRE=7), 22360 ( $\mathrm{NBG}=9$ ) , 24706, 27337, 28104, 28843, 31790 ( $\mathrm{NBG}=10$ ), 19724, 24518, 25139, 27149, 28249, 28281 (NBG=12), 26129, 29266 ( $\mathrm{NBG}, \mathrm{PRE}=12$ ), 27211 ( $\mathrm{NBG}=15$ ), 19217 ( $\mathrm{NBG}=17 \mathrm{a}$ ), 17668, 29340 ( $\mathrm{NBG}=17 \mathrm{~b}$ ), 21365 ( $\mathrm{NBG}=18$ ) , 10705, 17622, 23389, 23581 ( $\mathrm{NBG}=19 \mathrm{a}$ ) , 27243, 29335 ( $\mathrm{NBG}=21 \mathrm{~b}$ ), 17716 ( $\mathrm{NBG}=\mathrm{Hl}$ ); COTTERRELL 63 (GRA=16a); COURTENAY-LATMER s.n. (NBG, Oct. 1945=10); CROOK 5 ( $\mathrm{NH}=6 \mathrm{a}$ ).
D. DALY 78 (PRE, 2 sheets $=10$ ), 766 (GRA=16a); DALY \& CHERRY 977 (GRA=16a); DAHLSTRAND 1793 (STE=17b); DAVIDSON 484, 2007, $2450(\mathrm{~J}=12), 2548(\mathrm{~J}=20), 792(\mathrm{~J}=21 \mathrm{~b})$; DAVIDSON \& MOGG 33288 ( $\mathrm{J}=12$ ) ; DE VILLIERS 5 ( $\mathrm{NU}=17 \mathrm{a}$ ); DE VRIES 85 ( $\mathrm{PRE}=17 \mathrm{~b}$ ); DENY s.n. (J 35104=2) ; DEVENISH 1521, 1521a (NU $=12$ ), 181, 691, 766 ( $\mathrm{PRE}=12$ ) , 1174 ( $\mathrm{NH}, \mathrm{PRE}=14 \mathrm{a}$ ) ; DIETERLEN 149 (PRE, SAM=4), 2296 ( $\mathrm{NH}, \mathrm{PRE}, \mathrm{SAM}=5$ ) , 223, 6971 ( $\mathrm{SAM}=6 \mathrm{a}$ ), 1370 (PRE, SAM=6a), $230(\mathrm{NH}=6 \mathrm{~b}), 290(\mathrm{PRE}, \mathrm{SAM}=7), 709(\mathrm{SAM}=7), 223(\mathrm{NH}=9), 310$ ( $\mathrm{SAM}=9$ ), 230 (PRE, $\mathrm{SAM}=9$ ), 229 ( $\mathrm{SAM}=10$ ), 229a ( NH, PRE, SAM - 2 sheets $=10$ ), $650(\mathrm{NH}, \mathrm{SAM}=12), 176(\mathrm{NH}, \mathrm{SAM}=16 \mathrm{a}), 536$ (PRE=16b), s.n. (PRE 33599=16b), 639 (NH, PRE, STE=17a), 6968 (SAM $=17 \mathrm{a}), 536$ (SAM=17b); DIXON 6 (NU=10); DOUWESDEKKER 11 ( $\mathrm{NU}=10$ ) ; DOWNING 31 ( $\mathrm{NU}=6 \mathrm{a}$ ); DUTHIE 735 ( $\mathrm{J}=17 \mathrm{a}$ ), 692 (J=19a), s.n. (STE 15799=19a) s.n. (STE 22022=19b); DU TOIT s.n. (NU Aug. 1957=7); DUTTON 54 (PRE=17b); DYER 298 ( $\mathrm{PRE}=17 \mathrm{~b}$ ).
E. EDWARDS 2407 ( $\mathrm{NU}=10$ ) ; EVANS 438 ( $\mathrm{NH}=2 \mathrm{la}$ ).
F. FAIRALL $110(\mathrm{SAM}=10), 57(\mathrm{SAM}=15)$; FAKUDE 15 ( $\mathrm{NH}=15$ );

FARQUAMARSON 3 (NU=10); FELDUAN 8.r. (NU 7.7.57 2 7) ; FISCHER $81(\mathrm{NU}=10), 724(\mathrm{NH}, \mathrm{NU}=21 \mathrm{a})$; FLANAGAN 1173 (GRA, SAM=4), $1171,2842(S A M=6 a), 1170(S A M=6 b), 1642(S A M=9), 1810$ (PRE, SAM=10), 1811, 2771 (PRE, SAM=12), 1174 (SAM=17a), 813 (GRA, $S A M=19 a), 1172$ (GRA - 2 sheets, PRE, SAM=20); FORBES s.n. (STE 12803=10); FOURCADE 4548 (STE=19a); FRANKS s.n. (NH $14174=17 b$ ) 。
G. GALPIN 1098 (GRA, NH, PRE, SAM=2), 1058, 1603 (PRE=4), 8427 ( $\mathrm{PRE}=5$ ) , 1604 ( $\mathrm{GRA}, \mathrm{PRE}=6 \mathrm{a}$ ), $1099(\mathrm{PRE}=6 \mathrm{~b}), 412$ ( $\mathrm{GRA}, \mathrm{PRE}=9$ ), 1605 ( $\mathrm{PRE}=9$ ), 1101 (GRA, NH, $\mathrm{PRE}, \mathrm{SAM}=12$ ), 10244 ( $\mathrm{PRE}=12$ ), $13840(\mathrm{PRE}=16 \mathrm{~b}), 3343(G R A=17 \mathrm{a}), 3085(\mathrm{GRA}=19 \mathrm{~b}), 5810$ (SAM= 20); GARABEDIAN s.n. (NBG 49882=17a); GEEKIE 41 (NU=10); GERRARD \& MCKEN 1826 ( $\mathrm{NH}=6 \mathrm{~b}$ ) , 1828 ( $\mathrm{NH}=10$ ), 1827 ( $\mathrm{NH}=15$ ); GERSTNER s.r. (NH 28986=6a), 3855 ( $\mathrm{NH}, \mathrm{PRE}=6 \mathrm{~b}$ ) , 2382, 3588 $(\mathrm{NH}=7), 2590,3638(\mathrm{NH}=10), 3692(\mathrm{NH}=11), 4936$ ( $\mathrm{PRE}=11$ ), s.n. (NH 22261=17b); GILLET 1170 (STE=5), s.n. (STE 30403=9), 2316 (STE=16a), 1262 (STE - 2 sheets=19b); GILLILAND 26911 (PRE=2), 26127 ( $J=3$ ); GLASS 376 (SAM=10), 378 (SAM=16a); GLEN 274 ( $\mathrm{NH}=20$ ) ; GODFREY s.n. (PRE 33555=9) ; GOOSENS 289 (PRE=2), $91(\operatorname{PRE}=3), 138(P R E=4), 779$ (PRE=10); GORDON-GRAY C.G. 4 J.L. $117(\mathrm{NU}=6 \mathrm{a}), 1438(\mathrm{NU}=10), 65(\mathrm{NU}=12), 107(\mathrm{NU}=15), 696$ $(N U=17 a), 66(N U=21 a), 949$ (NU=H1); GORDON-GRAY K.D. 9950 ( $N U=$ 6a), 10011 ( $N U=8$ ); GRICE s.n. ( NU Dec. 1973=12) , s.n. ( NU 11.1973=2la); GUIfLARMOD 344B (PRE=12); GUNN_s.n. (PRE $33557=9$ ); GUTHRIE 4387 ( $N B G=19 b$ ); GUY \& JARMAN 84 (NU=10).
H. HAAGNER s.n. (GRA Oct. 1904=10); HALL 16 (NU=17b), 206 (NBG= 19a); HALSE 43 (NH=6b); HALT 27 (NH=16b); HARDS 19 (PRE= 10); HARRISON $245(\mathrm{NH}=6 \mathrm{~b}), 183(\mathrm{NH}=10), 21(\mathrm{NH}=12)$; HART 29 (NU=10); HAYGARTH s.n. (STE Oct. 1921=4), 18 (STE=10); s.n. (NH 12358=12), 76 (STE=15), s.n. (NH 9923=17a), 79 (STE= 17b), s.n. (PRE 22309=17b); HERBST s.n. (NBG 78441=1);
HEYDOORN 45 (STE=6b); HILLARY 60 (NU=2la); HILLIARD 5543 ( $\mathrm{NU}=17 \mathrm{~b}$ ) , s.n. ( NU 28395 m 20 ) ; HILNER 82 (GRA=17a) ; HITCHINS 71 (PRE=12); HODSON $7 \quad(N U=2)$, $10(N U=14)$; HOFSTROM \& ACOCKS

251 ( $\mathrm{PRE}=9$ ) ; HONE 28 (PRE=12); HOWLETT 9 ( $\mathrm{NH}=15$ ), 10 ( $\mathrm{NH}=$ 21a) ; HUNTLEY 247 (NH, NU=10); HUTCHINSON, FORBES \& MCCLEAN $31(\mathrm{NH}=10), 52(\mathrm{NH}=15)$; HUTCHINSON, FORBES \& VERDOORN 133 ( $\mathrm{NH}=13$ ) .
I. I'ONS 63/19 (NBG=9); IRWIN 18 (NU=15).
J. JACOBSZ 328 (PRE=5), 329 (PRE=6a), 141 (PRE=16b); JENKINS $10202(G R A=9), 7524(G R A=12), 10686(P R E=16 a) ;$ JOHNS s.n. (NBG $11.4 .1942=19 \mathrm{~b}$ ) ; JOHNSTONE 528 (NU=5); JOLLIFFE 52 (NU= 14); JUNOD 25453 ( $\mathrm{PRE}=17 \mathrm{a}$ ).
K. XEET 608 (GRA=19a); KEMP s.n. (NBG 65826=9); KERFOOT 6459 ( $\mathrm{J}=2 \mathrm{lb}$ ) , 7225 ( $\mathrm{J}=\mathrm{H} 1$ ) ; KJLLICK 1573 ( $\mathrm{NPC}=5$ ), 1574 ( $\mathrm{NPC}=7$ ), 1113 ( $\mathrm{NPC}, \mathrm{PRE}=7$ ) , 967 ( $\mathrm{NPC}, \mathrm{PRE}=12$ ), 225 ( $\mathrm{NU}=15$ ), 1539 ( $\mathrm{NPC}=$ 21a) ; KILLICK \& STREY 2548 (PRE=7); KILLICK \& VAHRMRIJER 4048 ( $\mathrm{NH}=12$ ); KOK 210 ( $\mathrm{PRE}=6 \mathrm{a}$ ); KOTZE 507 (PRE=10); KOWARSKY 12082 ( J=12).
L. LACHIN.SN 7 (NU=10); LAMBERT s.r.(J 24471=9); LANDSDELL s.g. (NH 34266=15) ; LANG F.R. 177 (GRA=10); LANG H. s.n. (NH 27190 =7); LAUGIITON s.n. (J 34644=9); LAWN 1297, 1788 (NH=6a), 335, 446, $1161(\mathrm{NH}=10), 53,1187(\mathrm{NH}=12), 783,1019,1019 \mathrm{a}$, 1684, 1746 ( $\mathrm{NH}=15$ ), 1188 ( $\mathrm{NH}=19 \mathrm{a}$ ) ; LAWSON 1220 ( $\mathrm{NH}=10$ ), 1219 (NH, PRE=15); LEEMAN s.n. (PRE 33554=10); LEENDERTZ 11970 ( $\mathrm{PRE}=3$ ) , 7746 ( $\mathrm{PRE}=4$ ), 10203 ( $\mathrm{PRE}=9$ ) , 7350 ( $\mathrm{PRE}=12$ ), 7892 ( $\mathrm{PRE}=15$ ) ; LEISGANG 46 ( $\mathrm{NU}=10$ ) ; LEVETT 101 ( $\mathrm{NH}=5$ ) , 40 ( $\mathrm{NH}=$ 13), 25 ( $\mathrm{NH}=15$ ); LEWIS 6184 (STE=6a), 6184 ( $\mathrm{NBG}=6 \mathrm{~b}$ ); LIEBENBERG 6956 ( $\mathrm{PRE}=5$ ) , 7329 ( $\mathrm{PRE}=9$ ) , 3306 ( $\mathrm{PRE}=17 \mathrm{~b}$ ); LUCAS s.n. (J 40087=16a); LUSSEM 50 (NBG=17a).
M. MACNAE 1339 ( $\mathrm{J}=6 \mathrm{a}$ ), 1496 ( $\mathrm{J}=9$ ); MACOWAN 104 (GRA=4), 104 $(S A M=5), 1649 \quad(G R A=6 a), 1649 \quad(S A M=9), 50 \quad(G R A, S A M=16 a)$, 1899 (GRA, SAM=19b); MAGUIRE s.n. (J 25555=9), s.n. (J 25554 =12) ; MALAN s.n. (SAM 58324=4), s.n. (SAM 58326=17a); MARKOTTER s.n. (STE 16304=6a), 8851 (STE=9), 8692 (STE=15) 8.n. (STE 19066=17b); MARTIN A.J. 663 (NU=12); MARTIN B.E. $476(N B G=12), 59,475(N B G=19 a) ;$ MATTISON $8(N J=6 a)$; MAUVE
$29(\mathrm{~N} U=10), 4504$ (J, STE=12); McCALLUM 323 (PRE=6a); McCLEAN 905 ( $\mathrm{NH}=7$ ); McGILLET 1306 (STE=17a); McKEN 1046 ( $\mathrm{NH}=12$ ); McKEOWN 54 ( $\mathrm{NU}=12$ ); MCMURTY C3( $\mathrm{NBG}=6 \mathrm{a}$ ); MEDLEY WOOD 1170 ( $\mathrm{NH}, \mathrm{SAM}=1$ ), 5193 ( $\mathrm{NH}=1$ ), $373(\mathrm{NH}=2)$, $1011 \quad(\mathrm{NH}=4)$, s.n. (SAM $22296=4$ ), 3456, 3870 ( $\mathrm{NH}=6 \mathrm{a}$ ), 754,2627 (SAM=6a), 2463 ( $\mathrm{NH}=$ $6 \mathrm{~b}), 1347$ ( $\mathrm{NH}=7$ ), 2435, 4372 ( $\mathrm{NH}=8$ ), 6922 ( $\mathrm{NH}=10$ ), 47 (SAM=10), $1030(\mathrm{NH}, \mathrm{SAM}=12), 4660(\mathrm{PRE}, \mathrm{SAM}=12), 327(\mathrm{NH}=15), 101 \quad(\mathrm{SAM}=$ 16a), 6633 ( $\mathrm{NH}=17 \mathrm{a}$ ), 9649 (SAM=17a), 426, 771 ( $\mathrm{NH}=17 \mathrm{~b}$ ), 3434 ( $\mathrm{NH}=19 \mathrm{~b}$ ), 862 ( $\mathrm{NH}=20$ ) , 3940 , 12077 ( $\mathrm{NH}=21 \mathrm{a}$ ), 877 (PRE=2la), s.n. (SAM 22265=2la); MEYER s.n. (STE 11292=19a); MILDRED 2 (NBG=6a), 1 ( $\mathrm{NBG}=9$ ); MLLLER 202 ( $\mathrm{NH}=1$ ); MILNER s.n. ( $\mathrm{NH} 23260=15$ ); MNGOMEZULU 7 ( $\mathrm{NH}=17 \mathrm{a}$ ); MOGG 3240 ( $\mathrm{PRE}=6 \mathrm{a}$ ); 3189, 5720, 5572 (PRE=7); 272, 3281-2 sheets, 935 (PRE=9); 3518 ( $\mathrm{PRE}=15$ ), 3449, 3479 ( $\mathrm{PRE}=16 \mathrm{~b}$ ), 5880 ( $\mathrm{NH}=17 \mathrm{a}$ ), 13930 ( $\mathrm{PRE}=17 \mathrm{a}$ ), 881 ( $\mathrm{PRE}=17 \mathrm{~b}$ ), 4486 ( $\mathrm{PRE}=19 \mathrm{a}$ ); MOLL 1044 (NU, PRE=4), $1255(N H=6 a), 1040(N U=6 a), 1255$ ( $N J=7$ ), 4740 (SAM= $11), 2545$ ( $\mathrm{NU}=12$ ), 3394 ( $\mathrm{NU}=15$ ), 1060 ( $\mathrm{NU}=17 \mathrm{a}$ ), 1362 ( $\mathrm{NU}=21 \mathrm{a}$ ); MOLL \& STREY 3807 ( $\mathrm{NH}=10$ ); MORRIS 365 ( $\mathrm{NU}=10$ ), 366 (NU=14); MOSS 13679 ( $\mathrm{J}=1$ ), 3332, 14026, $17720(\mathrm{~J}=2)$, 13585 ( $\mathrm{J}=4$ ), 12080 ( J=6b), 5211, 13889, 17713 ( $\mathrm{J} \approx 7$ ), 2556, 2655, 16184 ( $\mathrm{J}=9$ ), s.n. (J 22.10.27=9), 15428, 17125, 18515 ( $\mathrm{J}=12$ ), 5576, 11932, 13492, 17676 ( $J=16 \mathrm{a}$ ), 13981 ( $\mathrm{J}=16 \mathrm{~b}$ ), 15016,17328 , 19744 (J=17a), 5574, 17124 (J=17b), 16037 ( J=19b), 15429 ( $\mathrm{J}=2 \mathrm{lb}$ ); MOSS \& RUGERS 996 ( $\mathrm{J}=16 \mathrm{~b}$ ); MÜLLER 333, 369 (NBG= 17a).
N. NASS s.n. (NBG 63086=17a); NELSON 11768 (PRE=10); NICHOLSON 797 ( $\mathrm{PRE}=1$ ), 238 ( $\mathrm{NH}=10$ ), 713 (PRE=20); NIEUWOUDT 33 ( $\mathrm{NU}=15$ ); NOEL 1485 (GRA=17b).
O. OBERMEYER 37 ( $\mathrm{NH}=6 \mathrm{a}$ ).
P. PATERSON $8 . \mathrm{n}$. (GRA Oct. $1911=10$ ), 1218 (GRA=19a), 370 (GRA= 19b); PEACOCK s.n. (SAM 65759=6a), s.n. (SAM 65757=17b); PEGEL Jhb. 1 ( $\mathrm{NH}=6 \mathrm{a}$ ), s.n. ( $\mathrm{NH} 23.1 .68-5$ sheets $=9$ ); PEGLER 1143 ( $\mathrm{PRE}=10$ ), 814 ( $\mathrm{PRE}=17 \mathrm{a}$ ), 109 ( $\mathrm{PRE}=20$ ); PELLATT 2 ( $\mathrm{NU}=8$ ); PENNEFATHER 28 (NU=15); PHILLIPS E. 932 (SAM=16b), в.n.
$(S A M 6755=17 a)$, s.n. (SAM $6755=17 b) ;$ PHILLIPS J. s.n. (J 32949=7); PIEK 89 (NH=6a); PIENAAR 16, 74 ( $\mathrm{NU}=10$ ), 72 ( $\mathrm{NU}=12$ ) ; PONT 184 ( $\mathrm{PRE}=6 \mathrm{a}$ ) ; POOLEY 662 ( $\mathrm{NU}=10$ ); POTGIETER
 =16b); PRONSER s.n. (J 29380=9).
R. RAM s.r. (NU 2.3.74=15), s.n. (NU - 2 sheets=17a), 508 ( $N U=$ 17b); RANDLES 55 (NU=4); RATTRAY 117 ( $\mathrm{GRA}=20$ ) ; RENNIE 610 ( $\mathrm{NU}=9$ ) , 91 ( $\mathrm{NU}=15$ ); RENSBURG 25 ( $\mathrm{NU}=7$ ) ; REPTON 3512 ( $\mathrm{PRE}=12$ ) , 2763 ( $\mathrm{PRE}=16 \mathrm{a}$ ); REYBURN s.n. (PRE 14.11.66=20); ROBERTS 2388 ( $\mathrm{PRE}=16 \mathrm{~b}$ ); ROBERTSON C. 12 ( $\mathrm{NU}=14$ ); ROBERTSON R. s.n. (PRE 33558=6a), 2 (PRE=9); ROGERS 27505 ( $\mathrm{J}=2$ ), 28267 $(S T E=2), 27547(S T E=4), 23539(J=6 a), 27786(S T E=6 a), 237$ $(G R A=6 b), 11561(G R A=7), 24612,28367$ ( $\mathrm{STE}=10$ ), 21416 ( $\mathrm{GRA}=12$ ), 24583, 27546 (STE=15), 27404, 28367 (STE=16a), 2062, 24665 $(S T E=17 a), 27963(S T E=19 b), 28312$ (STE=21a); ROSS 1305 ( $N=4$ ), 1458 ( $\mathrm{NU}=6 \mathrm{a}$ ) , $2157(\mathrm{NH}=10), 2159$ ( $\mathrm{NH}, \mathrm{PRE}=15$ ), 45 ( $\mathrm{NH}=16 \mathrm{~b}$ ); ROSS \& MOLL 2281 ( $\mathrm{NH}, \operatorname{PRE}=10$ ) ; RUCH 1477 (PRE=13); RUDATIS 1791 ( $\mathrm{PRE}=1$ ), $472(\mathrm{STE}=6 \mathrm{a})$, 39 ( $\mathrm{STE}=9)$, 1736 ( $\mathrm{PRE}=10$ ), 1151 (STE=12), 1381 (STE=15), 75 (STE=16b), 1150 (STE=17b), 910 (STE=20); RUMP s.n. (NH 20300=10).
S. SALTMARSHE 984 (GRA, PRE, SAM=14); SCHEEPERS 1371 (PRE=16a); SCHELPE $466(N U=6 b), 920(N H, N U=12), 13(N U=15)$; SCHLECHTER 3348 (GRA, NH, PRE=13), 6634 (GRA=20); SCHONIAND 69 (GRA=4), 3925, $4179(G R A=10), 4144(G R A=15), 704(G R A=19 b) ;$ SCOTT 5
 (PRE 7234=9); SHIRLEY 6. $(\mathrm{NU}=4)$, 76 ( $\mathrm{NU}=6 \mathrm{a}$ ), s.r. ( NU 32355= $6 \mathrm{a}), 241(\mathrm{NU}=10), 5,90(\mathrm{NU}=12)$; SIM $1127(\mathrm{NU}=4)$, 1125 ( $\mathrm{NU}=$ $6 a), 1126(\mathrm{NU}=9), 1120,1124,1855(\mathrm{NU}=10), 1129$ ( $\mathrm{NU}-2$ sheets $\mathrm{SAM}=10), 1117(\mathrm{NU}=12), 1854(\mathrm{NU}=17 \mathrm{a}), 1122(\mathrm{NU}-2$ sheets $=$ 19b), 1123 (NBG, NU=19b), s.n. (NU Oct. 1921=19b); SKEAD 203 ( $\mathrm{NU}=15$ ) ; SMITH 1724 ( $\mathrm{PRE}=10$ ) , 3770 ( $\mathrm{PRE}=20$ ) ; SMOOR 636 (NU= $11), 613(N U=12), 615(N U=17 b), 614(N U=18), 649(N U=21 a) ;$
SMUTS \& GILLETT 2465 (PRE=5), 3165 (PRE, STE=17a); SOLE 393 (GRA $=16 a$ ); SOLOMON 61 (NU=12); STANTON 28 (NU=19a); STAPLES s.r. ( $\mathrm{NH} 18533=1$ ) ; STEPHANSON s.n. (STE 16223=6a) ; STEPIIEN V. GRAAN \& SCHWAKE 1151 (PRE=10); STEWART 10191
(PRE=2), 180 ( $\mathrm{NBG}=7$ ); STEYN 973 ( $\mathrm{NBG}=12$ ), 933 ( $\mathrm{NBG}=16 \mathrm{~b}$ ), 751 (NBG=19a); STIRTON s.n. (NU 21.10.74=1), 1054, 1219 (NU= $6 \mathrm{a}), 1317(\mathrm{NU}=9), 105(\mathrm{NU}=10)$, s.m. (NU $21.10 .74=10), 444$ ( $\mathrm{NU}=12$ ), 513, $1091(\mathrm{NU}=15)$, s.n. $(\mathrm{NU} 21.10 .74=15)$, 1044 ( $\mathrm{NU}=$ 17a), s.n. (NU 21.10.74=17b), 1078 (NU=18) ; STOKDE 1598 (STE= 7); STORY 689 (PRE=6a), 4221 (GRA, PRE=19a); STREY 4521, 5949 ( $\mathrm{NH}=1$ ) , $10117(\mathrm{NH}=4), 9100(\mathrm{NH}=6 \mathrm{a}), 9101$ ( $\mathrm{NH}, \mathrm{PRE}=9), 5163$, $9256(\mathrm{NH}=10), 5942,9962(\mathrm{NH}, \mathrm{NU}=10), 5538(\mathrm{NU}=12), 6182(\mathrm{NH}$, $\mathrm{NU}=12), 6838(\mathrm{NH}, \mathrm{NU}, \mathrm{PRE}=12), 5170(\mathrm{NH}=15), 5986(\mathrm{NH}=16 \mathrm{~b})$, 7261 ( $\mathrm{NH}, \mathrm{PRE}=16 \mathrm{~b}$ ) , 4386, 4772, 5813, 10023, 10066 ( $\mathrm{NH}=17 \mathrm{a}$ ), 4474 ( $\mathrm{NH}, \mathrm{PRE}=17 \mathrm{a}$ ), $5974(\mathrm{NH}, \mathrm{NU}=17 \mathrm{~b}), 10241$ ( $\mathrm{NH}, \mathrm{NU}, \mathrm{PRE}=20$ ), 5990 ( $\mathrm{NH}, \mathrm{NU}=\mathrm{H} \mathrm{l}$ ) , 6879 ( $\mathrm{NH}, \mathrm{PRE}=\mathrm{Hl}$ ), 9149 ( $\mathrm{NH}, \mathrm{NU}, \mathrm{PRE}=\mathrm{Hl}$ ); SYMONS $70(N B G=7), 72(\mathrm{PRE}=9)$.
T. TAYLOR H.C. 588 (NBG=19a); TAYLOR L.E. 5292 (STE=2), 5262 (STE=7), 3535, 4966, 5584 (NBG=10), 5047A (NBG=12), 5376 (NBG $=15), 3634,5946,(N B G=16 a), 5362(N B G=19 a), 3710(N B G=20)$; TAYLOR R.H. 255 ( $\mathrm{NJ}=11$ ); THERON 675, 1191, 2115 ( $\mathrm{NH}=6 \mathrm{a}$ ) , 1190 ( $\mathrm{NH}=10$ ) ; THINGES 1904 ( $\mathrm{PRE}=9$ ) ; THODE 2542, 4662 ( $\mathrm{STE}=1$ ), 2538, 2544 (STE=2), 2539 (STE=4), 2543 (STE=5), 2546, 2547 ( $\mathrm{STE}=7$ ), 2534 ( $\mathrm{STE}=8$ ), A 249 ( $\mathrm{NH}=9$ ), A 2856 ( $\mathrm{NH}=10$ ), 2541,2546 (STE=10), A1646 ( $\mathrm{NH}=12$ ), 2535, 2536 ( $\mathrm{STE}=12$ ), 2533 ( $\mathrm{STE}=15$ ), A703, Al049 ( $\mathrm{NH}=16 \mathrm{a}$ ), A1296 ( $\mathrm{NH}=16 \mathrm{~b}$ ), 2545 ( $\mathrm{STE}=16 \mathrm{~b}$ ), 2532 (STE=17a), 2549 (STE=17b), A2398 (NH=19a), 5293 (STE=21a); THOMAS 23 (SAM=12), 17 (SAM=21a); THOMPSON 2379 (NBG=12); THORNCROFT $634(N H=20)$; TOD $33(N U=10)$; TRACE 59 (NU=1); TRAUSELD 661, 866, 867 ( $\mathrm{NU}=6 \mathrm{a}$ ) , 865 ( $\mathrm{NU}=66$ ) , 868 ( $\mathrm{NU}=9$ ) , 869 ( $\mathrm{NU}=15$ ) , 683 ( $\mathrm{NU}=21 \mathrm{a}$ ); TYSON 1211 (SAM=6a), 1478 ( $\mathrm{CRA}=10$ ); 2745 ( $\mathrm{PRE}=10$ ) , $1125,1210(\mathrm{SAM}=10), 2745$ ( $\mathrm{STE}=10$ ) , 2880 (SAM $=20)$.
V. v. GINKEL 257 ( $\mathrm{PRE}=10$ ) ; v. RENSBURG 25556 (J=3), 25556, 25557 $(J=9)$; v. ROOYEN 5 ( $N U=10$ ); v. d. SCHYFF 4818 (PRE=21a); v.d. ZEYDE s.n. (NBG 9.11.69=2), s.n. (NBG 28.11.72=5), s.n. (NBG 9.11.69=7), s.n. (NBG, 4.2.70, 12.10.70=9), s.n. (NBG 9.11.69=12) .
W. WARD $6654(P R E=1), 1583$ ( $\mathrm{NH}, \mathrm{NU}=6 \mathrm{~b}$ ), 1586 ( $\mathrm{NH}, \mathrm{NU}, \mathrm{PRE}=10$ ), 398, 459 ( $\mathrm{NU}=10$ ) , 1484 ( $\mathrm{NH}, \mathrm{NU}=12$ ), 4284 ( $\mathrm{NH}, \mathrm{NU}, \mathrm{PRE}=12$ ), 1148 ( $\mathrm{NH}=15$ ), 1655 ( $\mathrm{NH}, \mathrm{NU}=15$ ), 971 ( $\mathrm{NU}=17 \mathrm{a}$ ), 4131 ( $\mathrm{NH}=17 \mathrm{a}$ ), 5208 ( $\mathrm{NH}=17 \mathrm{~b}$ ) ; WEBB 26 ( $\mathrm{NU}=15$ ); WEEKS 25 ( $\mathrm{J}=9$ ); WEYDERMANN \& OBERDIECK 1147 (PRE=6a), 1440 (PRE=15); WEISS s.n. (STE 16907=7); WELLS 1578 ( $\mathrm{NU}=10$ ), 2113 ( $\mathrm{NH}=10$ ); WEST 1404 ( $\mathrm{NH}=$ 21a); WESTHUIZEN 820 (PRE=6b); WLLKER 116 (NU=10); WILSENACH s.n. (J 35565=6b), s.n. (PRE 33559=17a); WINTER 617 (NBG=9); WOOD S.E. 73, $85,111,161$ ( $\mathrm{NU}=1$ ), $104,117,153,159$ ( $\mathrm{NU}=2$ ), 91, 97, 131 ( $\mathrm{NU}=4$ ), 155,158 ( $\mathrm{NU}=5$ ), $92,94,95,106,107,108$, $114,124,128,128 \mathrm{~A}, 140,143,152,161 \mathrm{~A}, 164 \mathrm{~A}, 175$ ( $\mathrm{NU}=6 \mathrm{a}$ ), 92A, 121, 132, 149, 154 ( $\mathrm{NU}=7$ ), 210 ( $\mathrm{NU}=8$ ), $72,75,76,77,79$, $82,86,100,102,106,118,145,165 \mathrm{~B}, 167,168,170,171,173$, 174 ( $\mathrm{NU}=1 \mathrm{C}$ ), $101,122,126,151,160$, 163 ( $\mathrm{NU}=12$ ), 130 ( $\mathrm{NU}=13$ ), $74,75,77 \mathrm{~A}, 78,80,81,83,90,98,103,103 \mathrm{~A}, 115,120,137$, $142,149 \mathrm{~A}, 156,164,209(\mathrm{NU}=15), 71 \mathrm{~A}, 80 \mathrm{~A}, 86 \mathrm{~A}, 133,179(\mathrm{NU}=$ $17 \mathrm{a}), 146,180(\mathrm{NJ}=17 \mathrm{~b}), 121 \mathrm{~A}, 125,138,139,141,144,145 \mathrm{~A}$, 150 ( $\mathrm{NU}=18$ ); WRIGHT 1549 ( $\mathrm{NU}=3$ ), 1566,1602 ( $\mathrm{NU}=12$ ), 24,1552 ( $\mathrm{NJ}=15$ ), 1567, 1600 ( $\mathrm{NU}=\mathrm{H} 2$ ); WURTS 467 ( $\mathrm{NBG}=19 \mathrm{a}$ ); WYLIE s.n. (PRE, NH 27973=1), (com. Medley Wood) 10606 ( $\mathrm{NH}=4$ ), s.n. ( NH 21681, 23357, 28016=6a), s.n. ( $\mathrm{NH} 21682,27977=7$ ), s.n. ( NH 22420, 23107=10), s.n. (NH 21683=12), s.n. (NH 28020, PRE Nov, 1936=15), s.n. (NH $23112=17 a$ ), s.r. (NH $27962=17 \mathrm{~b}$ ), s.n. (PRE Oct.-Nov. 1931=18).
2. ZEYHER 1670 ( $\operatorname{STE}=6 \mathrm{a}$ ), $6(\mathrm{STE}=16 \mathrm{a})$, 950 ( $\mathrm{SAM}, \mathrm{STE}=16 \mathrm{a}$ ), s.n. (STE 30404=16b).

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[^0]:    a - H. membranacea, b-H. angustifolia var. buchananii, $c$ - putative hybrid

